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A conserved fold for fimbrial components revealed by the crystal structure of a putative fimbrial assembly protein (BT1062) from *Bacteroides thetaiotaomicron* at 2.2 Å resolution

BT1062 from *Bacteroides thetaiotaomicron* is a homolog of Mfa2 (PGN0288 or PG0179), which is a component of the minor fimbriae in *Porphyromonas gingivalis*. The crystal structure of BT1062 revealed a conserved fold that is widely adopted by fimbrial components.

1. Introduction

The Gram-negative anaerobic bacterium *Bacteroides thetaiotaomicron* is a predominant member of the mammalian intestinal microbiota. It is important for the study of the symbiotic relationship between bacteria and humans, as well as for its abilities to digest complex plant polysaccharides and host-derived polysaccharides (Xu *et al.*, 2003). It is also an opportunistic pathogen and can cause serious infections. Extracellular proteins are expected to be crucial for such functions in *B. thetaiotaomicron* and other gut microbes. Therefore, we initiated a project to characterize the structures of proteins that are specific to the gut environment from the bacterial secretome of human gut microbiota, in order to gain further insights into the molecular mechanisms of bacteria-host symbiosis as well as of bacterial pathogenesis.

Here, we report the 2.2 Å crystal structure of a putative fimbrial assembly protein BT1062 from *B. thetaiotaomicron*, which was determined using the high-throughput pipeline of the Joint Center for Structural Genomics (JCSG; Lesley *et al.*, 2002) as part of the National Institute of General Medical Sciences Protein Structure Initiative (PSI; <http://www.nigms.nih.gov/Initiatives/PSI/>). The *BT1062* gene of *B. thetaiotaomicron* encodes a predicted lipoprotein with a molecular weight of 36 535 Da (residues 1–317) and a calculated isoelectric point of 4.8.

2. Materials and methods

2.1. Protein production and crystallization

Clones were generated using the Polymerase Incomplete Primer Extension (PIPE) cloning method (Klock *et al.*, 2008). The gene encoding BT1062 (Swiss-Prot Q8A8V5) was amplified by polymerase chain reaction (PCR) from *B. thetaiotaomicron* VPI-5482 genomic DNA using *PfuTurbo* DNA polymerase (Stratagene) and I-PIPE (Insert) primers (forward primer 5'-tgtacttccaggcGCTTCATGCG-ACAGCTTAATGAAGACC-3', reverse primer 5'-aattaagtgcgtta-TTGATTCTCTGAATGCGATGCACC-3'; target sequence in upper case) that included sequences for the predicted 5' and 3' ends. The expression vector pSpeedET, which encodes an amino-terminal tobacco etch virus (TEV) protease-cleavable expression and purification tag (MGSDKIHHHHHHENLYFQ/G), was PCR-amplified with V-PIPE (Vector) primers (forward primer 5'-taacgcgacttaatt-actcgttaaacggctccagc-3', reverse primer 5'-gccctggagaatcaggtttcgatgtatgtatgtatgt-3'). V-PIPE and I-PIPE PCR products were mixed to anneal the amplified DNA fragments together. *Escherichia coli* GeneHogs (Invitrogen) competent cells were transformed with the I-PIPE/V-PIPE mixture and dispensed onto selective LB-agar plates.

Table 1

Summary of crystal parameters, data-collection and refinement statistics for BT1062 (PDB code 3g8).

Values in parentheses are for the highest resolution shell.

	λ_1 MAD-Se	λ_2 MAD-Se	λ_3 MAD-Se
Space group	$P4_{1}2_{1}2$		
Unit-cell parameters (Å)	$a = 106.9, c = 79.1$		
Data collection			
Wavelength (Å)	0.9793	0.9116	0.9792
Resolution range (Å)	29.7–2.2 (2.26–2.20)	29.6–2.2 (2.26–2.20)	29.7–2.2 (2.26–2.20)
No. of observations	98494	94479	94046
No. of reflections	23880	23862	23868
Completeness (%)	99.9 (99.9)	99.9 (100)	99.9 (99.8)
Mean $I/\sigma(I)$	10.8 (1.8)	11.6 (2.2)	10.5 (1.6)
R_{merge} on I^{\dagger}	0.11 (0.72)	0.10 (0.65)	0.11 (0.80)
R_{meas} on I^{\ddagger}	0.13 (0.82)	0.11 (0.75)	0.13 (0.93)
Model and refinement statistics			
Resolution range (Å)	29.2–2.2		
No. of reflections (total)	23827		
No. of reflections (test)	1220		
Completeness (%)	99.8		
Data set used in refinement	λ_2 MAD-Se		
Cutoff criterion	$ F > 0$		
$R_{\text{cryst}}^{\$}$	0.191		
$R_{\text{free}}^{\ddagger}$	0.229		
Stereochemical parameters			
Restraints (r.m.s. observed)			
Bond lengths (Å)	0.016		
Bond angles (°)	1.52		
Average isotropic B value †† (Å 2)	37.6		
ESU ‡‡ based on R_{free} value (Å)	0.17		
Protein residues/atoms	284/2321		
Solvent molecules	175		

$^{\dagger} R_{\text{merge}} = \sum_{hkl} \sum_i |I_i(hkl) - \langle I(hkl) \rangle| / \sum_{hkl} \sum_i I_i(hkl)$. $\ddagger R_{\text{meas}}$ is the redundancy-independent R_{merge} (Diederichs & Karplus, 1997; Weiss & Hilgenfeld, 1997). $\$ R_{\text{cryst}} = \sum_{hkl} ||F_{\text{obs}}| - |F_{\text{calc}}|| / \sum_{hkl} |F_{\text{obs}}|$, where F_{calc} and F_{obs} are the calculated and observed structure-factor amplitudes, respectively. $\ddagger R_{\text{free}}$ is the same as R_{cryst} but for 5% of the total reflections chosen at random and omitted from refinement. $\dagger\dagger$ This value represents the total B that includes TLS and residual B components. $\ddagger\ddagger$ Estimated overall coordinate error (Collaborative Computational Project, Number 4, 1994; Cruickshank, 1999).

The cloning junctions were confirmed by DNA sequencing. Using the PIPE method, the gene segment encoding residues Met1–Glu22 was deleted as it was predicted to code for a signal peptide at the start of the protein. Expression was performed in a selenomethionine-containing medium at 310 K with suppression of normal methionine synthesis. At the end of fermentation, lysozyme was added to the culture to a final concentration of 250 µg ml $^{-1}$ and the cells were harvested and frozen. After one freeze–thaw cycle, the cells were sonicated in lysis buffer [50 mM HEPES pH 8.0, 50 mM NaCl, 10 mM imidazole, 1 mM tris(2-carboxyethyl)phosphine–HCl (TCEP)] and the lysate was clarified by centrifugation at 32 500g for 30 min. The soluble fraction was passed over nickel-chelating resin (GE Healthcare) pre-equilibrated with lysis buffer, the resin was washed with wash buffer [50 mM HEPES pH 8.0, 300 mM NaCl, 40 mM imidazole, 10% (v/v) glycerol, 1 mM TCEP] and the protein was eluted with elution buffer [20 mM HEPES pH 8.0, 300 mM imidazole, 10% (v/v) glycerol, 1 mM TCEP]. The eluate was buffer-exchanged with TEV buffer (20 mM HEPES pH 8.0, 200 mM NaCl, 40 mM imidazole, 1 mM TCEP) using a PD-10 column (GE Healthcare) and incubated with 1 mg TEV protease per 15 mg of eluted protein. The protease-treated eluate was run over nickel-chelating resin (GE Healthcare) pre-equilibrated with HEPES crystallization buffer (20 mM HEPES pH 8.0, 200 mM NaCl, 40 mM imidazole, 1 mM TCEP) and the resin was washed with the same buffer. The flowthrough and wash fractions were combined and concentrated to 19.1 mg ml $^{-1}$ by centrifugal ultrafiltration (Millipore) for crystallization trials. BT1062 was crystallized by mixing 100 nl protein solution with 100 nl crystallization

solution above a 50 µl reservoir volume using the nanodroplet vapor-diffusion method (Santarsiero *et al.*, 2002) with standard JCSG crystallization protocols (Lesley *et al.*, 2002). The crystallization reagent consisted of 1.4 M sodium citrate, 0.1 M HEPES pH 7.5. A cube-shaped crystal of approximate dimensions 40 × 40 × 30 µm was harvested after 23 d at 277 K for data collection. Ethylene glycol was added to the crystal as a cryoprotectant to a final concentration of 10% (v/v). Initial screening for diffraction was carried out using the Stanford Automated Mounting (SAM) system (Cohen *et al.*, 2002) and an X-ray microsource at Stanford Synchrotron Radiation Lightsource (SSRL, Menlo Park, California, USA).

The oligomeric state of BT1062 in solution was determined using a 1 × 30 cm Superdex 200 column (GE Healthcare) coupled with miniDAWN static light-scattering (SEC/SLS) and Optilab differential refractive-index detectors (Wyatt Technology). The mobile phase consisted of 20 mM Tris–HCl pH 8.0, 150 mM NaCl and 0.02% (w/v) sodium azide. The molecular weight was calculated using ASTRA v.5.1.5 software (Wyatt Technology).

2.2. Data collection, structure solution and refinement

Multi-wavelength anomalous diffraction (MAD) data were collected on beamline 9-2 at the SSRL at wavelengths corresponding to the inflection (λ_1), high-energy remote (λ_2) and peak (λ_3) of a selenium MAD experiment. The data sets were collected at 100 K using an MAR CCD 325 detector. The MAD data were integrated and reduced using MOSFLM and scaled with SCALA. Selenium sites were located using SHELXD (Sheldrick, 2008) and refined using autoSHARP (mean figure of merit of 0.46 with ten selenium sites; Bricogne *et al.*, 2003). Phase refinement and automatic model building were performed with RESOLVE (Terwilliger, 2003). Model completion and refinement were performed with Coot (Emsley & Cowtan, 2004) and REFMAC (Winn *et al.*, 2003). The refinement included experimental phase restraints in the form of Hendrickson–Lattman coefficients and TLS refinement with one TLS group per chain. CCP4 programs were used for data conversion and other calculations (Collaborative Computational Project, Number 4, 1994). Data-processing and refinement statistics are summarized in Table 1.

2.3. Validation, deposition and figures

The quality of the crystal structure was analyzed using the JCSG Quality Control server, which verifies the stereochemical quality of the model using AutoDepInputTool (Yang *et al.*, 2004), MolProbity (Lovell *et al.*, 2003) and WHAT IF v.5.0 (Vriend, 1990), the agreement between the atomic model and the data using SFCHECK v.4.0 (Collaborative Computational Project, Number 4, 1994) and RESOLVE (Terwilliger, 2003), the protein sequence using ClustalW (Thompson *et al.*, 1994), the atomic occupancies using MOLEMAN2 (Kleywegt, 2000) and the consistency of NCS pairs. It also evaluates the differences in $R_{\text{cryst}}/R_{\text{free}}$, expected $R_{\text{free}}/R_{\text{cryst}}$ and maximum/minimum B values by parsing the refinement log file and PDB header. All molecular graphics were prepared with PyMOL (DeLano Scientific). Sequence alignments were rendered using TEXshade (Beitz, 2000).

3. Results and discussion

3.1. Sequence analysis and functional assignment

BT1062 is a member of a functionally uncharacterized protein family [Pfam PF08842 or DUF1812 (domain of unknown function family 1812)] consisting of ~80 *Bacteroidetes* proteins of around 300–

400 residues. Homologous proteins are abundant in *Bacteroidetes* genomes. For example, at least four paralogs are found in *B. thetaiotaomicron* VPI-5482 (BT1062, BT2657, BT4225 and BT4226; sequence identity of >20%), three in *Porphyromonas gingivalis* (PGN0185, PGN0288 and PGN0289) and eight in *B. fragilis* NCTC 9343 (BF1578, BF1851, BF1976, BF2185, BF2264, BF2871, BF3328 and BF4229). The genomic context is conserved for *BT1062* homologs, which involves a cluster of four associated genes: *BT1066*, *BT1065*, *BT1063* and *BT1062* (Fig. 1). *BT1062* and *BF2185* of *B. fragilis* have almost identical genomic environments. A cluster of genes *BT1062–BT1068*, which are likely to be an operon, all contain signal peptides and are predicted to encode lipoproteins (with the exception of *BT1064*). This putative operon may be under the control of *BT1069*, which encodes a putative transcription regulator. Downstream of the operon is a putative histidine kinase (*BT1058*).

BT1062 is homologous to *Mfa2* (PGN0288, also previously known as PG0179) of *P. gingivalis* strain ATCC 33277 (19% sequence identity; Fig. 2a). *Mfa2* co-transcribes with the minor fimbrial antigen (*mfa1*) and is involved in the assembly of *Mfa1* fimbriae (Chung *et al.*, 2000; Hasegawa *et al.*, 2009). *BT1063* is a remote homolog of *Mfa1* (PGN0287; 15% identity; Fig. 1), which is the structural subunit of *P. gingivalis* minor fimbriae (Yoshimura *et al.*, 2009). *P. gingivalis* also contains homologous proteins to *BT1064* and *BT1066* (PGN0128 and PGN0179; PGN0129 and PGN0178). *BT1065* matches the N-terminal domain of PGN0128, indicating that PGN0128 is a fusion product of *BT1065*-like and *BT1064*-like proteins. Therefore, the *BT1062–BT1068* genes are most likely to encode a fimbriae (or pili) system similar to that of the minor fimbriae of *P. gingivalis*, with *BT1062* being equivalent to *mfa2*. *P. gingivalis* has at least two types of fimbriae: major (long) fimbriae with *FimA* as the main structural subunit (Yoshimura *et al.*, 1984) and minor (short) *Mfa1* fimbriae (Hamada *et al.*, 1996; Park *et al.*, 2005). Fimbriae were also observed in strains of *B. thetaiotaomicron* and *B. fragilis* (Shinjo & Kiyoyama, 1984); however, the fimbriae-assembly machinery are currently uncharacterized at the molecular level. The similarity of the potential fimbriae proteins to those of *P. gingivalis* could suggest a similar fimbriae-assembly system in *B. thetaiotaomicron* and *B. fragilis*. The fimbriae in gut bacteria, such as *B. thetaiotaomicron* and *B. fragilis*, may be required for adhesion to host tissues (Pumbwe *et al.*, 2006),

formation of biofilms with other bacteria in the gut, or play other as yet unknown functional roles.

3.2. Overall structure

The selenomethionine derivative of *BT1062* (residues 23–317) with an N-terminal His tag was expressed in *E. coli* and purified by metal-affinity chromatography. The predicted N-terminal signal peptide (residues 1–22) was not included in the construct. The crystal structure of *BT1062* was determined in the tetragonal space group $P4_321$ at 2.2 Å resolution using the MAD method. The final *BT1062* model includes a monomer (residues 34–317; Fig. 2b), one ethylene glycol and 174 water molecules in the asymmetric unit. The Matthews coefficient (V_M ; Matthews, 1968) for *BT1062* is 3.25 Å³ Da^{−1} and the estimated solvent content is 62%. The Ramachandran plot produced by MolProbity shows that 96.8 and 100% of the residues are in the favored and allowed regions, respectively. *BT1062* is composed of 21 β-strands (β_1 – β_{21}), three α-helices (α_1 – α_3) and five β_{10} -helices. The total β-sheet, α-helical and β_{10} -helical contents are 43.0, 6 and 5.3%, respectively. *BT1062* is likely to exist as a monomer in solution, which is consistent with crystal-packing analysis and analytical size-exclusion chromatography.

3.3. Structural comparisons

The structure of *BT1062* consists of a tandem repeat of two domains: I (34–170) and II (171–317). Using individual domains, the DALI structural similarity search server (Holm & Sander, 1995) indicated that both domains have transthyretin-like (previously known as prealbumin-like) folds with seven core β-strands (A–G) arranged in two sheets (DAG and CBEF; Fig. 2). The top hit for domain I is the C-terminal transthyretin subdomain of the carboxy-peptidase D domain II (Aloy *et al.*, 2001; PDB code 1h8l; Z = 7.0, r.m.s.d. of 2.1 Å for 78 aligned C^α atoms, 13% sequence identity). The best match for domain II is human transthyretin (Karlsson & Sauer-Eriksson, 2007; PDB code 2qel; Z = 6.1, r.m.s.d. of 3.6 Å for 96 aligned C^α atoms, 7% sequence identity). For the entire structure, the minor pilin GBS52 of the Gram-positive bacterium *Streptococcus agalactiae* (Krishnan *et al.*, 2007) is among the top hits (fifth), with an r.m.s.d. of 5.7 Å for 145 aligned C^α atoms (PDB code 2pz4; Z = 4.1, 10%

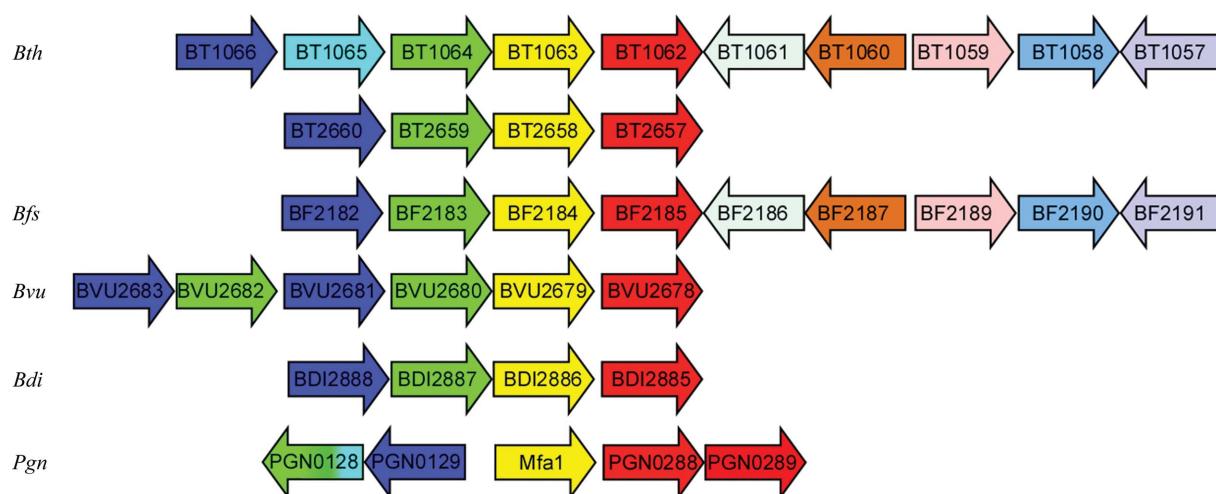
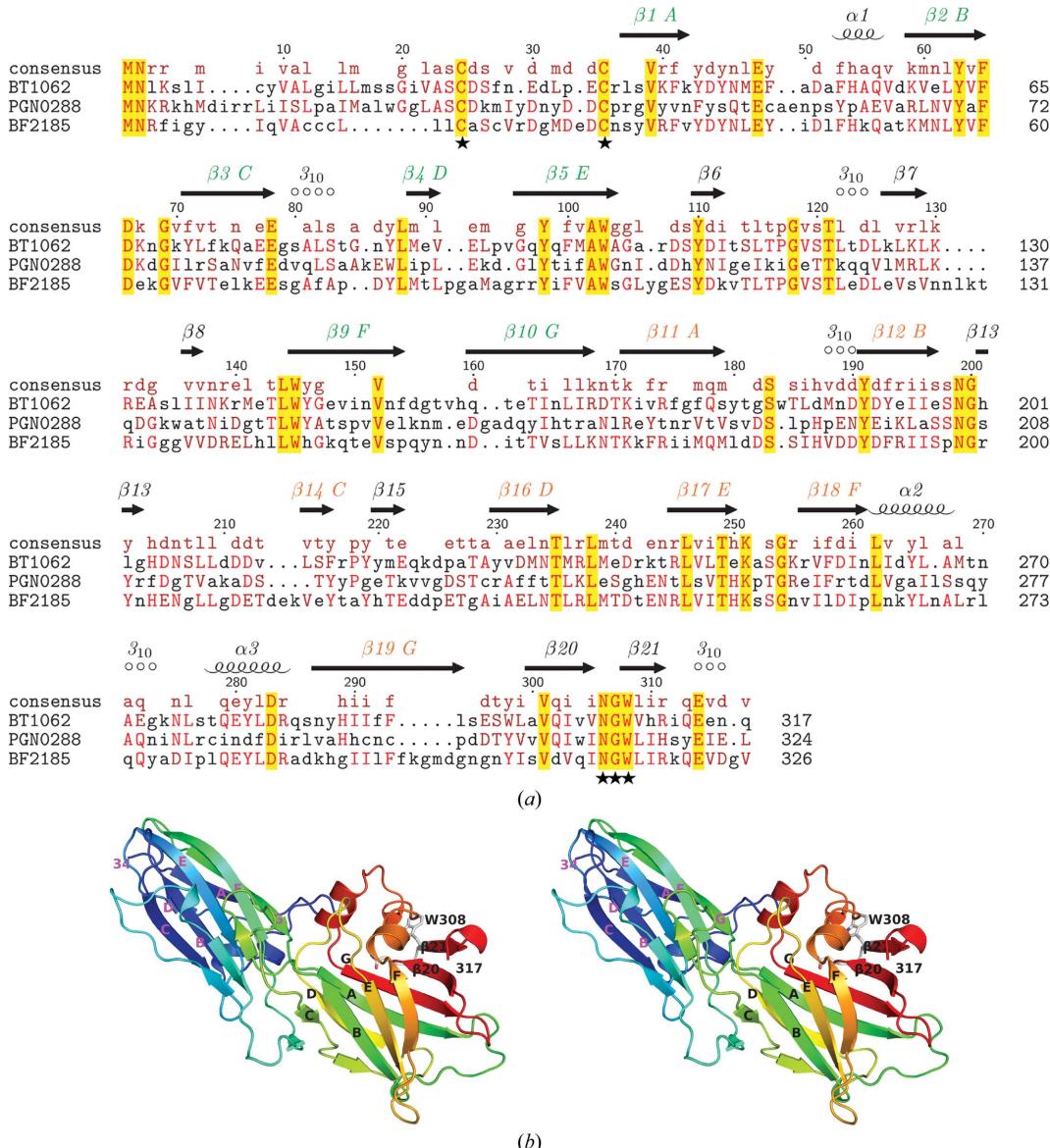


Figure 1

Gene context for representative *BT1062* homologs in *B. thetaiotaomicron* (*Bth*), *B. fragilis* NCTC 9343 (*Bfs*), *B. vulgatus* ATCC 8482 (*Bvu*), *Parabacteroides distasonis* ATCC 8503 (*Bdi*) and *Porphyromonas gingivalis* strain ATCC 33277 (*Pgn*). The lengths of the genes are not drawn to scale. Each homologous set of sequences is represented by one color.

**Figure 2**

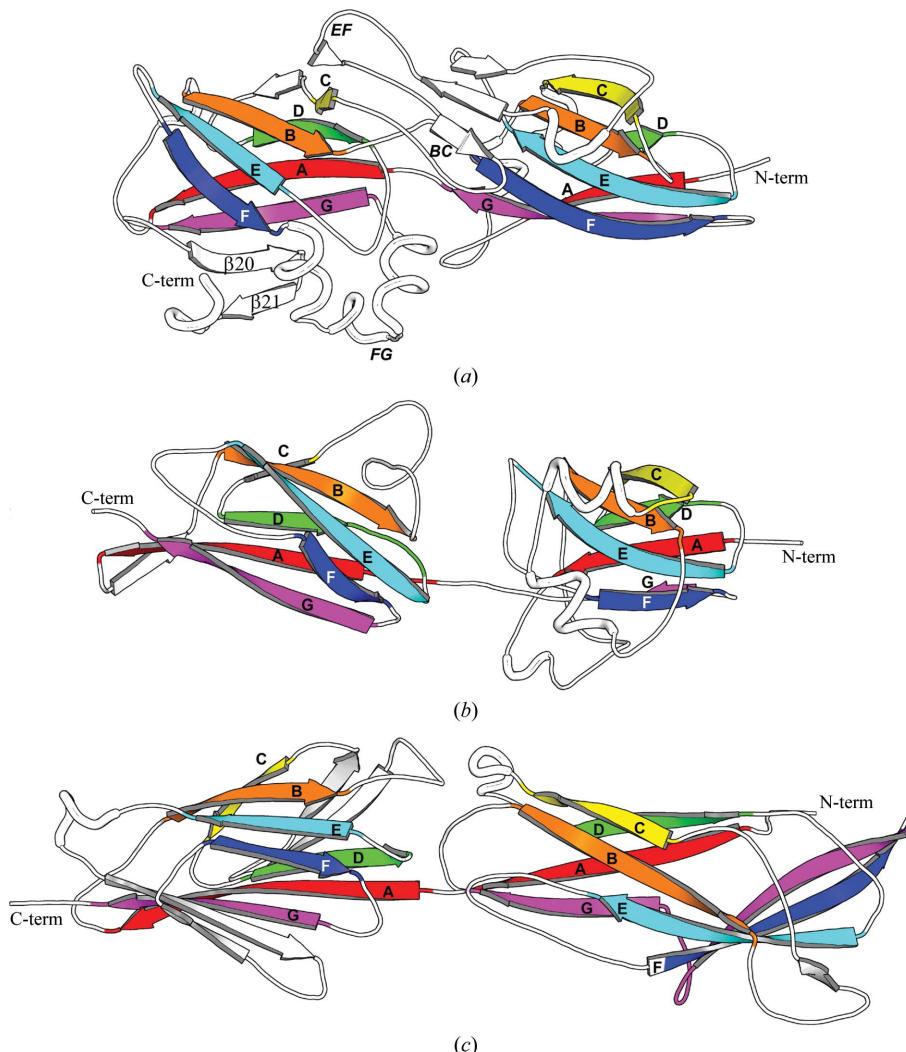
Crystal structure of BT1062. (a) Sequence alignment of BT1062, BF2185 and PGN0288 (Mfa2). The secondary-structural elements, residue numbering of BT1062 and consensus are shown at the top. The seven conserved β -strands (A–G) of the two transthyretin-like domains are highlighted. The potential membrane-attachment site and Mfa1-interaction site are labeled by stars at the bottom. (b) Stereo ribbon diagram of BT1062 monomer color coded from the N-terminus (blue) to the C-terminus (red).

sequence identity). *TM-align* (Zhang & Skolnick, 2005) aligned BT1062 to GBS52 with an r.m.s.d. of 4.8 Å for 175 C^α atoms. Despite the large r.m.s.d. value, this match is significant since both proteins are fimbrial components. The two domains of both proteins have an identical fold (*i.e.* the same topology of the seven core strands). GBS52 does not have long inserts between core β -strands in its two domains, except for the BC loop of the first domain, while BT1062 contains several significant insertions between core strands in both domains (Fig. 2). The most significant additional structural feature of BT1062 is a small β -sheet at the domain boundary formed by the EF loop of domain I and the BC loop of domain II (Fig. 3). Domain II contains a three-helix insertion between strands F and G as well as a β -hairpin attachment ($\beta 20$ – $\beta 21$) at the C-terminus. Thus, domain II of BT1062 deviates more significantly from the prototypical seven-stranded core domain, although some members of the transthyretin family have an additional β -strand that would correspond to $\beta 20$. A similar two-domain arrangement is also observed for the *S. pyogenes*

major pilin Spy0128 (*TM-align* r.m.s.d. of 5.3 Å for 178 aligned C^α atoms; Kang *et al.*, 2007; Fig. 3). Given the overall structural similarity and functional overlap, it seems possible that these pilin components might be derived from a common ancestral fold through divergent evolution. The basic fold of the seven core strands in these proteins has previously been described as IgG-like (Krishnan *et al.*, 2007). We have avoided such a description here owing to a lack of clear evidence to establish an evolutionary relationship between the IgG-like fold (SCOP ID 48725) and the transthyretin-like fold (SCOP ID 49451) (Andreeva *et al.*, 2004).

3.4. A conserved fold for fimbrial components

A sequence-similarity search using *PSI-BLAST* against the non-redundant (nr) database at the National Center for Biotechnology Information (NCBI) indicated that the family size of DUF1812 can be significantly expanded, with >1000 hits almost exclusively from

**Figure 3**

Structural comparisons of (a) BT1062, (b) the minor pilin of GBS52 (PDB code 2p24) and (c) the major pilin Spy0128 (PDB code 3b2m). All molecules are shown in a similar orientation with the same scale. The conserved core strands are labeled from A to G.

Bacteroidetes. There are 35 potential homologs from *B. thetaiotaomicron* alone, indicating the popularity of this fold in this bacterium. Interestingly, the identified homologs include components of both the major fimbriae and the minor fimbriae of *P. gingivalis*. In addition to the BT1062 homologs in minor fimbriae discussed above, major fimbrial components, such as FimA and the accessory proteins FimC, FimD and FimE, are expected to adopt a similar fold to BT1062. Thus, DUF1812 is a collection of diverse proteins that are likely to be fimbrial components. These proteins are likely to be adapted from a single fold to serve different functions. Many of these remote homologs also contain the highly conserved tryptophan (Trp308 in BT1062) described above.

The details of the biogenesis of Mfa1-like fimbriae are still unclear. A recent study suggested that *P. gingivalis* Mfa2 is likely to anchor the Mfa1 fimbriae to the outer membrane and to regulate the length of the Mfa1 filament (Hasegawa *et al.*, 2009). Mfa2 is present in the outer membrane and may directly interact with Mfa1. Most sequence homologs of BT1062 and Mfa2 contain two highly conserved cysteines at the N-terminus (Cys25 and Cys35) located near the tip of the bilobal molecule. The first invariant cysteine was predicted to be the lipoprotein signal-peptide cleavage site (between 24 and 25) by the LipoP server (Juncker *et al.*, 2003). This cysteine is likely to be the

last residue of the lipoprotein signal-sequence motif [lipobox motif (L/V)XXC, X = A/S/G/T] and is directly involved in membrane attachment of the matured lipoprotein via a thioether bond (Braun & Wu, 1994). The role of the second conserved cysteine is currently not clear. It may also be involved in membrane attachment owing to its close proximity to the first cysteine. The conformation of the peptide between the two conserved cysteines is likely to be flexible since residues 24–33 are exposed to solvent and disordered in the crystal with no interpretable electron density. The most conserved surface residues of BT1062 homologs correspond to a short sequence motif $^{306}\text{N}(\text{G/D})\text{W}^{308}$ located in the $\beta 20-\beta 21$ loop. This exposed site is likely to be involved in interaction with Mfa1 and thus to be important for the function of BT1062. The spatial arrangement of the potential membrane-attachment site and protein–protein interaction site may be functionally significant as the location of the potential membrane-attachment site would allow more freedom and accessibility of a membrane-attached elongated molecule.

4. Conclusions

Bioinformatics studies, combined with experimental evidence from the related bacteria *P. gingivalis*, allow us to identify at least one

putative operon that is likely to be involved in fimbrial assembly in *B. thetaiotaomicron* and other related bacteria. Our structural studies of the BT1062 protein in this operon revealed surprising structural similarities to the minor pilin GBSS52 of *S. agalactiae* and the major pilin Spy0128 of *S. pyogenes*, both of which are Gram-positive bacteria. We demonstrated that a tandem repeat of the transthyretin-like fold is also likely to be adopted by other components of *Bacteroides* fimbriae, such as the major pili subunit FimA of *P. gingivalis*. These results may suggest a common evolutionary origin for this type of fimbrial component in both Gram-negative and Gram-positive bacteria. Thus, our studies contribute new insights into the evolution of fimbriae (pili).

Additional information about BT1062 is available from TOPSAN (Krishna *et al.*, 2010) at <http://www.topsan.org/explore?pdbID=3gf8>.

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