pockets. This separation might be possible to accept three HMTGs on a gp120 at the same time.

Keywords: lectin proteins; HIV drug design; protein structure analysis

FA1-MS07-P08

Conformational Change of Adenosine Deaminase During Ligand-Exchange in Crystal State. <u>Takayoshi Kinoshita</u>^a, Toshiji Tada^a, Isao Nakanishi^b. ^aGraduate School of Science, Osaka Prefecture University, Japan. ^bDepartment of Pharmaceutical Sciences, Kinki University, Turkey, Japan. E-mail: kinotk@b.s.osakafu-u.ac.jp

Adenosine deaminase (ADA) perpetuates chronic inflammation by degrading extracellular adenosine which is toxic for lymphocytes. ADA has two distinct conformations: open form and closed form [1]. From the crystal structures with various ligands, the non-nucleoside type inhibitors bind to the active site occupying the critical water-binding position and sustain the open form of apo-ADA. In contrast, substrate mimics do not occupy the critical position, and induce the large conformational change to the closed form. However, it is difficult to predict the binding of (+)-erythro-9-(2-hydroxy-3-nonyl)adenine (EHNA), as it possesses characteristic parts of both the substrate and the non-nucleoside inhibitors. The crystal structure shows that EHNA binds to the open form through a novel recognition of the adenine base accompanying conformational change from the closed form of the PR-ADA complex in crystalline state [2]. The open form crystal structure of the EHNA-ADA complex supports our hypothesis that the occupancy at the trigger-water-position is critical for determining the open/closed conformational alternation, rather than the nucleoside framework binding. We believe that the structural penetration of the EHNA-ADA complex and structural comparison of the other inhibitor-ADA complexes will support the discovery of novel ADA inhibitors by structure-based drug design.

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Keywords: adenosine deaminase; EHNA; conformational change

FA1-MS07-P09

The Structure of the Ca²⁺-ATPase Bound to Cyclopiazonic Acid Reveals a Complexed Divalent Ion. J. Preben Morth^a, Mette Laursen^a, Maike Bublitz^a, Karine Moncoq^c, Claus Olesen^b, Jesper Vuust Moeller^b, Howard S. Young^c, Poul Nissen^a. *aDepartment of Molecular Biology, Aarhus, University, Denmark. bDepartment of Physiologyand Biophysics, Aarhus* University, Denmark. *cDepartment of Biochemistry* and National Institute for Nanotechnology, University of Alberta, Canada. E-mail: jpm@mb.au.dk We have determined the structure of the sarco(endo) plasmic reticulum Ca2+-ATPase (SERCA) in an E2.Pi-like form stabilised as a complex with MgF_{4}^{2-} , an ATP analogue (AMPPCP), and cyclopiazonic acid (CPA). The structure determined at 2.5 Å resolution leads to a significantly revised model of CPA binding compared to earlier reports [1,2] showing that a divalent metal ion is required for CPA binding through coordination of the tetramic acid moiety at a characteristic kink of the M1 helix found in all P-type ATPase structures which is expected to be part of the cytoplasmic cation access pathway. Our model is consistent with the biochemical data on CPA [3] function and provides new measures in structure based drug design targeting Ca2+-ATPase from e.g. pathogens. We also present an extended structural basis of ATP modulation pinpointing key residues at or near the ATP binding site. A structural comparison to the Na⁺.K⁺ -ATPase reveals that a Phe93 side chain occupies the equivalent binding pocket of the CPA site in SERCA suggesting an important role of this residue in stabilization of the potassium-occluded E2 state of Na⁺,K⁺-ATPase.

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Keywords: drug design; ATPase; membrane channel transport

FA1-MS07-P10

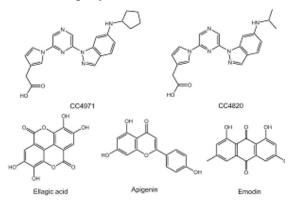
An Approach for Producing a CK2alpha Inhibitor Using X-ray, Calculation and ITC. <u>Yusuke</u> <u>Sekiguchi</u>^a, Harumi Fukada^b, Tetsuko Nakaniwa^a, Takayoshi Kinoshita^a, Shinya Nakamura^c, Isao Nakanishi^{c,d}, Kazuo Kitaura^d, Hiroaki Ohno^d, Yamato Suzuki^d, Akira Hirasawa^d, Gozoh Tsujimoto^d, Toshiji Tada^a. *aGraduate School of Sciences*. *bGraduate School of Life and Environmental Sciences, Osaka Prefecture University*. *CDepartment of Pharmaceutical Sciences, Kinki University*. *dGraduate School of Pharmaceutical Sciences, Kyoto University*. E-mail: <u>sekiguchi07@b.s.osakafu-u.ac.jp</u>

Protein kinase CK2alpha is a highly pleiotropic serine/ threonine protein kinase. CK2alpha plays important roles in cell growth, proliferation, and survival, while it is highly expressed in a wide variety of tumors.(1) Furthermore, CK2alpha is a target protein for glomerulo nephritis (GN) therapy, because an administration of either anitisense oligodeoxynucleotide against CK2alpha or low molecular weight CK2alpha-specific inhibitors effectively prevents the progression of renal pathology in the rat GN models. (2)

To design a novel and potent CK2alpha inhibitor, we determined four X-ray crystal structures of CK2alphainhibitor complexes (cc-04791, cc-04820, apigenin, ellagic acid), and measured enzyme kinetic parameters using ITC (Isothermal Titration Calorimetry) for the respective

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inhibitors. Thermodynamic data, specifically enthalpy (Δ H) and entropy (Δ S), reveal the forces that drive complex formation. Furthermore, binding affinity - K_d in range of millimolar to nanomolar is a powerful information for drug design targeting highly homologous kinases. Supported with computational analysis, these data show the specific contributions of some important residues in ligand-binding, and lead to design a potent inhibitor.



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Keywords: CK2; kinase inhibitor; drug design

FA1-MS07-P11

Structural Characterization of Human Glutathione Transferase A1-1 in Complex with the Anti-cancer Drug Chlorambucil. Lykourgos Chiniadis^a, Kostas Bethanis^a, Nikos Labrou^b, Irene Axarli^b, Katholiki Skopelitou^b, Michael Karpusas^a. *aPhysics Lab.*, Department of Science, Agricultural University of Athens. ^bLab. of Enzyme Technology, Department of Agricultural Biotechnology, Agricultural University of Athens.

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The glutathione transferases (GSTs) are detoxification enzymes that protect the cell from a wide range of potentially harmful electrophilic compounds [1]. Although this action of GSTs protects the organism from harmful environmental toxins, it results also in neutralization of certain therapeutic drugs such as chlorambucil, an anti-cancer drug used to treat chronic lymphocytic leukemia (CLL) [2]. Design of GST inhibitors that mimic chlorambucil binding may be useful in improving the bioavailability of chlorambucil when they are co-administered with the drug. To enable structure-based design of such inhibitors, human GST A1-1 was crystallized in the presence of glutathione and chlorambucil and its structure was determined at 2.3 Å resolution. Chlorambucil is observed to form a conjugate with glutathione, adopting a conformation different from the one observed in the structure of GST P1-1 and to induce local conformational changes in nearby protein residues. In addition it reacts with the only cysteine residue of the enzyme, in agreement with biochemical evidence.

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Keywords: glutathione transferase; anticancer drugs; crystallographic structure determination

FA1-MS07-P12

Targeting Telomeric RNA Quadruplexes. <u>Gavin</u> <u>Collie</u>^a, Stephen Neidle^a, Gary Parkinson^a. ^a*CRUK Biomolecular Structure Group, The School of Pharmacy, London, UK.* E-mail: gavin.collie@pharmacy.ac.uk

Following the discovery that mammalian telomeres are transcribed into telomeric-repeat containing RNA [1],[2], there has been a move to determine the role these molecules play in telomere regulation. We report here the first crystal structure of a telomeric RNA G-quadruplex, providing atomic-resolution insights into RNA-quadruplex folding. The structure, formed from the sequence $r(U_{\rm Br}AGGGUUAGGGU)$, folds as a bimolecular, parallel stranded G-quadruplex with linking propeller-type external loops, a topology previously observed for the equivalent telomeric DNA sequence [3].

As telomeric DNA G-quadruplexes are well established targets for small molecule anti-cancer therapeutics, it is reasonable to consider telomeric RNA G-quadruplexes as potential targets too. The structure reported here provides the basis for computer aided drug design, and will hopefully allow the differences between RNA and DNA G-quadruplexes to be identified and exploited for improved specificity of quadruplex-interacting ligands. These issues will be addressed and the structural basis for drug design explored.



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Keywords: RNA structure; anticancer drug structural study; drug targets

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