and limitations of WAXS data will thus enhance both our ability to analyze WAXS experiments and their use as experimental tests of computational models of molecular systems.

Keywords: WAXS characterization, X-ray solution scattering, SAXS

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“*Irena*” software package for analysis and modeling of small-angle scattering data

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Availability of high-quality small-angle scattering (SAS) data is increasing with the proliferation of a wide range of high-performance user-friendly synchrotron and desktop small-angle X-ray scattering (SAXS) instruments. Similarly, neutron sources also provide an increasing number of quality and well-supported small-angle neutron scattering (SANS) instruments. As the barrier to quality data is decreasing, non-expert users - highly skilled experts in their particular fields with only limited small-angle scattering expertise - are applying the SAS techniques in their science. These users often need extensive help to be able to correctly apply the small-angle scattering methods and generate high-level science in their areas of expertise. The software package “*Irena*” was designed to be a comprehensive package with tools for most of the steps a user commonly needs to perform, balancing ease of use for non-expert users with the controls and complexity expert users need. “*Irena*” provides a number of data modeling and analysis tools, mostly applicable in such fields as materials science, polymers, and chemistry. A wide range of support tools are also included such as ones to enable input, output, manipulations with, and graphing of the data, as well as a scattering contrast calculator. As an example: one of the tools in *Irena* enables modeling of multiple non-interacting dilute systems with a choice of form factors and, when necessary, including one of five available structure factors. This model can be fitted to one of five available structure factors. This connection between them can be presented as radial density functional $p(x,y,z)$ which describes the electron density map corresponding to peptides and its relation to some distance functional $r(x,y,z)$. The method is rotation invariant and its combination with the existing technique that uses the template matching improves the completeness of the protein structure provided by ARP/wARP.

Keywords: protein structure determination, electron density, pattern recognition

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Density modification by directed evolution of electron density maps

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There are numerous cases in macromolecular X-ray crystallography where current methods of density modification are not capable to yield an interpretable map. Since the performance of the automatic procedures for model building of the corresponding protein (e.g. RESOLVE, TEXTAL or ARP/wARP) strongly depends on the quality of the phases, reliable methods for modifying the density are absolutely essential. We are developing a new iterative density modification method which is based on directed evolution of the electron density map. One of the central aims for the new method is to be applicable to cases when the resolution of the data is lower than 3 Å. In a first step each grid point of the initially phased map is analysed by statistical and pattern recognition methods to define a likelihood whether the density value of the point should be changed. A small number of map points are subsequently ‘mutated’. Finally, the phases from the mutated map are used to generate a new density distribution. First encouraging results with 3 Å resolution test cases have been obtained, where this iterative procedure lead to a reduction of the phase error of about 0.2° for each iterative step of the procedure. These will be presented and discussed. The application of the method to the crystallographic use of low resolution images from electron microscopy studies will also be presented.

Keywords: density modification, pattern recognition, protein crystallography

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ARP/wARP: From noisy electron densities of proteins to complete structures

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Structural interpretation of the provided experimental X-ray data, which have a low level of the information content, that therefore leads to a noisy and troublesome interpretable electron density map is a challenging task in the protein crystallography. An important carrier of the structural information in electron density maps of proteins is the repetitive chemical motif of the peptide unit that links the amino acids. One efficient way for the peptide recognition is implemented in the chain tracing module of the ARP/wARP suite, where an advanced template matching technique is applied to the shapes of the density. However, the success of the recognition process strongly depends on the level of informational content in the electron density map. Our on-going development concerns studying the functional $p(x,y,z)$ which describes the electron density map corresponding to peptides and its relation to some distance functional $r(x,y,z)$. The distribution of values of the radial density functional can then be applied to the recognition of the peptide pattern. The method is rotation invariant and its combination with the existing technique that uses the template matching improves the completeness of the protein structure provided by ARP/wARP.

Keywords: density modification, pattern recognition, protein crystallography

**P03.10.10**


Validation and correction of carbohydrate 3D structures

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Carbohydrate structures in PDB entries exhibit a rather high rate of errors [1,2]. Some errors, such as wrong residue names, surplus atoms in the glycosidic linkage, or wrong connectivities, can be
corrected rather easily by changing the residue name, deleting surplus atoms or editing the LINK or CONEET records. Other problems, however, require more sophisticated methods to fix them. To fix e.g. stereochemistry problems, atoms have to be relocated. In some instances, e.g. when wrong stereochemistry is involved in glycosidic linkages, entire residues might have to be rearranged, which involves refinement of the 3d structure. Here, we present an approach to combine a tool for carbohydrate validation (pdb-care [3], www. glycosciences.de/tools/pdb-care/) with the modeling software WHAT IF [4] (swift.cmbi.ru.nl/whatif/) and the refinement tool PDBe-redo [5] (www.cmbi.ru.nl/pdb_redo/) to enable a (semi-)automated correction of erroneous carbohydrate structures in PDB entries.


Keywords: refinement, validation, glycoproteins

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**Restained anisotropic refinement with SHELXL**

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The successful refinement of macromolecular crystal structures usually requires the use of restraints based on our knowledge of similar structures. Whereas geometrical restraints are relatively well understood and quantified, there is still much room for improvement of the restraints applied to the atomic displacement parameters (isotropic and anisotropic temperature factors). We are currently investigating a variety of potential restraints and other features with a view to making the SHELXL program for crystal structure refinement more suitable for the refinement of macromolecular structures at modest resolution. The step from isotropic to anisotropic refinement increases the number of parameters refined by more than a factor of two and often results in over-fitting of the data. A good approach to this problem is the use of TLS constraints (Schomaker & Trueblood, 1968; Winn, Isupov & Murshudov, 2001) because, as usually implemented, the number of extra parameters is limited to 20 per ‘rigid’ domain. However, it is not always easy to subdivide a macromolecular crystal structure into suitable domains and an atom may not be in more than one domain at the same time. A flexible alternative is to extend the rigid bond restraints by Rollett (1970) and known as the DELU restraint in SHELX, to much greater distances (say 6-8 Å) than normally employed. Didisheim and Schwarzenbach (1987) showed that in the limit of very tight restraints this assumtes to the TLS model. The main technical problem in the implementation of such ‘TLS restraints’ for large structures is finding the appropriate atom pairs efficiently, since this needs to be performed each refinement cycle and should take symmetry equivalents into account.

Keywords: restrained refinement, anisotropic atomic displacement parameter, macromolecular structure

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**A molecular dynamics approach to equilibrium structures in crystals**

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The equilibrium structure of a crystal represents the system in a theoretical vibrationless state at the absolute minimum of its potential-energy surface. Such structures can be compared directly between different phases and polymorphs and with theoretical calculations. Structures determined using X-ray and neutron-diffraction experiments are time-averaged over all of the molecular vibrations occurring in the crystal. If these motions are anharmonic or curvilinear then the time-averaged and equilibrium structures will differ. There remains no general and simple method for removing the structural inconsistencies that result. We have recently developed a new method using molecular dynamics (MD) simulations that allows experimental positions to be corrected to equilibrium positions. The corrections are determined by taking the differences between theorectical equilibrium structures and time-averaged structures obtained from MD trajectories. The application of the method to the crystal structure of nitromethane using an empirical force field will be detailed. Comparisons will be made with literature X-ray and neutron data sets. Thermal motion is incorporated into the crystal-structure refinement process using the Debye-Waller factor, which is the Fourier transform of an atom’s vibrational probability density function. This probability function is typically assumed to be a trivariate Gaussian and yields the ubiquitous probability ellipsoids. The MD simulations allow us to determine the true function numerically. In the case of nitromethane this leads to highly curved probability functions for the H atoms.

Keywords: thermal motion, molecular dynamics simulations, Debye-Waller factor

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**Computational chemistry approach to polymorphism of aspirin**

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It is strongly expected that practical techniques on computational analysis of molecular crystal structures can provide knowledge for molecular designs of functional materials or drugs, and for crystallographic studies of solid state phenomena, such as nucleation, growth, polymorphism, and phase transition. To cope with the expectations, we have been developing a computational chemistry system (CONFLEX/KESSHOU) to crystallographic analysis and prediction of conformational and packing polymorphism based on rational evaluations for many energy minima of crystal structures and their dynamical behaviors. In this conference, as an extension of CONFLEX/KESSHOU, we propose our computational investigation for the recent topic on the aspirin crystal polymorphism [1-3]. In order to quantitatively elucidate this polymorphic transition between form I and form II without purported ambiguity, we have calculated energy minimum crystal structures by using CONFLEX/KESSHOU crystal calculations. In comparison with their free energies of two polymorphic forms at room temperature and 100 K, the predominant stability of form I has been computationally ascertained as well.