

**MS.35.3***Acta Cryst.* (2008). A64, C67**Apparent mismatch between XAFS and XRD structure of crystalline and amorphous electrochromic WO<sub>3</sub>**Alain Michalowicz<sup>1</sup>, Jacques Moscovici<sup>1</sup>, Aline Rougier<sup>2</sup>, Stephane Laruelle<sup>2</sup><sup>1</sup>Institut de Chimie et des Materiaux Paris Est, Universite Paris 12 and CNRS, 2 rue Henri Dunant, Thiais, France, 94320, France, <sup>2</sup>Laboratoire de Reactivite et Chimie des Solides, Universite de Picardie Jules Vernes, 33 rue Saint Leu, 80039 Amiens Cedex, France, E-mail : michalov@univ-paris12.fr

WO<sub>3</sub> are used in a great variety of inorganic materials dealing with a wide range of applications. We have studied four different crystalline WO<sub>3</sub> materials (monoclinic (1), monohydrate (2), hexagonal (3) and pyrochlore (4) ) as precursors to amorphous electrochromic species obtained by mechanosynthesis. The structures of the crystalline phases were determined by various diffraction techniques (single crystal, X-ray or Neutron powder diffraction). EXAFS was used to compare the local structure around W of these crystalline phases and those of the corresponding amorphous materials. The EXAFS W-O radial distribution functions were determined as sums of gaussians, in the frame of the errors and quality of fits estimation procedures recommended by the International XAFS Society (IXS) standard and criteria committee (2000). From this comparison, two main results are obtained :

The XAFS and diffraction rdf of (1) and (2) are in agreement, but the powder diffraction rdf of (3) and (4), which are metastable phases, are in total disagreement with the EXAFS results. In these two last cases, diffraction structures miss the existence of short and long W-O bonds, revealed by the EXAFS study. We explain this mismatch by disorder effects which lead to averaged crystal structures. On the contrary the EXAFS study is able to give a more precise picture of the local W structure, even if the long range order informations are poorer; The W-O rdf of the corresponding amorphous materials converge to a unique local structure, independent of its crystalline precursor.

Keywords: inorganic materials, XAFS and XRD, quality of fit in EXAFS

**MS.35.4***Acta Cryst.* (2008). A64, C67**High pressure and high temperature EXAFS and diffraction study of AgI**Akira Yoshiasa<sup>1</sup>, Hiroshi Arima<sup>2</sup>, Hiroshi Fukui<sup>3</sup>, Maki Okube<sup>4</sup>, Yoshinori Katayama<sup>3</sup>, Osamu Ohtaka<sup>2</sup><sup>1</sup>Kumamoto University, Graduate School of Science, Kurokami 2-36-1, Kumamoto, Kumamoto, 860-8555, Japan, <sup>2</sup>Osaka University, Toyonaka 560-0043, Japan, <sup>3</sup>JAERI, Spring-8, Hyogo 679-5198, Japan, <sup>4</sup>Tokyo Institute of Technology, Yokohama 226-850 Japan, E-mail : yoshiasa@sci.kumamoto-u.ac.jp

We have determined the precise P-T phase diagram of AgI by *in situ* high-pressure high-temperature synchrotron experiments [1]. We will report on structural details and effective potentials in various high pressure phases. EXAFS and X-ray diffraction measurements were performed up to 6.0 GPa and 1100 K using a multi-anvil high-pressure device and synchrotron radiation from Spring-8, Hyogo. In the disordered rock-salt phase, Ag ions occupy both octahedral and tetrahedral sites and twenty percent of Ag ions occupy the tetrahedral site as a maximum value at 2GPa. The transition between the rock-salt type and disordered rock-salt type phases is a broad disorder type within the same structure. From the viewpoint of the local

structure analyses, some sudden changes are recognized near the phase transition point. The Debye Waller factors in AgI phases were investigated by both the diffraction and EXAFS methods. Analysis of EXAFS Debye-Waller factor is useful because the force constant can be decided directly even at high pressure and high temperature [2]. Pressure influences greatly the effective pair potential and anharmonicity decreases with increasing pressure. Phonon dispersion relations in various phases have been derived from high pressure experiments.

[1] O.Ohtaka, et al. (2002) *Solid State Comm.*, 123, 213-216.[2] A.Yoshiasa et al. (2000) *Jpn. J. Appl. Phys.* 39, 6747-6751.

Keywords: high pressure, superionic conductor, Debye-Waller factor

**MS.35.5***Acta Cryst.* (2008). A64, C67***In situ* XRD and XAFS studies of oxidation/reduction and water gas shift reactions of Cu doped ceria**Jonathan C. Hanson<sup>1</sup>, Jose A Rodriguez<sup>1</sup>, Xainqin Wang<sup>1</sup>, Wen Wen<sup>1</sup>, Zhong Zhong<sup>1</sup>, Anatoly Frenkel<sup>2</sup>, Qi Wang<sup>2</sup>, Peter Chupas<sup>3</sup><sup>1</sup>Brookhaven National Laboratory, Chemistry, PO Box 5000, Upton, NY, 11973, USA, <sup>2</sup>Physics Dept., Yeshiva Univ., New York, NY 10016. USA, <sup>3</sup>Advanced Photon Source, Argonne National Laboratory, Argonne, IL 60439 USA, E-mail : hanson1@bnl.gov

Metal doped and impregnated ceria has been shown to be an active catalyst for the water gas shift (WGS) reaction. The structural transformations under REDOX and WGS conditions have been shown to involve reversible movement of the Cu out and back into the cerium atomic positions in the ceria structure. The diffraction studies were carried out with high energy X-rays and the high Q data allowed for improved profile refinement [1, 2] and pair distribution function (PDF) studies [3] during structural transformations (typical sampling time 3 minutes). The XAFS studies were collected in Quick EXAFS mode [4] during transformations (typical sampling times 15 seconds). Intermediate Cu<sub>2</sub>O like structure was demonstrated from principal component analysis of the time resolved QXANES data and subsequent modeling of the QEXAFS data. A comparison of PDF and EXAFS results will be presented. The research carried out at BNL was financed through contracts DE-AC02-98CH10886, DE-FG02-03ER15476 and DE-FG02-05ER15688 with the US Dept. of Energy (Division of Chemical Sciences).

1. Wang, X., et al. *J. Phys. Chem B*, 2006. 110: p. 428-434.2. Wang, X.Q., et al. *Journal of Chemical Physics*, 2005. 122(15): p. 154711-1 154711-10.3. Chupas, P.J., et al. *J. Appl. Crystallography*, 2003. 36: p. 1342-1347.4. Caliebe, W.A., et al. *Rad. Phys. and Chem.*, 2006. 75: p. 1962-1965.

Keywords: *in situ* diffraction, catalyst structure, X-ray absorption

**MS.36.1***Acta Cryst.* (2008). A64, C67-68**Zooming into the overall architecture of the giant muscle protein titin**Matthias Wilmanns<sup>1</sup>, Christina Vega<sup>1,2</sup>, Simone Muller<sup>1,3</sup>, Qing Chen<sup>1</sup>, Young-Hwa Song<sup>1</sup>, Nikos Pinotsis<sup>1,4</sup><sup>1</sup>EMBL, EMBL-Hamburg, Notkestrasse 85, Hamburg, Hamburg, 22603,

## Microsymposia

Germany, <sup>2</sup>IBMB - CSIC, Parc Científic de Barcelona, Josep Samitier 1-5, 08028 Barcelona, Spain, <sup>3</sup>Cardiovascular Research Institute, California Institute for Quantitative Biomedical Research, University of California, San Francisco, CA, USA, <sup>4</sup>ICR, 237 Chester Beatty Laboratories, Fulham Road, London SW3 6JB, UK, E-mail: wilmanns@embl-hamburg.de

The giant muscle protein titin extends over one half of the muscle sarcomere. In its largest isoform titin comprises more than 38,000 residues and about 300 domains. Its structural complexity does not allow the application of classical structural biology methods to determine its overall architecture. Therefore, we have decided to chop the protein into smaller fragments and determine the high resolution structures of representative parts. Within this endeavor, we have also become interested to consider known ligands involved in the titin interactome for structural/functional analysis. Over the last decade, we determined structures of the N-terminal assembly complex (Zou et al., 2006), from the I-band (Mayans et al., 2001; Vega et al., unpublished) and from the A-band including the kinase domain and down-stream signaling complexes (Mayans et al., 1998; Muller et al., 2006; Muller et al., 2007; Muller et al., unpublished; Chen et al., unpublished). The available data allow modeling a large part of the titin proteome and to interpret available low resolution data of the entire titin filament. Combined with complementary functional data, our findings reveal key structural/functional relationships of titin and its interactions partners. Structural biology results from a related sarcomeric filament protein, myomesin (also known as mini-titin) will be presented in a separate contribution. Some of the data have been published recently (Pinotsis et al., 2008).

References:

- Mayans et al. (1998) *Nature* 395, 863-869.  
Mayans et al. (2001) *Structure* 9, 331-340.  
Muller et al. (2007). *J Mol Biol.* 371, 469-480.  
Muller et al. (2006) *FEBS Lett.* 580, 341-444.  
Pinotsis et al. (2008) *EMBO J.* 27, 253-264.  
Zou P, Pinotsis, N et al. (2006) *Nature* 439, 229-33.

Keywords: muscle proteins, protein/protein interactions, kinase structure

### MS.36.2

*Acta Cryst.* (2008). A64, C68

#### **Ion channel structures by single particle analysis using EM: Sodium and TRP channels, IP3 receptor**

Chikara Sato, Kazuhiro Mio, Yuusuke Maruyama, Toshihiko Ogura  
National Institute of Advanced Industrial Science and Technology (AIST), Neuroscience Research Institute and JBIRC, Umezono 1-1-4, Tsukuba, Ibaraki, 305-8568, Japan, E-mail: ti-sato@aist.go.jp

Six-transmembrane (6-TM) type channels are plasmamembrane integral components of cellular signaling pathways conserved in almost all species including animals, plants, and some kinds of prokaryotes. These channels selectively permeate cations in response to various signals. In excitable and non-excitable mammalian cells, 6-TM cation channels play fundamental roles, including the generation of action potential and its transmission, the regulation of intracellular ion concentrations, and the activation of signaling cascades by humoral or mechanical pathways. We have recently determined the structure of four different 6-TM type cation channels: the voltage-sensitive sodium channel<sup>1</sup>, the IP3 receptor<sup>2</sup>, the TRPC<sup>3</sup> and TRPM<sup>2</sup><sup>4</sup> channels, using single particle analysis from cryo-EM images. The basic structure of the molecules was shown to be similar: a bell-like shape composed of a relatively small extracellular (or luminal) domain, a protein-dense transmembrane domain, and an expanded cytoplasmic domain. However in detail, the cytoplasmic

architectures are quite different from each other and are diversely evolved to their specific physiological functions.

1. Sato, C., Ueno, Y., Asai, K., Takahashi, K., Sato, M., Engel, A., and Fujiyoshi, Y. *Nature* 409, 1047-1051, (2001).
2. Sato, C., Hamada, K., Ogura, T., Miyazawa, A., Iwasaki, K., Hiroaki, Y., Tani, K., Terauchi, A., Fujiyoshi, Y. & Mikoshiba, K. *J. Mol. Biol.* 336, 155-164, (2004).
3. Mio, K., Ogura, T., Kiyonaka, S., Hiroaki, Y., Tanimura, Y., Fujiyoshi, Y., Mori, Y. & Sato, C., *J. Mol. Biol.* 367, 373-383 (2007).
4. Maruyama, Y., Ogura, T., Mio, K., Kiyonaka, S., Kato, K., Mori, Y. & Sato, C. *J. Biol. Chem.* 282, 36961-36970, (2007).

Keywords: electron microscopy analysis, ion channel structures, three-dimensional, image reconstruction

### MS.36.3

*Acta Cryst.* (2008). A64, C68

#### **The assembly process of the double-layered capsids of phytoeoviruses**

Toshihiro Omura<sup>1</sup>, Naoyuki Miyazaki<sup>2</sup>, Hisashi Naitow<sup>3</sup>, Taiyun Wei<sup>1</sup>, Takumi Shimizu<sup>1</sup>, R. Holland Cheng<sup>4</sup>, Atsushi Nakagawa<sup>2</sup>, Tomitake Tsukihara<sup>2</sup>

<sup>1</sup>National Agricultural Research Center, 3-1-1 Kannondai, Tsukuba, Ibaraki, 305-8666, Japan, <sup>2</sup>Institute for Protein Research, Osaka University, <sup>3</sup>RIKEN Harima Institute, Spring-8 Center, <sup>4</sup>Department of Molecular and Cellular Biology, University of California, E-mail: toomura@affrc.go.jp

Viruses in the family *Reoviridae* have an inner core with a large interior cavity that contains the 10 to 12 segmented double-stranded RNA as a genome, and a transcriptional complex that includes proteins with RNA polymerase, helicase, guanylyltransferase and transmethylese activities and an RNA-binding protein. The core particle is surrounded by one or two layers of outer capsid proteins. During X-ray crystallographic and Cryo-electron microscopic studies of the structural organization of *Rice dwarf virus*, a member of the genus *Phytoreovirus* in the family *Reoviridae*, we have identified possible structural mechanisms that allow creation of a large cavity inside a double-layered spherical particle that consists of heterologous proteins with different lattices. The viral particle seems to be created in a genetically economical manner, with the sealing of joints between inner-layer proteins by a second layer of proteins, suggesting the organization of a rigid protein layer that separate from and, probably, protects the interior of the virus from the cytoplasmic environment within infected cells. Procedure of the virus assembly was analyzed combined with molecular cytopathological data in virus infected cells.

Keywords: virus assembly, viral structure and function, virus host interactions

### MS.36.4

*Acta Cryst.* (2008). A64, C68-69

#### **A new virus structure: The nucleosome-like organization of the filamentous archaeal virus AFV1**

Adeline Goulet<sup>1</sup>, Victor Kostyuchenko<sup>2</sup>, Nicolas Leulliot<sup>3</sup>, Herman van Tilbeurgh<sup>3</sup>, David Prangishvili<sup>4</sup>, Christian Cambillau<sup>1</sup>, Michael G Rossmann<sup>2</sup>, Valerie Campanacci<sup>1</sup>

<sup>1</sup>CNRS, Université Aix-Marseille I & II, AFMB, case 932, 163 avenue de Luminy, Marseille cedex 09, PACA, 13288, France, <sup>2</sup>Departement of biological sciences, Purdue university, 915 West state street, West