

Oral presentation

Study of the conformational dynamics of a bacterial photoactivated adenylate cyclase

S.M. Kapetanaki^{1,*,}, N. Coquelle¹, D. von Stetten², M. Byrdin¹, R. Rios-Santacruz¹, R. Bean³, J. Bielecki³, E. Bódis⁴, M. Boudjelida¹, Z. Fekete⁴, G. W. Grime⁵, H. Han³, C. Hatton⁶, S. Kantamneni³, A. Kengyel⁴, K. Kharitonov³, C. Kim³, M. Kloos³, F.H.M. Koua³, I. de Diego Martinez³, C. Mas¹, D. Moussaoui⁷, D. Melo³, M. Nyitrai⁴, I. Pécsi⁴, P. Pernot⁷, L. Rane¹, A. Round³, E. Round³, A. Sarma³, R. Schubert³, J. Schulz³, M. Sikorski³, E. Telek⁴, M.D. Tully⁷, K. Ujfalusi-Pozsonyi⁴, M. Vakili^{3,§}, J. Valerio³, N. Varnyuné Kis-Bicskei⁴, J. Vitas¹, R. de Wijn³, A. Wrona³, N. Zala¹, A. Pearson⁶, K. Dörner³, G. Schiró¹, E. F. Garman⁸, A. Lukács⁴, M. Weik¹

¹Univ. Grenoble Alpes, CEA, CNRS, Institut de Biologie Structurale, F-38044 Grenoble, France, ²European Molecular Biology Laboratory (EMBL), Hamburg unit c/o DESY, Notkestr. 85, 22607 Hamburg, Germany, ³European XFEL, Holzkoppel 4, 22869 Schenefeld, Germany, ⁴Department of Biophysics, Medical School, University of Pécs, Szigeti Street 12, 7624 Pécs Hungary, ⁵University of Surrey Ion Beam Centre, GU2 7XH Guildford, United Kingdom, ⁶Institute for Nanostructure and Solid-State Physics, Universität Hamburg, HARBOR, Luruper Chaussee 149, 22761 Hamburg, Germany, ⁷European Synchrotron Radiation Facility (ESRF), Grenoble, France, ⁸Department of Biochemistry, University of Oxford, Dorothy Crowfoot Hodgkin Building, South Parks Road, Oxford OX1 3QU, UK, # Current address: Department of Biophysics, Medical School, University of Pécs, Szigeti Street 12, Pécs 7624, Hungary, § Current address: Center for Free-Electron Laser Science CFEL, Deutsches Elektronen-Synchrotron DESY, Notkestr. 85, 22607 Hamburg, Germany, sofia.kapetanaki@aok.pte.hu

Structural insights into the photoactivated adenylate cyclases can be used to develop new ways of controlling cellular cyclic adenosine monophosphate (cAMP) levels for optogenetic applications. OaPAC is a recently discovered blue-light using flavin adenosine dinucleotide (BLUF) photoactivated adenylate cyclase from the cyanobacterium *Oscillatoria acuminata* that uses adenosine triphosphate and translates the light signal into the production of cyclic adenosine monophosphate [1,2]. Here, we report the crystal structures of the enzyme in the absence of its natural substrate determined from room temperature serial crystallography data collected at both an X-ray free electron laser and a synchrotron and we compare them with the cryo macromolecular crystallography structures obtained at a synchrotron by us [3] and others [1,2,4]. These results reveal slight differences in the structure of the enzyme due to data collection at different temperatures and X-ray sources. We further investigate the effect of the Y6 mutation in the blue-light using flavin adenosine dinucleotide domain, a mutation which results in a rearrangement of the hydrogen-bond network around the flavin and a notable rotation of the side-chain of the critical Q48 residue [3]. These changes in the hydrogen bond network around the flavin triggered by the mutation point towards an allosteric regulation mechanism for the enzyme and are discussed in the light of the small and large-scale movements observed in the enzyme in solution upon ATP binding [5].

[1] Ohki, M., Sugiyama, K., Kawai, F., Tanaka, H., Nihei, Y., Unzai, S., Takebe, M., Matsunaga, S., Adachi, S., Shibayama, N., Zhou, Z., Koyama, R., Ikegaya, Y., Tame, J.R., Iseki, M. & Park, S.Y. (2016) *Proc. Natl. Acad. Sci.* **113**, 6659.

[2] Ohki, M., Sato-Tomita, A., Matsunaga, S., Iseki, M., Tame, J. R., Shibayama, N., & Park, S-Y. (2017) *Proc. Natl. Acad. Sci.* **114**, 8562.

[3] Kapetanaki, S.M., Coquelle, N., von Stetten, D., ..., Lukacs, A., Weik, M. (2024) BioRxiv doi: <https://doi.org/10.1101/2024.04.21.590439>.

[4] Chretien, A., Nagel, M.F., Botha, S., de Wijn, R., Brings, L., Dorner, K., Han, H., Koliyadu, J.C.P., Letrun, R., Round, A., Sato, T., Schmidt, C., Secareanu, R.-C., von Stetten, D., Vakili, M., Wrona, A., Bean, R., Mancuso, A., Schulz, J., Pearson, A.R., Kottke, T., Lorenzen, K. & Schubert, R. (2023) *J.Mol. Biol.* **436**, 168439.

[5] Ujfalusi-Pozsonyi, K., Bodis, E., Nyitrai, M., Kengyel, A., Telek, E., Pécsi, I., Fekete, Z., Varnyune Kis-Bicskei, N., Mas, C., Moussaoui, D., Pernot, P., Tully, M.D., Weik, M., Schiro, G., Kapetanaki, S.M. & Lukacs, A. (2024) *Commun. Biol.* **7**, 147.

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