

edge (XANES) and extended (XAFS) at the Zn K-edge have been explored. We will present further insights into the divalent metal ions regulation of the hut operon in *Bacillus subtilis*, by combining both crystallography and X-ray absorption spectroscopic studies.

1) Kumarevel et al., *Nature* 434, 183-191 (2005), 2) Kumarevel et al., *Nucleic Acids Res.* 33, 5494-5502 (2005)

Keywords: X-ray absorption fine structure, protein crystallography with synchrotron radiation, protein refinement

MS.63.5

Acta Cryst. (2008). A64, C111

The structure of the Amyloid β -peptide high affinity copper II binding site in Alzheimer's disease

Victor A. Streltsov, Stephen J. Titmuss, Vidana Epa, Jose N. Varghese

CSIRO, Molecular and Health Technologies, 343 Royal Parade, Parkville, Victoria, 3052, Australia, E-mail: victor.streltsov@csiro.au

A major source of neurodegeneration observed in Alzheimer's disease (AD) is believed to be related to the toxicity from reactive oxygen species (ROS) produced in the brain by the Amyloid- β ($A\beta$) protein bound primarily to copper ions. The evidence for an oxidative stress role of $A\beta$ -Cu redox chemistry is still incomplete. Details of the copper binding site in $A\beta$ may be critical to the etiology of AD. Here we present the structure determined by combining X-ray absorption fine structure (XAFS) and Density Functional Theory analysis of truncated $A\beta$ (1-16) peptide complexed with Cu(II) in solution under a range of buffer conditions. PBS buffer salt (NaCl) concentration does not affect the copper binding mode. The XAFS spectra for truncated $A\beta$ (1-16)-Cu(II) and full length $A\beta$ (1-40/42)-Cu(II) peptides are similar. The novel six-coordinated (3N3O) geometry around copper in the $A\beta$ -Cu(II) complex includes three histidines, glutamic or/and aspartic acid and axial water. The structure of high affinity Cu²⁺ binding site is consistent with the hypothesis that the redox activity of the metal ion bound to $A\beta$ can lead to the formation of di-tyrosine linked dimers found in AD. X-ray absorption near-edge spectroscopy (XANES) has been used to probe the substrate mediated reduction of Cu(II) to Cu(I) in $A\beta$ -Cu(II) complexes by ascorbate and the neurotoxin 6-hydroxydopamine (6-OHDA), however dopamine and, in particular, cholesterol are incapable of reducing soluble monomeric $A\beta$ -Cu(II) complexes. The results are in agreement with assignment of the redox potentials for $A\beta$ -Cu(II), ascorbic acid and dopamine.

Keywords: beta-amyloids, Alzheimer's proteins, X-ray absorption

MS.64.1

Acta Cryst. (2008). A64, C111

Structural basis of a plant photosystem I sunlight conversion

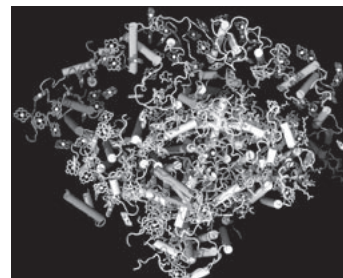
Alexey Amunts, Omri Drory, Nathan Nelson

Tel Aviv University, Biochemistry, Levanon st., Tel Aviv, Tel Aviv, 69978, Israel, E-mail: amunts@tauex.tau.ac.il

A plant Photosystem I (PSI) is a large membrane super-complex that drives photosynthesis. PSI captures sunlight through sophisticated pigment network and uses the energy to perform transmembrane electron transfer. It consists of the reaction center

complex (RC), where the charge separation reaction takes place and the light harvesting complex (LHCI), which serves as an additional antenna system. PSI performs a photochemical activity with the unprecedented quantum yield of close to 1.0, being the most efficient light capturing and energy conversion device. We determined the X-ray structure of intact PSI at 3.4 Å resolution [1]. The crystal structure provides a picture at near atomic detail of 17 protein subunits; 3038 amino acids were assigned, as well as 168 chlorophylls, 2 phyloquinones, 3 Fe₄S₄ clusters and 5 carotenoids. The remarkable feature of PSI is the unprecedented high content of non-protein components, approximately one third of the total mass of about 650 KDa consists of different co-factors. The structure reveals intriguing insights regarding unique interactions between the RC and the LHCI complexes.

[1] Amunts, A., Drory, O. & Nelson, N. (2007) *Nature*, 447, 58-63.



MS

Keywords: photosynthesis, membrane protein, electron transfer

MS.64.2

Acta Cryst. (2008). A64, C111

Inhibitor complexed structures of the Cyt bc1 from the photosynthetic bacterium *R. sphaeroides*

Di Xia¹, Lothar Esser¹, Chang-An Yu²

¹Center for Cancer Research, National Cancer Institute, NIH, Laboratory of Cell Biology, 37 Convent Dr., Building 37, Room 2122C, Bethesda, Maryland, 20892, USA, ²Department of Biochemistry and Molecular Biology, Oklahoma State University, Stillwater, OK, 74078, USA, E-mail: dixia@helix.nih.gov

The cytochrome bc₁ complex (bc₁) is a major contributor to the proton motive force across the membrane by coupling electron transfer to proton translocation. The crystal structures of wild type and mutant bc₁ complexes from the photosynthetic purple bacterium *Rhodospirillum rubrum* (*Rsbcb1*), stabilized with the quinol oxidation (QP) site inhibitor stigmatellin alone or in combination of with the quinone reduction (QN) site inhibitor antimycin, were determined. The high quality electron density permitted assignments of a new metal-binding site to the cytochrome c₁ subunit and a number of lipid and detergent molecules. Structural differences between *Rsbcb1* and its mitochondrial counterparts are mostly extra membranous and provide a basis for understanding the function of the predominantly longer sequences in the bacterial subunits. Functional implications for the bc₁ complex are derived from analyses of 10 independent molecules in various crystal forms and from comparisons with mitochondrial complexes.

Keywords: membrane protein crystallization, cytochrome bc₁ complex, mechanism of proton pumping

MS.64.3

Acta Cryst. (2008). A64, C111-112

Structure and mechanism of the DsbB-DsbA protein disulfide generation system in *E. coli*

Kenji Inaba