Structural studies of a novel ubiquitin-modifying enzyme, SdeA using various tools

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Post-translational attachment of ubiquitin (Ub) is one of the most common protein modifications and is involved in a number of key cellular processes in eukaryotes. Conventional ubiquitylation occurs through an ATP-dependent three-enzyme cascade (E1, E2, and E3) that mediates the covalent conjugation of the C-terminus of ubiquitin to a lysine on the substrate. SdeA, which belongs to the SidE effector family of Legionella pneumophila, can transfer ubiquitin to endoplasmic reticulum-associated Rab-family GTPases in a manner independent of E1 and E2 enzymes [1]. The novel ubiquitin-modifying enzyme SdeA utilizes NAD+ as a cofactor to attach ubiquitin to a serine residue of the substrate [2,3].

Here, we determined the overall structure of the 180 kDa full-length SdeA and the spatial orientation of the N-terminal deubiquitylase (DUB), phosphodiesterase (PDE), mono ADP-ribosyltransferase (mART), and C-terminal coiled-coil (CC) domains using small angle X-ray scattering (SAXS) technique. The full-length shape was checked through negative staining. Besides, crystal structures of the mART-C domain of SdeA was determined in free form and in complex with NAD+ at high resolution. The sub-domains of mono ADP-ribosyltransferase termed mART-N and mART-C were characterized to elucidate the coupled enzymatic reaction of NAD+ hydrolysis and ADP-ribosylation of ubiquitin. These results provide insight into the unusual ubiquitylation mechanism of SdeA and expand our knowledge of ubiquitin biology.

References

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