

**PS04.05.15 THE CRYSTAL STRUCTURE OF THE HUMAN PAPILLOMAVIRUS 31 E2 DNA BINDING DOMAIN IN THE ABSENCE OF DNA.** V. L. Giranda, X. Kong, D. Egan, F. Lindh, T. Holzman, H. S. Yoon, and T. Robins, Abbott Laboratories, Abbott Park, IL 60064

Human papillomaviruses (HPVs) are a causative agent for proliferative epithelial lesions (e.g., warts). Certain HPV serotypes have been causally linked to the development of cervical carcinoma. The papillomavirus E2 gene product is a transacting transcriptional regulator. E2 is required for viral replication and is comprised of a DNA binding and activation domains. Previously only the structure of the bovine papillomavirus (BPV) E2 DNA binding domain bound to DNA has been reported.

The crystal structure of the HPV type 31 E2 DNA binding domain has been solved to 2.5 Å resolution in the absence of DNA. The structure is similar to the BPV E2 DNA binding domain. The monomer is comprised of four beta strands which form a beta sheet as well as two alpha helices. The first helix is the DNA recognition helix. The HPV structure, like that of BPV, shows that the DNA binding domains form a tightly associated dimer, with a four stranded beta sheet from each monomer contributing to an eight stranded beta sandwich at the dimer core. The remaining two helices (per monomer) reside on the outside of the dimer. The DNA recognition helices in the absence of DNA are similar to the those seen in the presence of DNA.

The loop between the second and third beta strand is disordered in the HPV structure (no DNA). In the BPV structure this loop is not disordered and associates with the DNA sugar phosphate backbone. Buried at the center of the eight stranded beta sandwich are four hydrophilic residues, two from each monomer, that coordinate a water or solvent ion. This differs significantly from the BPV where the core is occupied exclusively by hydrophobic residues.

This structure will provide a platform on which to base future structure aided drug design. The aim of this design is to create compounds that will inhibit the replication of HPV.

**PS04.05.16 STRUCTURE OF THE LACTOSE OPERON REPRESSOR AND ITS COMPLEXES WITH DNA AND INDUCER.** M. A. Kercher\*, G. Chang†, N. C. Horton\*, P. Lu\*, J. H. Miller‡, H. C. Pace\*, M. Lewis‡, †Department of Biochemistry and Biophysics and \*Department of Chemistry, University of Pennsylvania, Philadelphia, PA 19104, USA and ‡Department of Microbiology and Molecular Genetics, University of California, Los Angeles, CA 90024, USA

In 1961 Jacob and Monod proposed a model for gene regulation which was widely adopted as a paradigm. The model was based on a "repressor" molecule that had not at that time been isolated but later became known as *lac* repressor. Determination of the 3-dimensional structures of three different forms of the *lac* repressor protein (the intact *lac* repressor, the *lac* repressor bound to the gratuitous inducer isopropyl-β-D-thiogalactoside (IPTG) and the *lac* repressor complexed with two 21 base-pair symmetric operator DNA duplex deoxyoligonucleotides) makes the analysis of the mechanism of gene regulation possible. Comparison of the induced and repressed forms of the protein at the dimer level show that a conformational change in the core of the structure results in changes in the distant DNA-binding areas of the protein. Examination of the protein at the tetramer level leads to a model of a 93 base-pair repression loop corresponding to the *lac* operon -82 to +11 region in which *lac* repressor acts with the catabolite activating protein is presented. A correlation of the site-specific mutational analysis with the 3-dimensional structures is also presented. (supported by NIH and US ARO)

**PS04.05.17 INHIBITION OF HIV-1 REVERSE TRANSCRIPTASE VIA A SUBSTRATE INHIBITOR COMPLEX: STRUCTURAL IMPLICATIONS.** Mary L. Kopka, Mark Filipowsky and Richard E. Dickerson, Molecular Biology Institute, University of California at Los Angeles, CA 90095, USA

A bis-linked distamycin drug using m-pyridyl as the linker was studied by kinetics, gel shift and nucleic acid melting point, ΔT<sub>m</sub>, analysis as an inhibitor of HIV-1 reverse transcriptase (RT). IC50 curves run with enzyme pre-incubated with template-primer (TP) show a 4-fold increase in inhibition over conditions with TP pre-incubated with drug. This fact, coupled with increased binding of TP to RT in response to drug in gel shift assays, and weak binding of the inhibitor to the RNA/DNA TP alone from ΔT<sub>m</sub> assays, indicates formation of a dead-end ternary complex of drug, enzyme and TP. The kinetics indicate that the inhibition occurs before processive synthesis begins. The drug-bound to the RNA/DNA template-primer is the inhibiting species, i.e. a substrate-inhibitor complex (SI). This SI complex is competitive with TP on the same site in a transition enzyme-TP complex and to a lesser extent on free enzyme. This dead-end ternary transition complex is the main inhibition intermediate.

These results will be interpreted considering the relevant conformational changes the enzyme undergoes immediately prior to and during reverse transcription.

**PS04.05.18 CRYSTAL STRUCTURE OF KLENOW-ANALOGOUS FRAGMENT OF *THERMUS AQUATICUS* DNA POLYMERASE I AT 2.5 Å RESOLUTION.** Sergey Korolev, Murad Nayal, Wayne M. Barnes, Enrico Di Cera, and Gabriel Waksman, Department of Biochemistry and Molecular Biophysics, Washington University School of Medicine, St Louis, MO 63110, USA

The crystal structure of the large fragment of the *Taq* DNA polymerase (Klentaq1), determined at 2.5 Å resolution, demonstrates a compact, two domain architecture. The C-terminal domain is identical in fold to the equivalent region of the Klenow fragment of *Escherichia coli* DNA polymerase I (Klenow Pol I). Although the N-terminal domain differs greatly in sequence from its counterpart in Klenow Pol I, it has clearly evolved from a common ancestor. The structure of Klentaq1 reveals the strategy utilized by this protein to maintain activity at high temperatures and provides the structural basis for future improvements of the enzyme.

Ref:

S. Korolev et al., (1995) Proc. Natl. Acad. Sci. USA 92, 9264-9268.

**PS04.05.19 STRUCTURAL ANALYSIS OF THE TBP/TFIIB/DNA COMPLEX FROM THE HYPERTHERMOPHILIC ARCHAEA PYROCOCCLUS WOESSEI.** P. Kosa, B. DeDecker, G. Ghosh, P. B. Sigler, Department of Molecular Biophysics and Biochemistry, Yale University, New Haven, CT 06510

Eukaryotic transcription Factor IIB is necessary for basal or activated transcription. TFIIB forms a complex with TATA-binding protein (TBP) and DNA. The crystal structure of this complex has been solved. TFIIB has been shown to directly interact with TBP, TFIIF, and RNA polymerase II, and is proposed to be involved in mediating activated transcription through an interaction with activators.

Homologs of TBP and TFIIB have been found in several Archaea species, suggesting that Archaea have a eukaryotic like pol II transcriptional apparatus. *Pyrococcus woesei*, an archaea from deep sea thermal vents which exhibits a maximum growth temperature of 110°C, has an A/T rich, TATA-like promoter se-