

# Time-resolved serial femtosecond crystallography on photoswitchable fluorescent proteins

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Time-resolved serial femtosecond crystallography (TR-SFX) at X-ray free electron lasers (XFELs) allows studying the structural dynamics of crystalline biological macromolecules down to the sub-picosecond time scale [1]. According to a pump-probe scheme, optical pump pulses initiate activity in light sensitive crystalline proteins and XFEL pulses generate diffraction patterns that allow determining intermediate-state structures. We apply TR-SFX to study light-induced dynamics in a reversibly photoswitchable fluorescent protein, rsEGFP2.

Reversibly photoswitchable fluorescent proteins are essential tools in advanced fluorescence nanoscopy of live cells. They can be repeatedly toggled back and forth between a fluorescent (*on*) and a non-fluorescent (*off*) state by irradiation with light at two different wavelengths. Our consortium (\*) combines TR-SFX at XFELs, ultrafast absorption spectroscopy and simulation methods to study photoswitching intermediates in rsEGFP2 on the picosecond to nanosecond time scale. We have been able to identify the transient structure of rsEGFP2 in its excited state 1 ps after photoexcitation, and to observe the chromophore in a twisted state, midway between the stable configurations of the *on* and *off* states [2]. This observation, together with a ground-state intermediate structure determined 10 ns after photoexcitation, has allowed us to uncover details of the photo-switching mechanism of rsEGFP2 [3].

Based on the reaction intermediates determined by TR-SFX [2, 3] two rationally designed mutants of the rsEGFP2 have been generated. Pico- to nanosecond TR-SFX results experiments on these rsEGFP2 variants have been carried out at SACLA and the LCLS and provide insight into modified energy landscapes (unpublished).

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