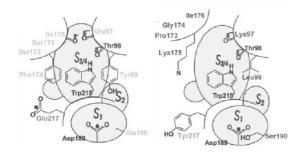
s1.m8.o3 Understanding Protein - Ligand Interactions For Drug Design: A Structural Perspective. <u>Milton T.</u> <u>Stubbs</u>^a, Daniel Rauh^a and Gerhard Klebe^b, ^aInstitut für Biotechnologie, Martin-Luther Universität Halle-Wittenberg, Germany; ^bInstitut für Pharmazeutische Chemie, Philipps-Universität Marburg, Germany. E mail: stubbs@biochemtech.uni-halle.de

Keywords: Protein-Ligand Recognition; Drug Design; Serine-Proteinases

A high resolution crystallographic structure determination of a protein - ligand complex is generally accepted as the 'gold standard' for structure-based drug design. Yet how much does a crystal structure actually reveal about ligand affinity? In a combined crystallographic, kinetic and mutagenic approach, we have set out to analyse the multivariate determinants for ligand selectivity.

In so doing, we have observed pH-dependent changes in binding mode, as well as ligand-induced large scale reorganisation of secondary structure. The availability of multiple crystal forms allows us to factorise the relative contributions of competing processes to protein - ligand affinity. The implications for



computational drug design will be discussed.

Fig.1 Schematic diagram showing the ligand binding sites of factor Xa (left) and trypsin (right). The aromatic pocket of factor Xa was introduced into that of trypsin through mutation of the residues shown.

- Renatus, M., Bode, W., Huber, R., Stürzebecher, J. and Stubbs, M.T. (1998) Structural and functional analyses of benzamidinebased inhibitors in complex with trypsin: Implications for the inhibition of factor Xa, tPA and urokinase. J. Med. Chem. 41, 5445-5456.
- [2] Dullweber, F., Stubbs, M.T., Musil, D., Stürzebecher, J. and Klebe, G. (2001) Factorising ligand affinity: A combined thermodynamic and crystallographic study of trypsin and thrombin inhibition. J. Mol. Biol. 313, 593-614.
- [3] Stubbs, M.T., Reyda, S., Dullweber, F., Möller, M., Klebe, G., Dorsch, D., Mederski, W.W.K.R. and Wurziger, H. (2002) pH-dependent binding modes observed in trypsin crystals – Lessons for structure-based drug design. Chem. Bio. Chem. 3, 246-249.
- [4] Rauh, D., Reyda, S., Klebe, G. and Stubbs, M.T. (2002) Trypsin mutants for structure based drug design: expression, refolding and crystallisation. Biol. Chem. 383, 1309-1314.
- [5] Reyda, S., Sohn, C., Klebe, G., Rall, K., Ullmann, D., Jakubke, H.-D. and Stubbs, M.T. (2003) *Reconstructing the binding site of factor Xa in trypsin reveals ligand-induced structural plasticity.* J. Mol. Biol. 325, 963-977.
- [6] Rauh, D., Klebe, G., Stürzebecher and Stubbs, M.T. (2003) ZZ made EZ: Influence of inhibitor configuration on enzyme selectivity. J. Mol. Biol. 330, 761-770.
- [7] Rauh, D., Klebe, G. and Stubbs, M.T. (2004) Understanding protein-ligand interactions: The price of flexibility. J. Mol. Biol. 335, 1325-1341.

www.biochemtech.uni-halle.de/xray

s1.m8.o4 Structural Basis of Diverse Substrate Recognition by PMM/PGM from P. Aeruginosa. Catherine Regni, Peter A. Tipton, Lesa J. Beamer, University of Missouri -Columbia, Biochemistry, USA. E-mail: car963@mizzou.edu

Keywords: Enzyme; Protein-Ligand Interactions; P. Aeruginosa

P. aeruginosa is an opportunistic human pathogen that produces life-threatening infections in cystic fibrosis patients, burn victims, and immunocompromised individuals. The bacterium is known to form antibiotic resistant biofilms, and its infections are further complicated by the production of exoproducts that contribute to its virulence. The enzyme phosphomannomutase/phosphoglucomutase (PMM/PGM) participates in the biosynthesis of three virulence factors from P. aeruginosa: alginate, rhamnolipid and lipopolysaccharide. Enzyme-substrate complexes of PMM/PGM reveal the structural basis of the enzyme's ability to use four different substrates in catalysis. High-resolution structures with glucose 1-phosphate, glucose 6-phosphate, mannose 1-phosphate, and mannose 6-phosphate show that the position of the phosphate group of each substrate is held constant by a conserved network of hydrogen bonds. This produces two distinct, and mutually exclusive, binding orientations for the sugar rings of the 1-phospho and 6-phospho sugars. Specific binding of both orientations is accomplished by key contacts with the O3 and O4 hydroxyls of the sugar, which must occupy equatorial positions. Dual recognition of glucose and mannose phosphosugars uses a combination of specific protein contacts and nonspecific solvent contacts. The ability of PMM/PGM to accommodate these four diverse substrates in a single active site is consistent with its highly reversible phosphoryl transfer reaction and allows it to function in multiple biosynthetic pathways in P. aeruginosa.