

P04.01.60*Acta Cryst.* (2008). A64, C249**SAXS and crystal structural analysis of *Helicobacter pylori* GroES**Haur Lee¹, Kuo-Long Lou², Lu-Ping Chow¹¹National Taiwan University College of Medicine, Institute of Biochemistry and Molecular Biology, No.1 Jen Ai Road Section I Taipei 100, Taiwan R.O.C, Taipei, Taiwan, 100, Taiwan, ²Graduate Institute of Oral Biology, Medical College, National Taiwan University, Taipei, Taiwan., E-mail: f95442019@ntu.edu.tw

Helicobacter pylori is a major risk factor of gastric cancer. Although many *H. pylori* virulence factors have been reported, the pathogenic mechanism by which *H. pylori* infection causes gastric cancer remains unclear. Previous studies identified the gastric cancer-related antigens: GroES from *H. pylori* and characterize their roles in the development of gastric cancer. In proteomics study, *H. pylori* GroES was shown as a dominant gastric cancer-related antigen, with a much higher seropositivity of gastric cancer samples compared to gastritis and duodenal ulcer. GroES seropositivity was more commonly associated with antral gastric cancer than with non-antral gastric cancer. Since GroES of *H. pylori* is a novel gastric cancer-associated virulence factor and may contribute to gastric carcinogenesis via induction of inflammation and promotion of cell proliferation. The structure of GroES is important to realize its role in pathogenic mechanism pathway. We try to get the GroES and target molecule complex structure. Until present, we got some SAXS structures and low resolution crystal structures of *H. pylori* GroES. The credo data show that the high similarity between *H. pylori* and *Thermus Thermophilus* GroES. It may tell us the role of *H. pylori* GroES in pathogenic mechanism pathway which can causes gastric cancer. Hopefully we will get into the structural and functional details of GroES molecular mechanism in the near future.

Keywords: GroES, *Helicobacter pylori*, SAXS**P04.01.61***Acta Cryst.* (2008). A64, C249**Crystallographic study of the bacterial prolipoprotein posttranslational lipid modification system**

Yujia Zhai, Zhijie Li, Zhuqing Ouyang, Fei Sun

Institute of Biophysics, Chinese Academy of Sciences, 15 Datun Road, Chaoyang District, Beijing, Beijing, 100101, China, E-mail: yujia@ibp.ac.cn

More than 130 bacterial lipoproteins of diverse structures and functions have been identified to date, and they all contain N-acyldiacylglyceryl-cysteine as their N-terminal amino acid. Posttranslational lipid modification of prolipoproteins in bacteria involves three sequential reactions catalyzed by cytoplasmic membrane enzymes, i.e., prolipoprotein: phosphatidylglycerol diacylglyceryl transferase (LGT), prolipoprotein signal peptidase (LSP), and apolipoprotein N-acyl transferase (LNT), resulting in the formation of N-acyl diacylglycerylcysteine as the N-terminal amino acid of these lipid-modified proteins. This pathway appears to be essential since mutants defective in the activity of any of these three enzymes are temperature sensitive in growth, suggesting that one or more lipoproteins are required for normal growth, division, and viability of bacterial cells. The third and last step of the pathway is the acylation of the N-terminal glyceride-cysteine residue by apolipoprotein N-acyl transferase (Lnt), resulting in mature lipoprotein. Defective mutant study suggests that apolipoprotein N-acyltransferase is an essential enzyme in *S. typhimurium* and *E.*

coli, and lack of this enzyme results in cell death. Moreover Lnt is necessary for efficient recognition of outer membrane lipoproteins by the Lol system, which transports them from the plasma to the outer membrane. The important roles of Lnt mentioned above have hastened us to investigate its structure and its structure-function relationship. In this study, we have initiated crystallographic studies on LGT, LSP and LNT. We have successfully purified these three membrane proteins and obtained the crystals of LGT and LNT.

Keywords: lipid modification, membrane proteins, apolipoprotein N-acyl transferase

P04.01.62*Acta Cryst.* (2008). A64, C249**Preliminary crystallographic studies on ACAP1 BAR-PH and GAP-ANK domains**

Xiaoyun Pang, Jun Ma, Fei Sun

Institute of Biophysics, Chinese Academy Sciences, 15 Datun Road, Chaoyang District, Beijing, 100101, P.R.China, Beijing, Beijing, 100101, China, E-mail: pangxy@moon.ibp.ac.cn

Endocytic recycling is critical for many cellular events, including cell polarity, cell mobility, signal transduction and phagocytosis. ACAP1, a GTPase-activating protein (GAP) for ADP-ribosylation factor (ARF) 6, is part of a novel clathrin coat complex that is regulated by ARF6 for endocytic recycling in two key physiological settings, stimulation-dependent recycling of integrin that is critical for cell migration and insulin-stimulated recycling of glucose transporter type 4 (Glut4), which is required for glucose homeostasis. However, in contrast to TfR recycling that undergoes constitutive recycling, the role of ACAP1 in integrin beta1 recycling requires its phosphorylation by Akt, which is, in turn, regulated by a canonical signaling pathway. ACAP1 and ACAP2, together with ASAP1 and PAP, can be grouped into a protein family defined by several common structural motifs including coiled coil (BAR), pleckstrin homology (PH), Arf GAP, and ankyrin-repeat domains (ANK). Crystals of BAR-PH and GAP-ANK suitable for X-ray crystallography have been obtained and diffraction data have been collected in Photon Factory of KEK separately to 2.6 Å and 3.0 Å resolution. BAR-PH crystal belongs to space group $P2_1$, with unit-cell parameters $a=42.4$, $b=59.8$, $c=167.8$ Å, $\alpha=90^\circ$, $\beta=91^\circ$, $\gamma=90^\circ$. There are two molecules per asymmetric unit. GAP-ANK crystal belongs to space group $P2_12_12_1$, with unit-cell parameters $a=40.4$, $b=107.4$, $c=162.0$ Å, $\alpha=\beta=\gamma=90^\circ$. There are two molecules per asymmetric unit. And crystals of two mutants of GAP-ANK and their complexes with interaction partners of integrin beta1 have been obtained.

Keywords: endocytic recycling, coiled coil, integrin

P04.02.63*Acta Cryst.* (2008). A64, C249-250**A comparative study on substrate specificity, activity and thermal stability of some plant proteases**

Chandana Chakrabarti, Debi Choudhury, Sampa Biswas, Sumana Roy, Sruti Dutta, Jiban K Dattagupta

Saha Institute of Nuclear Physics, Crystallography and Molecular Biology Division, 1/AF Bidhannagar, Kolkata, West Bengal, 700 064, India, E-mail: chandana.chakrabarti@saha.ac.in

The papain-like cysteine proteases constitute an important family of

enzymes. Papain, the archetype member of this family has industrial and pharmaceutical applications. The papain-family cysteine proteases share a typical overall fold, comprising two domains, with the active site located in a cleft at their interface. Their enzymatic activity is related to a catalytic dyad formed by a Cys(-) and His(+). Despite the similarities, variations in properties like substrate specificity, activity and thermal stability have been observed in some of these proteases. Three such proteases, Ervatamins A, B and C, have been isolated from the latex of a tropical plant *Ervatamia coronaria* and characterized. Structural and biochemical studies on these proteases have helped us to identify a few amino acid residues which may be thought to be responsible for substantial changes in their functional properties. This structure-based knowledge is being utilized to design proteases for improved industrial applications.

Keywords: proteases, structure-based protein engineering, industrial applications

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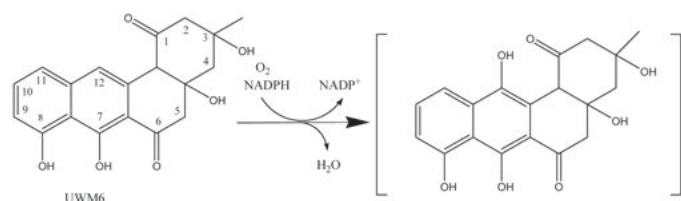
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Aromatic hydroxylases in polyketide antibiotic biosynthesis

Hanna Koskiniemi¹, Mikko Metsa-Ketela^{1,2}, Pauli Kallio², Doreen Dobritzsch¹, Liu Zhanliang², Pekka Mantsala², Jarmo Niemi², Gunter Schneider¹

¹Karolinska Institutet, Department of Medical Biochemistry and Biophysics, Tomtebodavägen 6, 4. tr., Stockholm, Stockholm, 17177, Sweden, ²University of Turku, Vatselankatu 2, 20014 University of Turku, Finland, E-mail: hanna.koskiniemi@ki.se

Aromatic polyketides are a major class of natural products with great medical significance: many representatives are used as e.g. anticancer and antimicrobial agents. However, their use is limited by harmful side-effects and drug resistance, hence new therapeutic agents are needed. Polyketides are produced in complicated enzymatic pathways by certain bacteria, plants and fungi. Engineering of the biosynthesis routes is a promising means of producing novel polyketide drugs, but requires detailed structural and mechanistic information of the biosynthetic enzymes. PgaE and CabE are homologous aromatic hydroxylases from the biosynthesis route of angucycline class of polyketides in *Streptomyces* sp. PGA64 and S. sp. H021. They catalyze the hydroxylation of the C12 of the substrate, UWM6. Their structures have been determined by X-ray crystallography to 1.8 Å and 2.7 Å. CabE and PgaE belong to the p-hydroxybenzoate hydroxylase (pHBH)-family of flavin adenine dinucleotide (FAD)-dependent aromatic hydroxylases. The ordered reaction mechanism includes dynamic rearrangements of the protein and the bound FAD. Unlike pHBH, PgaE and CabE do not appear to activate their substrate via deprotonation.



Keywords: flavin, polyketide antibiotic, aromatic hydroxylase

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Mechanism of stereospecific substrate recognition by LL-diaminopimelate aminotransferase

Nobuhiko Watanabe¹, Maia M Cherney¹, Marco J van Belkum², Sandra L Marcus², Mitchel D Flegel², Matthew D Clay², Michael K Deyholos³, John C Vederas², Michael N. G. James¹

¹University of Alberta, Biochemistry, 429 Medical Sciences Buildings, Department of Biochemistry, Edmonton, Alberta, T6G 2H7, Canada, ²University of Alberta, Department of Chemistry, Edmonton, Alberta, T6G 2G2, Canada, ³University of Alberta, Department of Biological Sciences, Edmonton, Alberta, T6G 2E9, Canada, E-mail: nobuhiko@ualberta.ca

The lysine biosynthetic pathway is an attractive target for the development of new antibiotics or herbicides because it is absent in humans. LL-diaminopimelate aminotransferase (LL-DAP-AT) is a newly discovered enzyme in the novel lysine biosynthetic pathway in *Chlamydia* and plants. Previously, three different lysine biosynthetic pathways have been characterized in bacteria. However, none of the previous bacterial lysine biosynthetic pathways were found in *Chlamydia* or in plants. Recently, LL-DAP-AT was discovered to be the missing piece in *Chlamydial* and plant lysine biosynthetic pathways, and this enzyme bypasses three enzymatic pathways in the previously described bacterial lysine biosynthetic pathway. In order to understand the mechanism of this enzyme and to assist in the design of inhibitors, we have determined the three-dimensional structures of LL-DAP-AT from *A. thaliana* in native and with two substrate-analogues (LL-DAP-PLP, Glu-PLP) bound. LL-DAP-AT is a pyridoxal-5'-phosphate (PLP) dependent enzyme and belongs to the type I fold family of PLP-dependent enzymes. Comparison of the active site residues of LL-DAP-AT and aspartate aminotransferases revealed that the PLP binding residues in LL-DAP-AT are well conserved in both enzymes. However, Tyr37, Tyr152, Glu97 and Asn309 are unique to LL-DAP-AT. Tyr37 and Tyr152 are positioned to recognize distal carboxylate groups of both LL-DAP and glutamate. Glu97, Asn309 and water molecules form an array of hydrogen-bonds to stereospecifically recognize LL-DAP in the active site. Our studies revealed the unique stereospecific recognition mechanism used by this newly discovered LL-DAP-AT.

Keywords: drug targets, aminotransferases, enzymatic mechanisms

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L-Threonine dehydrogenase (TDH) from *T. kodakaraensis*, an enzyme involved in amino acid metabolism

Alexandra Bowyer, Jon B Cooper, Muhammad Akhtar
University of Southampton, School of Biological Sciences, Biochemistry, Bassett Crescent East, Southampton, Hampshire, SO16 7PX, UK, E-mail: alexr@soton.ac.uk

We have determined the structure of threonine dehydrogenase (TDH) from the hyperthermophilic archaeon *Thermococcus kodakaraensis*. It exists as a homotetramer with 1 structural zinc ion per monomer, but is unclear whether a second zinc is required for catalytic activity at the active site, as in many alcohol dehydrogenases. Data was collected to 2.3 Å and molecular replacement was used to solve the structure. Amino acids are essential for cellular growth, repair, and maintenance, although organisms are unable to synthesise all the ones they need themselves. Whilst they are able to synthesise some from chemicals and amino acids, others must be absorbed through the