

XAS structural insights into Cu binding with amyloid Aβ4-γ peptides

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Alzheimer's disease (AD) is a progressive neurodegenerative disorder characterized by the presence of amyloid (Aβ) plaques. In addition to modulating senile Aβ plaque formation, Aβ-Cu complexes are involved in extensive redox chemical reactions. The first protein sequencing studies of the Aβ plaque core (APC) of AD patients identified NH2-terminal heterogeneity and the majority (64%) of the APC-AD Aβ peptides begins with a Phe4 residue. This N-truncated sequence produced the high-affinity (dissociation Kd of femtoM) amino-terminal Cu Ni (ATCUN, H2N-Xaa-Yaa-His) binding motif.

X- ray absorption spectroscopy (XAS) combined with electrochemical control (XAS-SEC) was utilised to derive structural models for the interaction of CuI and CuII with N-truncated Aβ peptides. First, the conventional XAS of CuII:Aβ1-16 and CuII:Aβ4-γ (γ=8,12,16) as frozen solutions (5-10 K, Fig. 1a,b) and secondly XAS-SEC at room temperature under potentiostatic control have been measured (Fig. 1c). The performance of the XAS-SEC cell in the Australian Synchrotron XAS beamline is demonstrated by the current response to a step potential and to application of a triangular waveform (Fig. 1d). The XAS of CuII:Aβ4 8 and CuII:Aβ4-16 show the oscillations characteristic for CuII binding to the ATCUN site and prove that Tyr10 or Glu11 are NOT involved in CuII binding at pH of 7.4 in our studies. The binding geometry is different from the CuII binding site of Aβ1-16 involving 3 Histidine: His6-His13-His14 (Fig. 1a) [1].

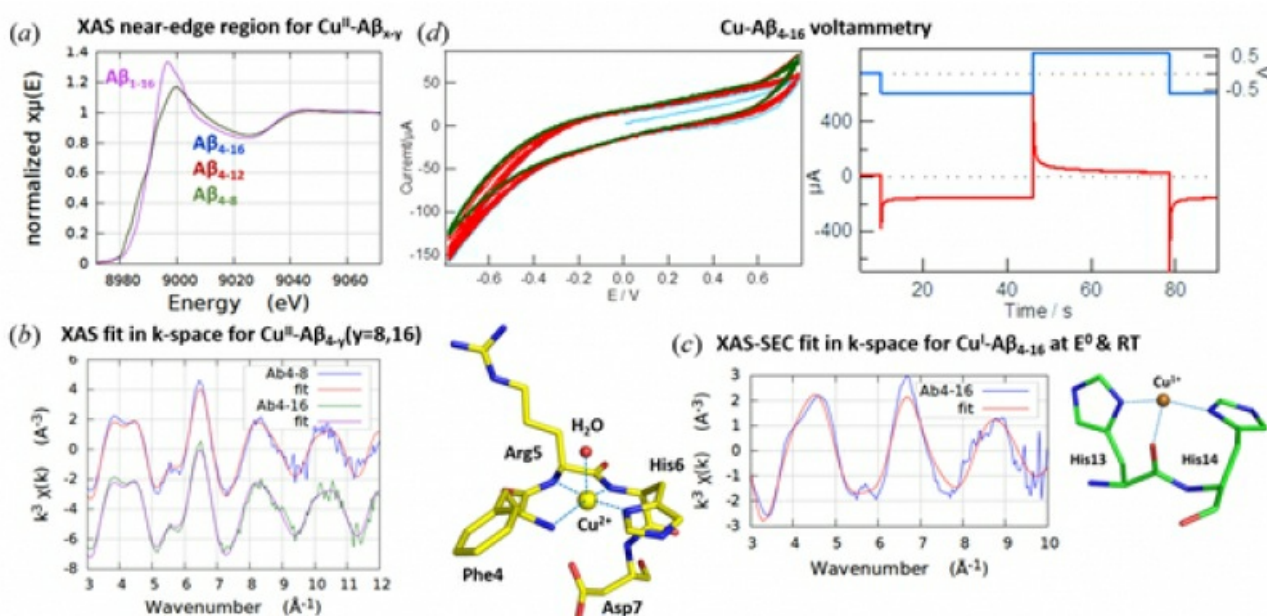
The reduction potential, E°(CuII/I), depends on the relative strength of CuII/CuI binding where the ATCUN motif available only for the truncated Aβ4-γ peptide giving both higher Kd and more negative reduction potential, i.e. E°(CuII/I:Aβ1-16) >> E°(CuII/I:Aβ4-γ). The reduction of CuII:Aβ1-16 is irreversible with CuI rearrangement of the 3His to almost linear geometry His13-His14 binding site [2]. While not observable by electrochemical measurements, reduction of CuII:Aβ4-γ can be monitored by XAS-SEC. More forcing conditions are needed to reduce CuII:Aβ4-12 than CuII:Aβ4-16. CuI binding at the His13-His14 site is inferred by the XAS analysis of spectra recorded during in situ reduction of CuII:Aβ4-16 (Fig. 1c).

These results suggest that reduction of CuII:Aβ4-16 proceeds from CuII bound to the His13-His14 site (Aβ4-16:CuIHis13,14), possibly via His6-His14 "preorganization" [3] which is in equilibrium with Aβ4-16:CuIIATCUN.

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