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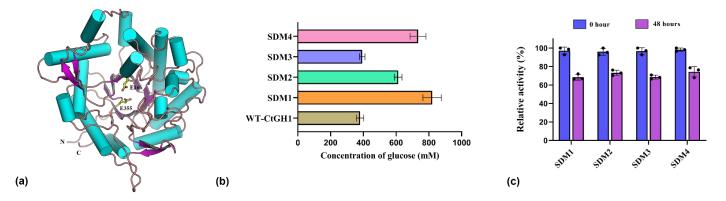
## Structure-guided improvement of glucose tolerance and catalytic efficiency of $\beta$ -glucosidase from *Clostridium thermocellum* for industrial bioethanol production

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Cellulases are utilized by bioethanol production industries for converting complex cellulose into fermentable sugars.  $\beta$ -glucosidase, constituting the last enzyme of the cellulose degradation pathway, is inhibited by its product – glucose [1]. This inhibition becomes a bottleneck of the industrial cellulose hydrolysis process. Hence, engineering  $\beta$ -glucosidases with high glucose tolerance along with high catalytic efficiency is a need of the hour in bioethanol production industries. Our current study encompasses the characterization of β-glucosidase from *Clostridium thermocellum* (WT-CtGH1). The purified WT-CtGH1 showed its higher specificity towards cellobiose over pNPG as a substrate. However, WT-CtGH1 showed a low glucose tolerance of 380 mM glucose compared to the desired industrial criterion of > 800 mM of glucose. Hence, we proceed with improving the glucose tolerance of WT-CtGH1. For this, the crystal structure of WT-CtGH1 elucidated by us at 3 Å resolution was used as a reference for the rational design approach (Figure 1a). A comprehensive understanding of the mechanism by which glucose binds to the active site pocket was obtained through sequence and structural analyses of WT-CtGH1 with other glucose-tolerant  $\beta$ -glucosidase enzymes [2 - 4]. These sequence and structure comparisons aided us in introducing two point mutations - SDM1 (Gly to Trp) and SDM2 (Ser to Trp) in the active site crater for improved glucose tolerance [5]. The purified SDM1 and SDM2 mutant enzymes showed enhanced glucose tolerance of ~ 800 mM and ~ 600 mM compared to WT-CtGH1 while retaining the kinetic properties (Figure 1b). Surprisingly, a point mutant - SDM3 (Ser to Leu) showed a more than two-fold increase in the catalytic efficiency (with cellobiose as compared with WT-CtGH1. A combinatorial double mutant of earlier mentioned mutations - SDM4 (Gly to Trp and Ser to Leu) showed a 1.5-fold increase in the catalytic turnover as well as the catalytic efficiency with cellobiose as a substrate along with the enhanced glucose tolerance of ~750 mM glucose (Figure 1b). Along with glucose tolerance, the temperature ( $45^{\circ}C - 60^{\circ}C$ ) and pH (4.8 -5.5) of the saccharification bioreactor limit the bioethanol production in separate hydrolysis and fermentation process (SHF). WT-CtGH1, along with all the above-mentioned mutants, showed considerable thermal stability at industrial operational conditions (Figure 1c). The mutant  $\beta$ - glucosidases developed in this research work would stand as potential candidates to be employed for the saccharification process in biofuel production, hence, have immense commercial value.



**Figure 1. (a)** Overall structure of WT-CtGH1 elucidated using X-ray crystallography, showing  $(\alpha/\beta)_8$  barrel fold with catalytic glutamates – E166 and E355; (b) Comparison of glucose tolerance of WT-CtGH1 with point mutants – SDM1, SDM2, SDM3 and a double mutant – SDM4; (c) Comparison of thermal stabilities of mutants – SDM1, SDM2, SDM3 and SDM4.

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