

02.1-42 THE STRUCTURE OF HUMAN LACTOFERRIN AT 3.2Å RESOLUTION

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Lactoferrin, a member of the transferrin family of proteins is an iron binding glycoprotein ($M_r \sim 80\ 000$ daltons), present in mammalian milks, mucous secretions and white blood cells. Because of its ability to bind iron tightly, ($K \sim 10^{20}$) lactoferrin exerts a bacteriostatic effect *in vitro* by depriving micro-organisms of essential iron (Bullen, J.J., Rogers, H.J. & Leigh, L., Brit. Med. J., 1972, 3, 69-75). As well as a role in non-immunological host defence, there are reports which suggest it may modulate immune and inflammatory processes (Birgens, H.S., Scand. J. Haematol., 1984, 33, 225-230) and prevent the catalytic formation of potentially harmful oxygen radicals (Baldwin, D.A., Jenny, E.R. & Aisen, P., J. Biol. Chem., 1984, 259, 13391-13394). The three dimensional structure of lactoferrin determined at 3.2Å resolution will be presented. The 703 amino acid residues are organised into two homologous lobes that are connected by a short α helix. Each of these lobes carries one iron binding site, one carbohydrate chain, and is divided into two domains of supersecondary structure. The iron is found at the domain interface where it is bonded to four protein ligands, 2 tyrosine, 1 histidine and 1 aspartate. The fifth and sixth coordination sites correspond with a region of positive electron density adjoining the iron and provide a possible location for the associated anion (CO_3^{2-} or HCO_3^-). This region is adjacent to an arginine sidechain and a helix N-terminus. Some remarkable structural similarities between lactoferrin and certain other binding proteins are also apparent.

02.1-43 THE CRYSTAL STRUCTURE OF 4-ZN RHOMBOHEDRAL DES-B30 CROSS-LINKED HUMAN INSULIN. By U. Derewenda, Z. Derewenda, G. G. Dodson, Dept. of Chem., Univ. of York, U.K. & J. Markusson, NOVO Research Institute, Denmark.

Human des-B30 insulin in which there is a peptide bond formed between Lys B29 and Gly A1 by the synthetic action of trypsin is a byproduct of the industrial conversion of pig to human insulin. This modification leaves the hormone without detectable activity. Rhombohedral crystals of this insulin have been grown in conditions that produce 4 Zn insulin with native hormone. The crystals' cell dimensions and diffraction pattern showed they were similar to native rhombohedral 4Zn insulin. Refinement of the structure proceeded smoothly from the 4Zn insulin coordinates and converged with the agreement factor $R = .175$. The main structural change in this cross-linked insulin is the movement of B27 - B29 into an approximate helix which is continuous with the helix at A1 - A9. There are two zinc ions on the 3-fold axis coordinated by the B10 His, in contrast to the off-axial coordination seen in native 4Zn insulin. The overall structure of the des-B30 insulin molecule is, apart from the cross-bridge, essentially the same as the native molecule. This structure of des-B30 cross-linked insulin is therefore similar to the beef diamino suberic acid B29 - A1 cross-linked insulin. The latter modification leaves some flexibility in the molecule but is associated with a large drop in potency (to ca. 15%). We conclude that the explanation for the absence of activity in the des-B30 is the more complete loss of flexibility at the B chain C terminus.

02.1-44 A NEW CRYSTAL FORM OF INSULIN. Z.H. Rao and N.W. Isaacs. St. Vincent's Institute of Medical Research, Victoria Parade, Melbourne, Victoria 3065, Australia.

A new crystal form of pig insulin has been obtained. The crystals are orthorhombic, with space group C222₁, and cell dimensions $a = 60.2\text{Å}$, $b = 228\text{Å}$, $c = 222\text{Å}$.

Matthews (J. Mol. Biol., (1968) 33, 491) has found that for protein crystals the value of the crystal volume per unit mass, V_m , lies within the range 1.8 to 3.6 with a median value of 2.4. For a V_m of 1.8 the asymmetric unit of these crystals would contain 36 insulin molecules and for a V_m of 3.6 there would be 18 molecules. Assuming the insulin exists as a hexamer, for a V_m of 2.7, which is close to the median of observed values, there would be 24 molecules or 4 hexamers in the asymmetric unit.

A model for the structure, based on the packing of nearly spherical hexamers subject to the constraints of the space group symmetry, has been obtained.

The crystals diffract to a resolution of about 3.0Å on precession photographs. The same crystals can be obtained from bovine insulin.

02.1-45 CRYSTAL STRUCTURES OF BOWMAN-BIRK PROTEASE INHIBITOR AND ITS COMPLEX WITH TRYPSIN. by Y. Tsunogae, A. Suzuki, I. Tanaka, T. Yamane and T. Ashida, Faculty of Engineering, Nagoya University, Chikusa-ku, Nagoya 464, Japan

Bowman-Birk inhibitors found in the seeds (beans) of the leguminous plants are small proteins which inhibit the serine proteases by making stable enzyme-inhibitor complexes. The inhibitors usually consist of 60-80 amino acid residues including 7 disulfide linkages, all are evolutionarily conserved. They are double-headed inhibitors consisting of two tandem homologous domains each with a binding site. Each domain consists of three peptide loops made by disulfide linkages, and each loop is made of 8-11 residues. Crystals have been obtained of AB-I and IIa from azuki beans, A-I, II and B-III from peanuts, and their complexes with trypsin or chymotrypsin.

A-II, 3 Å study: The molecule has an elongated shape with an approximate dimension of $45 \times 15 \times 15$ Å, consisting of two distinct domains which are connected by two rather flexible chains (Fig. 1.). The structures of the domains are very similar to each other and are related by an intramolecular pseudo two-fold symmetry. The binding sites are in the outermost loops, which protrude from the core of the molecule to the opposite direction. The electron densities for both binding sites are very low, indicating a considerable flexibility or a disorder in the conformations.

AB-I+trypsin complex, 2.3 Å study: The SIR method with the MR method solved the structure. Of the inhibitor only the structure of the trypsin-binding domain could be determined. The electron density for the chymotrypsin-binding domain, however, is so low that any model could not be built. The structure was refined to $R=0.21$ including trypsin, the trypsin-binding domain of 29