# MS19 P01

Molecular assembiles of protein degradation pathways in prokaryotes and eukaryotes Borissenko L, Groll M. Department of Structural research, Institute of Biochemistry, Charité Medical School of Humbold University of Berlin, Monbijoustrasse 2, 10117 Berlin, Germany

### Keyword 1 : proteolysis, crystal structure, proteasome

Protein degradation is an essential and strictly controlled process with proteasome and functionally related proteases representing its central part. Tricorn protease (TRI) has been shown to act downstream of the proteasome, degrading produced peptides. Recently, a novel large prokaryotic aminopeptidase oligomeric complex, named TET, has been identified. This complex degrades peptides of different length in organisms where TRI is not present. We determined the crystal structure of TET from the thermophilic archaeon Pyrococcus horikoshii at 1.6 A resolution in native form and in complex with the inhibitor amastatin. We demonstrate that, beside the novel tetrahedral oligomerisation pattern, TET possesses a unique mechanism of substrate attraction and orientation. TET sequentially degrades peptides produced by the proteasome to single amino acids. Furthermore, we reconstituted in vitro the minimal protein degradation system from initial unfolding of labelled protein substrates, up to release of free amino acids. We propose that TET and TRI act as functional analogues in different organisms, with TET being more widely distributed. Thus, TET and TRI represent two evolutionarily diverged pathways of peptide degradation in prokaryotes. Our current interest is to follow the mechanisms of supramolecular assemblies which occur during the process of proteolysis in pro- and eukaryotes, using combination of various structural, biochemical and biophysical methods. Participants of the proteolytic process such as unfoldases, proteasome and downstream peptidases are marked with fluorescent tag-proteins or dyes in order to visually follow the formation of complexes upon addition of the substrate protein/peptide.

#### MS19 P02

Synthesis, Structure and Non-Linear Optical Activity of New 1D, 2D, and 3D Cadmium(II) and Zinc(II) Azido complexes. <u>Morsy M.A.M. Abu-Youssef</u><sup>a</sup>, Vratislav Langer<sup>b</sup>, Alshima'a A. Massoud<sup>b</sup>, Maher Abd El-Megeed Farhoud<sup>c</sup>, <sup>a</sup>Chemistry Department, Faculty of Science, Alexandria University, Egypt. <sup>b</sup>Environmental Inorganic Chemistry, Department of Chemical and Biological Engineering, Chalmers University of Technology, Sweden. <sup>c</sup>Physics Department, Faculty of Science, Alexandria University, Egypt. E-Mail: morsy5@link.net

#### Keywords: complexes, azides, NLO

The design and synthesis of versatile molecular building blocks towards self-assembly functional materials is one of our interests. New thermally stable coordination compounds based on 3D nets were synthesized and characterized. Cadmium (II) and zinc (II) azido polynuclear complexes were found to have not only an intriguing structures but also Non-Linear Optical (NLO) properties. Recently we have synthesized a series of high dimensionality Cd (II) and Zn (II) azido complexes: **3D**-  $[Cd_3(nic)_4(N_3)_2(H_2O)_2]_n$  (1),  $[Zn(nic)(N_3)]_n$  (2), and  $[Cd(2,5-dmpyz)(N_3)_2]_n$  (3), 2D-  $[Cd(Quz)_2(N_3)_2]_n$  (4), 1D- $[Zn(bipy)(N_3)_2]_n$  (5),  $[Cd(bipy)(N_3)_2]_n$  (6),  $[Cd(Qux)_2(H_2O)]_n$  (7) and  $[Cd(2-acpy)(N_3)_2]_n$  (8), where (nic = nicotinate anion; 2,5-dmpyz = 2,5dimethylpyrazine, Quz = quinazoline, bipy = 2,2'bipyridyl, Qux = quinoline-4- carboxylato anion and 2acyp = 2-acetylpyridine), [1-4]. Most of these complexes show higher second harmonic generation (SHG) efficiency than LiNbO<sub>3</sub> and potassium dihydrogen phosphate (KDP). Coordination polyhedra of  $[Cd_3(nic)_4(N_3)_2(H_2O)_2]_n$  (1)

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## MS19 P03

#### Redetermination of the A-amylose crystal structure

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#### Keywords: amylose, single-crystal X-ray diffraction

The structure of A-amylose has been revisited by singlecrystal microdiffraction. This allows determining of structural differences as compared to previous fiber and powder diffraction data [1]. Recent progress in synchrotron radiation microdiffraction [2] has now made feasible the collection of X-ray data sets on micrometer sized single crystals. In the present note we report the crystal structure of A-amylose based on low temperature synchrotron radiation microdiffraction. Needle-shaped Aamylose single crystals of less than 15 m length and about 2 m thickness have been crystallized from diluted aqueous solutions of amylose fractions [3]. Experiments were performed at the ESRF ID13 beamline at wavelength of  $\lambda$ =0.9465 Å. The beam was focused by parabolic Berefractive lenses and collimated to either 10µm or 30µm at the ID13 microgoniometer [2]. Diffraction patterns were recorded at 100K by a MAR165 CCD and processed by XDS data reduction package [4]. We have already reported previously unit cell parameters and space group (B2) of A-amylose [5]. The intensity data collected on 14 crystals were merged together using XSCALE software [4]. The resulting data set has 1.3Å highest resolution. average I/ $\sigma$  9.37 and redundancy 5.9. The structure of Aamylose was solved by molecule replacement technique using PATSEE software [6]. A fragment of a regular 6fold helix of 3 residues length was used as a search fragment. Positions of primary hydroxyl groups and all positions of water molecules were found on electron difference map. The structure was refined in isotropic approach of thermal parameters using restraints up to R1=0.1749, R<sub>free</sub>=0.2216. The principal structural model obtained is the same as determined previously by X-ray powder and fiber methods [1]. The basic units of the Aamylose crystal framework are six-fold left-handed double helices having gauche-gauche conformations of primary hydroxyl groups. The helices are arranged around two fold symmetry axes and have repeat 2c=2.116 nm. The present