Oral presentation

Fine slicing or instant crystal growth: two different approaches to prepare macromolecular crystalline samples for 3D electron diffraction experiments

Christine Moriscot¹, Camille Feuiller², Eric Girard², Dominique HOUSSET²

¹Univ. Grenoble Alpes, CNRS, CEA, DISBG, IBS, F-38000 Grenoble, France ²Univ. Grenoble Alpes, CEA, CNRS, IBS, F-38000 Grenoble, France dominique.housset@ibs.fr

Electron diffraction of 3D nanometer sized crystals, most commonly named microED or 3D ED, has recently emerged as a new technique to solve the structure of both small organic molecules and proteins [1-3]. 3D ED is clearly a promising technique in structural biology, both quite easy to implement and very complementary to X-ray crystallography and single particle cryo-EM. Electrons are actually a very interesting probe for small samples as they strongly interact with matter, and more importantly, they deposit much less energy than X-rays per diffracted particle [4]. Sub-atomic resolution data can routinely be collected for small organic compounds and data up to 0.9 Å resolution have been obtained from protein crystals [5]. However, the major bottleneck of the technique when applied to protein samples is to produce nano-crystals that do not exceed 200 to 300 nm in at least one dimension and to deposit them on a grid while keeping the minimum amount of solvent around them. Here, we present two possible approaches. One uses cryo-sectioning after high pressure freezing of dextran embedded protein crystals. 150 to 200 nm thick cryo-sections of hen egg white lysozyme tetragonal crystals were used for electron diffraction experiments. Complete diffraction data up to 2.9 Å resolution were collected and the lysozyme structure was solved by molecular replacement and refined against these data [6]. The second approach used minute made lysozyme nano-crystals. By mixing lysozyme dissolved in water with a crystallant solution containing NaCl and Tb-Xo4 [7], a milky suspension of nanometer to micrometer size crystals were obtained that proved perfectly appropriate for 3D ED studies. Complete diffraction data up to 3.2 Å resolution were collected on 5 nano-crystals and the lysozyme structure was solved by molecular replacement with the Tb-Xo4 compound clearly visible in the residual $\{F_{obs} - F_{calc}\}$ Coulomb potential map [8]. Despite the speed of crystal growth, these crystals appear perfectly suitable for structural investigations.

[1] M.T.B. Clabbers, E. van Genderen, W. Wan, E.L. Wiegers, T. Gruene, J.P. Abrahams, Protein structure determination by electron diffraction using a single three-dimensional nanocrystal, Acta Cryst D 73 (2017) 738–748.

[2] M. Gemmi, E. Mugnaioli, T.E. Gorelik, U. Kolb, L. Palatinus, P. Boullay, S. Hovmöller, J.P. Abrahams, 3D Electron Diffraction: The Nanocrystallography Revolution, ACS Cent Sci 5 (2019) 1315–1329.

[3] T. Gruene, J.T.C. Wennmacher, C. Zaubitzer, J.J. Holstein, J. Heidler, A. Fecteau-Lefebvre, S. De Carlo, E. Müller, K.N. Goldie, I. Regeni, T. Li, G. Santiso-Quinones, G. Steinfeld, S. Handschin, E. van Genderen, J.A. van Bokhoven, G.H. Clever, R. Pantelic, Rapid Structure Determination of Microcrystalline Molecular Compounds Using Electron Diffraction, Angew Chem Int Ed Engl 57 (2018) 16313–16317.

[4] R. Henderson, The potential and limitations of neutrons, electrons and X-rays for atomic resolution microscopy of unstained biological molecules, Q. Rev. Biophys. 28 (1995) 171–193.

[5] M.W. Martynowycz, M.T.B. Clabbers, J. Hattne, T. Gonen, Ab initio phasing macromolecular structures using electron-counted MicroED data, Nat Methods 19 (2022) 724–729.

[6] C. Moriscot, G. Schoehn, D. Housset, High pressure freezing and cryo-sectioning can be used for protein structure determination by electron diffraction, Ultramicroscopy 254 (2023) 113834.

[7] S. Engilberge, F. Riobé, S. Di Pietro, L. Lassalle, N. Coquelle, C.-A. Arnaud, D. Pitrat, J.-C. Mulatier, D. Madern, C. Breyton, O. Maury, E. Girard, Crystallophore: a versatile lanthanide complex for protein crystallography combining nucleating effects, phasing properties, and luminescence, Chem Sci 8 (2017) 5909–5917.

[8] C. Sauter, D. Housset, J. Orlans, R. de Wijn, K. Rollet, S. Rose, S. Basu, P. Benas, J. Perez, D. de Sanctis, O. Maury, E. Girard. The nucleating agent crystallophore induces instant protein crystallization. submitted to Crystal Growth & Design (2024)