This structure and the result of inhibition by ADP will be available for design of new drugs which inhibit ADPRTs such as S. enterica SpvB, which is required for human macrophage infection.

H. Tsuge, et al. *J Mol Biol* **2003**, *325*, 471-483
H. Tsuge, M. Nagahama, M. Oda, S. Iwamoto, H. Utsunomiya, V.E. Marquez, N. Katunuma, M. Nishizawa, J. Sakurai, *PNAS* **2008**, *105* (*21*), 7399-7404.

#### Keywords : ADPRT, iota toxin, complex structure

#### MS36.P16

Acta Cryst. (2011) A67, C475

## Structural basis for the helicobacter pylori-carcinogenic $TNF\langle -inducing\ protein$

Hideaki Tsuge,<sup>a,b</sup> Toshiharu Tsurumura,<sup>a</sup> Hiroko Utsunomiya,<sup>b</sup> Daisuke Kise,<sup>c</sup> Takashi Kuzuhara,<sup>c</sup> Tatsuro Watanabe,<sup>d</sup> Hirota Fujiki,<sup>c</sup> Masami Suganuma<sup>d</sup> <sup>a</sup>Faculty of Life Sciences, Kyoto Sangyo University, Kyoto, (Japan). <sup>b</sup>Institute for Health Sciences, Tokushima Bunri University, Tokushima, (Japan). <sup>c</sup>Faculty of Pharmaceutical Sciences, Tokushima Bunri University, Tokushima, (Japan). <sup>d</sup>Research Institute for Clinical Oncology, Saitama Cancer Center, Saitama, (Japan).

Stomach cancer is strongly associated with infection by Helicobacter pylori. In 2005, we identified a new H. pylori gene encoding a TNF- $\langle$  inducing protein (Tip $\langle$ ) that acts as a carcinogenic factor. Tip $\langle$  is secreted from H. pylori as a homodimer whose subunits are linked by disulfide bonds. Using blast search, there is no similar sequence with Tip $\langle$ , thus it is unique carcinogenic factor protein. We also characterized a Tip $\langle$  deletion mutant (del-Tip $\langle$ ) that lacks the N-terminal six amino acid residues (LQACTC), including two cysteines (C5 and C7) that form disulfide bonds, but nonetheless shows a weak ability to induce TNF-a expression.

Here we report the crystal structure of del-Tip( at 2.47Å resolution. As expected, the structure of del-Tip( is novel; It has a novel elongated structure containing a 40 Å-long a helix, and forms a heart-shaped homodimer via non-covalent bonds. Moreover, their circular dichroism spectra strongly suggest that the structures of the del-Tip( and Tip( homodimers are very similar. We conclude that the TNF-( inducing activity is correlated with the dimer formation. It means that strong dimer via covalent bond shows the strong activity for Tip( and weak activity for del-Tip( with weak interaction. Tip('s unique mode of dimer formation provides important insight into protein-protein interactions and into the mechanism underlying the carcinogenicity of H. pylori infection.

[1] H. Tsuge, T. Tsurumura, H. Utsunomiya, D. Kise, T. Kuzuhara, T. Watanabe, H. Fujiki, M. Suganuma. *Biochem Biophys Res Commun.* **2009**, *388*(2), 193-8.

Keywords: TNF-( inducing protein, helicobacter pylori, carcinogenic factor

#### MS36.P17

Acta Cryst. (2011) A67, C475

# A Model of action for peripheral Membrane-Associated GT-B Glycosyltransferases

Saioa Urresti,<sup>a,b</sup> David Giganti,<sup>a,b,c</sup> Marco Bellinzoni,<sup>c</sup> Mary Jackson,<sup>e</sup> Pedro M. Alzari<sup>c</sup> and Marcelo E. Guerin,<sup>a,b,d</sup> <sup>a</sup>Unidad de Biofísica, Centro Mixto Consejo Superior de Investigaciones Científicas -Universidad del País Vasco/Euskal Herriko Unibertsitatea (CSIC-UPV/EHU), Barrio Sarriena s/n, Leioa, Bizkaia, 48940, (Spain). <sup>b</sup>Departamento de Bioquímica y Biología Molecular, UPV/EHU, (Spain). <sup>c</sup>Unité de Biochimie Structurale (CNRS URA 2185), Institut Pasteur, 25 rue du Dr. Roux, 75724, Paris Cedex 15, (France). <sup>d</sup>IKERBASQUE, Basque Foundation for Science, 48011, Bilbao, (Spain). <sup>e</sup>Department of Microbiology, Immunology and Pathology, Colorado State University, Fort Collins, Colorado 80523-1682, Email: mrcguerin@gmail.com

Peripheral membrane-associated GT-B glycosyltransferases (GTs) are a ubiquitous family of enzymes that play essential roles in a variety of important biological processes in all living organisms. They transfer a sugar moiety from nucleotide- or lipid-phospho-sugar donors to a wide range of membrane-associated acceptors. Here we focus in PimA, an essential enzyme involved in the biosynthesis of phosphatidylmyo-inositol mannosides (PIMs), which are key glycolipids of the mycobacterial cell envelope<sup>1</sup>. PimA is a paradigm of this family of GTs, which the molecular mechanism of substrate/membrane recognition and catalysis is still unknown. We have solved the crystal structure of PimA from M. smegmatis in complex with its donor substrate GDP-Man<sup>2</sup>. The notion of a membrane-associated protein via electrostatic interactions is consistent with the finding of an amphipathic *a*-helix in the N-terminal domain of PimA. Based on structural, biophysics and biochemical studies, we proposed a model of interfacial catalysis in which PimA recognizes the fully acylated acceptor substrate, phosphatidyl-myo-inositol (PI), with its polar head within the catalytic cleft and the fatty acid moieties only partially sequestered from the bulk solvent. In addition, we provided strong evidence showing that PimA undergoes significant conformational changes upon substrate binding<sup>3</sup>. Altogether, our experimental data support a model wherein the flexibility and conformational transitions confer adaptability of PimA to the substrates, which seems to be of importance during catalysis. The proposed mechanism has fundamental implications for the comprehension of the peripheral membrane-associated GTs at the molecular level.

M.E. Guerin, J. Korduláková, P.M. Alzari, P.J. Brennan, M. Jackson, *Journal of Biological Chemistry* 2010, 285, 33577-33583. [2] M.E. Guerin, F. Schaeffer, A. Chafotte, P. Gest, D. Giganti, J. Korduláková, M. Van der Woerd, M. Jackson, P.M. Alzari, *Journal of Biological Chemistry* 2009, 284, 21613-21625. [3] M.E. Guerin, J. Korduláková, F. Schaeffer, Z. Svetlikova, A. Buschiazzo, D. Giganti, B. Gicquel, K. Mikusova, M. Jackson, P.M. Alzari, *Journal of Biological* Chemistry 2007, 282, 20705-20714.

Keywords: membrane glycosyltransferase, tuberculosis, catalysis

### MS36.P18

Acta Cryst. (2011) A67, C475-C476

### Structural studies on FlhBc from *Salmonella typhimurium* and *Aquifex aeolicus*

<u>Vladimir A. Meshcheryakov</u>, Clive S. Barker, Irina V. Meshcheryakova, Fadel A. Samatey, *Trans-membrane Trafficking Unit, Okinawa Institute of Science & Technology, 1919-1 Tancha, Onna-son, Kunigami-gun, Okinawa (Japan)*. E-mail: v.meshcheryakov@oist.jp

Many bacteria swim in liquid environment by means of flagella. The bacterial flagellum is a huge complex structure made from more than 30 different proteins. All flagellar axial proteins are transported across cytoplasmic membrane by the flagellum-specific secretion apparatus, which shares similarity to the type III secretion system of the bacterial needle utilized by some bacteria in pathogenesis. The protein transport is highly regulated. Membrane protein FlhB has been found to play an active role in this regulation. The protein consists of two domains: a hydrophobic N-terminal part, which is predicted to have four transmembrane helices, and a C-terminal cytoplasmic