Bispecifics and CAR T cells that target intracellular cancer driver mutations

Sandra Gabelli¹, Katharine Wright², Micheller Miller³, Emily Han-Chung Hsiue⁴, Michael S. Hwang⁵, Puchong Thirawatananond⁶, P. Aitana Azurmendi⁷, Jackie Douglass⁸, Tihitina Aytenfisu⁹, Michael Murphy¹⁰, Brian J Mog¹¹, Alexander H. Pearlman¹², Drew M. Pardoll¹³

¹Johns Hopkins University ²No affiliation given, ³Johns Hopkins University, ⁴Sidney Kimmel Comprehensive Cancer Center, ⁵Sidney Kimmel Comprehensive Cancer Center, ⁶Johns Hopkins University, ⁷Department of Biophysics and Biophysical Chemistry, ⁸Johns Hopkins University, ⁹Johns Hopkins University School of Medicine, ¹⁰Cytiva, ¹¹Sidney Kimmel Comprehensive Cancer Center, ¹²Sidney Kimmel Comprehensive Cancer Center ¹³Sidney Kimmel Comprehensive Cancer Center

gabelli@jhmi.edu

The emergence of immunotherapy as an important tool in the fight against cancer takes advantage of the exquisite selectivity of antibodies. Until now successful targets, however, are limited to those on the cell surface, while most driver mutations occur in the genes encoding intracellular proteins. To overcome this limitation, antibodies can be engineered to target mutation derived neoantigens, peptides derived from mutant proteins that are presented on the cell surface by the Major Histocompatibility Complex Class I (MHC-I). In cases where both the wild-type (WT) and mutant peptide are presented, antibodies can be developed that selectively target the specific mutant peptide-HLA complex. We have selected antibody fragments by phage display that target mutant peptides derived from a tumour suppressor gene, specifically the R175H mutation in the protein p53 and from an oncogene (1), the R140Q mutation in isocitrate dehydrogenase 2 (IDH2).

The IDH2R140 and IDH2R140Q peptide-HLA:B*07 complexes revealed each peptide bound in an almost identical conformation with the mutant residue buried deep in the peptide-HLA-B*07 binding cleft. The structure of the antibody fragment, 2Q.1, in complex with the IDH2R140Q peptide-HLA:B*07 displayed a subtle backbone movement in the mutant peptide that underlies the observed antibody selectivity. Structure-guided optimization of the targeting moiety resulted in the development of a CAR T cell that lacks WT binding making it more selective and with improved on-target activity in response to IDH2R140Q-HLA-B*07, as assessed by IFN-γ release. In contrast to the buried epitope in IDH2R140Q-HLA:B*07, the histidine residue in the p53 neoantigen-HLA:A*02 structure is exposed and available for antibody binding1. Structural basis of selectivity of the antibody against p53R175H, H2, is supported by a "cage-like" structure formed by tyrosine residues in three complementarity determining regions. Grafting of H2 anti p53R175H into a bispecific antibody with an anti CD3 fragment is effective in activating T cells to secret cytokines and kill cancer cells. Moreover, treatment with H2 resulted in regression of human tumor xenografts in mice. Moreover, to investigate potential on-target off-tumour reactivity, the human peptidome was screened in silico for the recognition motif of the H2 antibody. Of the three possible candidate proteins identified, none showed evidence of triggering T cell activation via H2 bispecific in vitro.

The exquisite selectivity achievable with antibodies provides the added benefit of distinguishing between WT and mutant proteins-the foundation for developing effective treatments with minimal adverse effects to patients. While it remains to be determined which class of precision therapeutic will prove more efficacious in humans, we present here two avenues to treat cancer harboring public intracellular targets(2). The 2Q.1 CAR T cell approach allows for the incorporation of antigen dependent co-stimulation, a property that has proved essential in the successful translation of CAR T cells to the clinic. On the other hand, the H2 bispecific to treat p53R175H dependent cancers has the potential to be an "off-the-shelf" therapy, that does not require manipulation of patients own T cells and has a relatively simple manufacturing process.

1. Han-Chung Hsie E, Wright KM, Douglas J, Hwang MS, Mog BJ, Pearlman AH, Paul S, DiNapoli SR, Konig MF, Wang Q, Schaefer A, Miller MS, Skora AD, Azurmendi PA, Murphy MB, Liu Q, Watson E, Li Y, Pardoll DM, Bettegowda C, Papadopoulos N, Kinzler KW, Vogelstein B, Gabelli SB*, Zhou S. Targeting a neoantigen derived from a common TP53 mutation. Science. 2021 Mar 5;371(6533):eabc8697. doi: 10.1126/science.abc8697. Epub 2021 Mar 1. PMID: 33649166

2. Pearlman AH, Hwang MS, Konig MF, Han-Chung Hsie E, Douglass J, DiNapoli SR, Mog BJ, Bettegowda C, Pardoll DM, Gabelli SB, Papadopoulos N, Kinzler KW, Vogelstein B, Zhou S. Targeting public neoantigens for cancer immunotherapy. Nature Cancer volume 2, pages 487-497 (2021),