topical reviews



IUCrJ ISSN 2052-2525 NEUTRON SYNCHROTRON

Received 27 September 2024 Accepted 19 February 2025

Edited by I. Robinson, UCL, United Kingdom

Keywords: coherent X-ray diffraction imaging; CXDI; materials science; nanoscience; X-ray microscopy; synchrotron radiation.



Prospects for coherent X-ray diffraction imaging at fourth-generation synchrotron sources

Yuriy Chushkin* and Federico Zontone

ESRF – The European Synchrotron, 71 avenue des Martyrs, 38000 Grenoble, France. *Correspondence e-mail: chushkin@esrf.fr

Coherent X-ray diffraction imaging is a lens-less microscopy technique that emerged with the advent of third-generation synchrotrons, modern detectors and computers. It can image isolated micrometre-sized objects with a spatial resolution of a few nanometres. The method is based on the inversion of the speckle pattern in the far field produced by the scattering from the object under coherent illumination. The retrieval of the missing phase is performed using an iterative algorithm that numerically phases the amplitudes from the intensities of speckles measured with sufficient oversampling. Two- and three-dimensional imaging is obtained by simple inverse Fourier transform. This lens-less imaging technique has been applied to various specimens for their structural characterization on the nanoscale. Here, we review the theoretical and experimental elements of the technique, its achievements, and its limitations at thirdgeneration synchrotrons. We also discuss the new opportunities offered by modern fourth-generation synchrotrons and outline the developments necessary to maximize the potential of the technique.

1. Introduction

There has been growing interest in using X-rays as a probe for high-resolution imaging. According to the Abbe resolution criterion, the short wavelength of X-rays enables high resolving power, allowing them to penetrate the internal structure of thick optically opaque specimens. Thanks to these properties, X-ray imaging is used intensively in many fields of applications and is being constantly improved and developed (Baruchel et al., 2008; Jacobsen, 2019). In contrast to visible light microscopy, the use of lenses for image formation in the X-ray regime is limited by the challenges of manufacturing high-quality lenses and their resulting low efficiency (Chao et al., 2005). The advent of powerful X-ray sources such as synchrotrons and free-electron lasers paved the way for the emergence of lens-less imaging techniques that use X-ray beams with a high degree of coherence. In particular, coherent X-ray diffraction imaging (CXDI), demonstrated in 1999 (Miao et al., 1999), enabled innovative high-resolution X-ray imaging modalities capable of achieving nanometre-scale resolution without the need for lenses. With this method, an image of a specimen is recovered using numerical algorithms applied to its diffraction pattern.

Modern detectors, the development of phase-retrieval algorithms and the boosting of computational power laid the foundations for developing new coherent X-ray imaging techniques (Chapman & Nugent, 2010; Cloetens *et al.*, 1999). This led to the implementation of two classes of X-ray microscopy. The first recovers an image from diffraction patterns recorded in the near-field (Fresnel diffraction) regime, with notable techniques including in-line holography

(Cloetens et al., 1999; Mokso et al., 2007; Soltau et al., 2021; Kalbfleisch et al., 2022), speckle tracking imaging (Bérujon et al., 2012) and near-field ptychography (Stockmar et al., 2013). The resolution of the obtained images is given by the detector pixel size and/or source size. The second class of techniques uses far-field diffraction patterns (Fraunhofer diffraction) to recover an image at the resolution given by the largest scattering angle where the diffraction signal is observed. Among the implemented methods are CXDI in forward-scattering geometry for microscopy of isolated non-crystalline specimens (Miao et al., 1999; Chapman et al., 2006), CXDI in Bragg geometry for imaging strain fields in nanocrystals (Pfeifer et al., 2006; Robinson & Harder, 2009; Sun & Singer, 2024) and their shapes (Robinson et al., 2001), Fourier transform holography (Stroke & Falconer, 1964; McNulty et al., 1992; Eisebitt et al., 2004) and scanning coherent imaging via far-field ptychography to visualize extended specimens (Rodenburg et al., 2007; Thibault et al., 2008; Dierolf et al., 2010; Pfeiffer, 2018). Owing to the robustness of the ptychographic reconstruction algorithm, ptychography became a leading technique for 3D quantitative high-resolution imaging (Holler et al., 2017; Aidukas et al., 2024).

Clearly, all the above-mentioned imaging techniques can gain a lot from the upgrades of synchrotron sources that have been accomplished recently or that are currently underway worldwide. In this paper, we focus on CXDI in forward-scattering geometry. The development of CXDI has been driven by the prospect of single-particle imaging of biomolecules at X-ray free-electron laser (XFEL) facilities (Neutze *et al.*, 2000). At synchrotrons, CXDI has found its utility for 3D imaging of isolated microscopic specimens, and its capabilities can be further enhanced at fourth-generation synchrotron sources.

2. Numerical lens

Classical microscopy schemes use lenses for magnified image formation. The manufacture of X-ray lenses is technically challenging and even the best quality lenses still can suffer from low efficiency (Chao *et al.*, 2005; Bajt *et al.*, 2018). CXDI operates without objective lenses. Fig. 1(*a*) shows the standard experimental geometry for plane-wave CXDI. An isolated specimen is placed on a thin Si_3N_4 membrane (transparent to X-rays) and can be rotated about the vertical axis for tomographic measurements. The microscopic specimen is illuminated by a monochromatic X-ray beam with a high degree of coherence, typically twice as large as the specimen's size.

The resulting far-field diffraction pattern $I(\mathbf{q})$ (\mathbf{q} is the scattering vector) from an isolated sample contains high- and low-intensity interference modulations called speckles. The speckle pattern encodes information on the exact spatial distribution of the electron density f in the sample. Within the Born approximation, the Fourier transform \mathcal{F} of the projected two-dimensional electron distribution is proportional to the scattered electromagnetic wavefield F at the detector plane in the far-field regime. Unfortunately, only the square amplitude

of the wavefield is measured and the phase ϕ information, essential for the inversion, is lost. It was proposed (Sayre, 1952; Miao *et al.*, 1998) and demonstrated (Miao *et al.*, 1999) that the missing phase can be retrieved by sampling the diffraction pattern at a resolution twice higher than the Nyquist frequency.

Numerical iterative phase retrieval (IPR) algorithms have been designed (Gerchberg & Saxton, 1972; Fienup, 1987; Luke, 2004) to retrieve the missing phase and hence obtain an image of the sample via the inverse Fourier transform \mathcal{F}^{-1} [Fig. 1(b)]. In this respect, the iterative algorithm can be regarded as a numerical lens. The algorithm uses the measured amplitudes $\sqrt{I(\mathbf{q})}$ to replace the Fourier amplitudes in reciprocal space and support constraints (Fienup, 1987) in real space. Generally, the iterative reconstruction starts from a random initial guess of an object and its support. The support is the finite and compact region in real space with non-zero electron density. During the iterations, the initial support can be gradually refined; for example, by using the shrink-wrap algorithm (Marchesini et al., 2003). Usually, a few thousand iterations are sufficient to achieve convergence. Typically, tens of reconstruction attempts are performed each time, starting from a random guess of the object. Only the results with the lowest error metric are retained for averaging to obtain the final reconstruction (Favre-Nicolin et al., 2020b).

There are alternative approaches for final image formation; for example, one approach uses the standard averaging method combined with an optimization procedure (Chen *et al.*, 2007). Algorithms first developed for 2D imaging can be applied to 3D data sets using a 3D fast Fourier transform (FFT). High-performance software is available for this



Figure 1

(a) Experimental geometry of CXDI. (b) Iterative phase retrieval algorithm scheme – numerical lens.

purpose (Chapman *et al.*, 2006; Favre-Nicolin *et al.*, 2020*a*). In 3D CXDI, the 3D diffraction volume is assembled from 2D diffraction patterns recorded at different sample tilts ω , taking into account the Ewald sphere curvature, thus overcoming the depth-of-focus limitations encountered in thick objects (Chapman *et al.*, 2006).

Although the IPR is a key element in CXDI, its ability to converge to an optimal solution depends on the quality of the measured diffraction patterns. Two major criteria determine the quality of the data. Firstly, the linear sampling ratio of the data σ must be $\sigma > 2^{1/N}$, where N is the space dimension (Miao et al., 1998); that is, the number of correlated intensities must be larger than the number of unknown variables. The experimental reality dictates that σ must be 3 or higher to ensure convergence of the IPR and reduce image artifacts (Song et al., 2007). Secondly, the quantity of missing data [white areas on the detector, Fig. 1(a)], in particular the number of speckles in proximity to the direct beam, must be reduced to a fraction of the central speckle (Miao et al., 2005). The problem in CXDI is that the direct beam must be blocked, as current detectors do not have enough dynamic range to cope with the intensity of the direct beam. Therefore, a beamstop is always used to block the direct beam, which necessarily masks the central speckles to a certain extent. Strategies to address this issue include the use of a semitransparent beamstop (Wilke et al., 2013) or the numerical mitigation of low-frequency information loss through a constrained power operator (Thibault et al., 2006).

The challenge of missing data becomes more relevant for 3D CXDI. In this case, the object is rotated about one axis to collect a series of diffraction data at different object tilts ω [Fig. 1(a)]. The available angles for the tomographic scan are usually limited to $\pm 80^{\circ}$ at best, since the object is sitting on a support (e.g. an Si_3N_4 membrane) that can cast a shadow on the beam and leave an unmeasured angular interval called the missing wedge. The issue of the missing wedge becomes more pronounced when large detectors are used, as the number of pixels with unknown intensity increases cubically with the detector size. In addition, large pixel array detectors have gaps between sensor modules, which cause additional artifacts in the reconstructed images (Carnis et al., 2019; Masto et al., 2024). This issue can be overcome to some degree by postprocessing treatments (Thibault et al., 2006; Malm, 2021; Masto et al., 2024; Malm & Chushkin, 2025).

The IPR works as a numerical lens and, with a sufficiently large collection angle and signal-to-noise ratio, it can outperform the best X-ray lenses (De Andrade *et al.*, 2021) in terms of dose efficiency and resolution. In CXDI, the Abbe criterion translates into a resolution limit given by the largest scattering angles (highest spatial frequencies) where the speckles are recorded with a sufficiently good signal-to-noise ratio, and it is ultimately limited by the detector size (Skjønsfjell *et al.*, 2018*b*). Since CXDI uses plane-wave illumination, small sample translations do not impact the resolution, making reconstructions resilient to mechanical instabilities. Resolution of the order of tens of nanometres has been demonstrated in several studies (Miao *et al.*, 2003; Takahashi et al., 2010; Vila-Comamala et al., 2011; Schropp et al., 2012).

3. Applications

Various X-ray methods have been used for imaging biological objects (Dierolf et al., 2010; Bartels et al., 2012; Nishino et al., 2012; Jacobsen, 2019). Growing interest in developing CXDI was triggered by its potential for high-resolution microscopy of biological specimens. In comparison, the application of electron microscopy commonly requires laborious sample preparation, often involving chemical staining to enhance contrast in biological specimens, in order to obtain images of thin sections with a resolution of a few ångströms (McIntosh, 2007). Here, CXDI promises to outperform previous techniques, thanks to extremely high phase sensitivity, even for specimens in a natural state. An example of Deinococcus radiodurans bacteria, imaged with CXDI using 8 keV radiation, is shown in Fig. 2(a). Unstained cells were deposited on an Si₃N₄ membrane, with alcohol replacing water to fix the cells. The resolution element (pixel) in these pictures is \sim 32 nm and, thanks to the high sensitivity, it is possible to distinguish the cell envelope, DNA toroids, septa and dense grains without chemical staining.

There are numerous examples of CXDI being applied to biological imaging. Early work by Jiang *et al.* (2008) demonstrated the application of CXDI for imaging biominerals in



Figure 2

CXDI images. (a) A 2D projected image of *Deinococcus radiodurans* bactaria. (b) Three-dimensional renderings of (top) fused and individual Si nanocrystals and (bottom) central slices of a large cluster. Scale bars are 1 μ m.

fish bone and in bovine bones (Verezhak, 2016) at nanometre resolution. Subsequent imaging of a whole unstained Schizosaccharomyces pombe yeast spore cell using chemical fixation (Jiang et al., 2010) revealed the 3D morphology and structure of cell organelles at 50-60 nm resolution. The cell structure can be preserved in a close-to-natural state using cryogenic freezing. Freeze-plunging is widely used in X-ray crystallography, and in electron and X-ray microscopy. For example, it was applied for imaging frozen-hydrated D. radiodurans bacteria (Lima et al., 2009), a Neospora caninum cell (Rodriguez et al., 2015) and near-surface structures of frozenhydrated malaria-infected human erythrocytes (Frank et al., 2017). Cryogenic cooling is also a standard approach for mitigating radiation damage of biological specimens, which is a major limitation for achieving high resolution. This damage can result in mass loss, as demonstrated in metal-coated polymeric spheres (Skjønsfjell et al., 2018a). Extrapolating from experimental data and theoretical calculations, Howells and co-workers estimated that biological imaging at 10 nm resolution should be achievable using cryogenic cooling (Howells et al., 2009), yet it still remains to be demonstrated.

The unique capabilities of CXDI are particularly well suited for imaging materials science specimens, which are generally more resistant to radiation damage. Fig. 2(b) shows CXDI images of a cluster of Si nanoparticles deposited on an Si₃N₄ membrane. The cluster contains individual octahedral nanoparticles, each ~200 nm in size, present on both the surface of the membrane and within the cluster. The central large cluster has a porous internal morphology, which makes such materials interesting for optoelectronics applications (de Jong *et al.*, 2016). CXDI has also been used to investigate TiO₂ sponges infiltrated with a perovskite for photovoltaic applications (Sanzaro *et al.*, 2019).

CXDI has also been used to study complex biomineral architectures, such as the exoskeleton structure of coccolithophores, known as coccospheres (Beuvier *et al.*, 2019). A coccosphere consists of an assembly of imbricated coccoliths, composed of calcite crystals produced by the cells. The strong scattering power of the CaCO₃ specimens, along with their robustness to X-rays, enabled high-quality 3D imaging. This revealed correlations between coccolith mass and grid size, as well as size variability within a coccosphere (Beuvier *et al.*, 2019).

 $CaCO_3$, a widespread product of biomineralization by living organisms, is ubiquitous in nature. As a light and biocompatible substance, it has found numerous industrial applications and can be used as a model system to study biomineralization and non-classical nucleation and growth mechanisms (De Yoreo *et al.*, 2015). CaCO₃ appears in three polymorph crystalline phases: calcite, aragonite and vaterite, all of which can be synthesized in a laboratory under specific conditions of temperature, concentration, additive presence and time. CXDI is an indispensable tool to visualize the 3D internal porosity of CaCO₃ microparticles. The method was used to follow the modification of pore geometry during the solidstate transformation of vaterite spherical particles to calcite (Cherkas et al., 2017) during high-temperature annealing. Another study investigated the phase-transformation kinetics of dried vaterite particles immersed in deionized and tap water (Cherkas et al., 2018). Recently, a comprehensive study uncovered the mechanism that governs the formation of hollow vaterite microparticles (Beuvier et al., 2022). The particles were synthesized by varying the ion concentration and the amount of an additive (polystyrene sulfonate, PSS) that slows down the kinetics of the process. The specimens were collected as a function of reaction time to follow the time evolution of the particle morphology. The work showed that Ostwald ripening cannot explain the observed diversity of crystal sizes, shapes and spatial organization. For this, other factors, such as PSS concentration and dipole-dipole interactions, must be accounted for in order to explain the transformation from a whole to a hollow particle (Beuvier et al., 2022). In order to resolve the crystalline phases of $CaCO_3$ or other single-particle minerals (Chattopadhyay et al., 2020), combined CXDI and Bragg diffraction measurements were developed (Chushkin et al., 2019).

These selected examples demonstrate the power of CXDI at third-generation synchrotrons. Nevertheless, new-generation synchrotron sources are expected to boost the capabilities of the method significantly.

4. Fourth-generation synchrotron sources

Fourth-generation synchrotron sources are based on the multi-bend achromat (MBA) lattice design (Einfeld *et al.*, 1995), which reduces the horizontal emittance by more than an order of magnitude, producing X-ray beams with a high degree of coherence (Raimondi *et al.*, 2023). The MBA lattice has been exploited to construct new synchrotron storage rings, such as Max IV and Sirius, but it can also be efficiently implemented at third-generation synchrotrons through the hybrid MBA design (Biasci *et al.*, 2014; ESRF, 2014), as demonstrated by the ESRF's Extremely Brilliant Source (ESRF-EBS) (Raimondi *et al.*, 2023). This approach paves the way for upgrades of existing third-generation synchrotrons worldwide.

High brilliance and low emittance are essential for coherent scattering techniques, as transverse coherence length scales inversely with emittance, while coherent flux is proportional to brilliance. For CXDI, such properties result in a larger field of view (object size), faster data collection and higher image resolution. The large spatial coherence enables the imaging of larger objects, in particular biological cells and materials of approximately 10-20 µm in size. Generally, biological cells are studied either with conventional optical microscopy, which can provide images of transparent specimens at a resolution of hundreds of nanometres (250 nm in Fig. 3), or with electron microscopy, which provides atomic scale resolution but only of thin sections a few hundred nanometres (250 nm) thick (Fig. 3). X-ray microscopy, in particular CXDI, at highly brilliant sources bridges the gap between these two techniques by imaging specimens a few micrometres thick at nanometre resolution.

topical reviews



Sample size and resolution coverage of CXDI, electron and optical microscopy techniques. The gray vertical line is the 10 nm resolution limit and the black dashed line defines the DOF boundary at $\lambda = 1.55$ Å.

Fig. 3 summarizes the expanded capabilities of CXDI with a fourth-generation synchrotron source. The red area delineates the accessible region at third-generation sources. The maximum sample size S is limited by the transverse coherence length and available sample-to-detector distance D, which follows the linear sampling ratio $\sigma = D\lambda/pS = 3$, where λ is the wavelength and p is the detector pixel size. The slope of the region defining the resolution element Δr is given by the sample size S and the detector size in pixels N_{pix} , assuming sufficient signal is recorded at the highest frequencies and maintaining a fixed sampling ratio of 3, giving $\Delta r = 3 \times S/N_{\text{pix}}$. Following the assumption of Porod's law, that the average speckle intensity $I(\mathbf{q})$ falls as $I(\mathbf{q}) \simeq \mathbf{q}^{-4}$, a hundredfold increase in brilliance results in a resolution gain of about $100^{1/4}$ \simeq 3 for the same signal-to-noise ratio. The improvement expected with fourth-generation sources and a detector four times larger is represented by the green region in Fig. 3. For our calculations, we assume a detector with 2048×2048 pixels and a pixel size $p = 75 \,\mu\text{m}$ positioned 30 m from a sample illuminated by a coherent 8 keV plane wave. Under such conditions, imaging samples up to 20 µm in size at (sub-) 30 nm resolution should make CXDI even more attractive for biologists, as many interesting biological specimens fall within this size range.

We note that samples made of radiation-hard materials that are smaller than 7 µm can be routinely imaged at sub-10 nm resolution. As an example, Fig. 4 shows the capability of the new ESRF-EBS coupled with a large detector to provide a resolution of below 10 nm of a test logo sample patterned in a 216 nm thin tungsten layer. It is important to note that the sample thickness is a parameter that can constrain the resolution of an image. When the sample thickness exceeds the depth of field (DOF), given by DOF = $2\Delta r^2/\lambda$, the resulting 2D image exhibits defocus artifacts (Chapman *et al.*, 2006). The DOF boundary, defined for a wavelength of $\lambda = 1.55$ Å, is shown in Fig. 3 and is more restrictive than the detector size constraints (green area). Using shorter wavelengths shifts the





(Top row) A 5 μ m ESRF logo test sample imaged by (left) SEM, (middle) CXDI before the EBS at 32.5 nm pixel size and (right) with the EBS at 7.6 nm pixel size. (Bottom row) Enlargements of the letter 'R' demonstrate the increase in resolution.

boundary towards higher resolution, but it also reduces the sample size/thickness for which oversampling can be achieved within the available sample-to-detector distance. Never-theless, alternative approaches can be applied. In ptycho-graphy, the defocus artifacts in 2D imaging can potentially be solved numerically using the multi-slice method (Maiden *et al.*, 2012; Tsai *et al.*, 2016). In CXDI, the issue is resolved by taking the Ewald sphere curvature into account when building the 3D diffraction volume (Chapman *et al.*, 2006) or a 2D slice (Takahashi *et al.*, 2010) from the 2D diffraction patterns measured at different sample tilts. It is important to note that other X-ray imaging techniques can cover certain portions of the sample size domain shown in Fig. 3, but CXDI holds the potential to achieve the finest resolution.

Coherent X-rays can also be used for imaging magnetic nanostructures and their dynamics with soft X-rays (Eisebitt *et al.*, 2004; Flewett *et al.*, 2012; Ukleev *et al.*, 2018) and, recently, with hard X-rays (Donnelly *et al.*, 2017) to probe magnetic domains, domain walls, vortexes and skyrmions. Hard X-rays, in the range of 6–12 keV, are necessary for imaging microscopic specimens that are too thick (>400 nm) for soft X-rays, but magnetic contrast in this energy range is rather low (Donnelly *et al.*, 2016). Thus, the high coherent flux provided by fourth-generation sources is mandatory for exploiting hard X-rays to characterize magnetic textures in materials (Donnelly & Scagnoli, 2020).

One of the little-exploited but high-potential applications of CXDI is time-resolved imaging to study dynamic phenomena (Grote *et al.*, 2022). Due to its non-scanning nature, fast recording (kilohertz frame rate) of diffraction data is possible with current state-of-the-art pixel array detectors. Plane-wave illumination, used in CXDI, provides robustness to any unwanted sample nano-translations and ensures high resolution. Real-time imaging of dynamic phenomena was demonstrated in a proof-of-principle experiment using a coherent optical light source (Lo *et al.*, 2018). In the hard X-ray regime,

the combination of X-ray photon correlation spectroscopy with CXDI for time-resolved imaging was recently proposed (Takazawa et al., 2023; Hinsley et al., 2024). Rapid data collection, combined with analysis of photon-sparse random projections (Ayyer et al., 2016; Giewekemeyer et al., 2019), can be useful in applications with optical trapping and manipulation (Gao et al., 2019) or levitating specimens. Sample levitation/manipulation in air or liquid (Kolb et al., 2015; Müller et al., 2015) could be exploited for random sample rotations in order to generate sufficient data for a complete 3D diffraction volume without the missing wedge. The challenge of constructing a 3D diffraction volume with sufficient statistics from random sample orientations is central in single-particle imaging at XFELs, and a solution has been developed by Loh & Elser (2009). Recent studies have demonstrated significant progress by reconstructing 3D structures of viruses (Rose et al., 2018) and nanoparticles (Ayyer et al., 2021; Nakano et al., 2022) from a series of single-shot diffraction patterns at an XFEL. The proposed solutions for data sorting, background subtraction, sample handling and image reconstruction can be applied directly to synchrotron CXDI imaging. For example, the expand-maximize-compress algorithm (Loh & Elser, 2009) for assembling 3D diffraction volumes from random sample orientations was adopted for Bragg CXDI (Björling et al., 2020) to manage uncontrolled particle rotations during measurements.

To capitalize fully on the new possibilities offered by fourthgeneration synchrotron sources, specialized instruments (Chushkin *et al.*, 2014; Johansson *et al.*, 2021) must be developed, alongside advancements in detectors (Grimes *et al.*, 2023; Takahashi *et al.*, 2023), to enhance detection efficiency for both high- and low-intensity signals.

Further development of CXDI should focus on improving the performance of the numerical lens. Efforts should be directed towards improving algorithm robustness to noise, handling sparse data sets, reducing artifacts caused by missing information and overcoming depth-of-focus limitation in thick samples to achieve the ultimate resolution. For example, the new general proximal smoothing algorithm (Pham *et al.*, 2019) outperforms classical algorithms when phasing noisy data. The latest phase retrieval developments incorporate the total variation regularization to compensate for missing data (Chang *et al.*, 2016) and demonstrate efficiency in imaging magnetic domains (Yokoyama *et al.*, 2022).

A promising approach is to employ machine learning methods (Harder, 2021). There are numerous examples of the effectiveness of machine learning for IPR. The introduction of data treatments based on neural networks is useful for recovering missing data and reducing noise (Bellisario *et al.*, 2022), as well as for removing artifacts caused by the detector gaps (Masto *et al.*, 2024). Deep neural network models are also effective for complex single-particle imaging (Wu *et al.*, 2021). Additionally, machine learning can be used to find reliable object support (Shang *et al.*, 2025). New approaches for resolution enhancement via the extrapolation of experimental data could also be effective in some cases (Latychevskaia *et al.*, 2015).

5. Conclusions

After 25 years of existence, coherent X-ray imaging (CXDI) has proved to be a very powerful method for imaging biological and materials science specimens on the nanoscale. A wide range of objects, from biological cells, bacteria and bones to the exoskeletons of algae, minerals, mesostructures and porous particles for electronics have been scrutinized using CXDI.

At third-generation synchrotrons, CXDI has occupied a niche for visualizing the 3D structure of naturally microscopic and isolated specimens. Typically, the largest size of the investigated objects was limited to \sim 6–7 µm, due to the small transverse coherence length of the beam, and with a resolution of around 15–30 nm, due to the size of the detector and the coherent flux. The new generation of synchrotron sources with highly coherent X-ray beams will enhance the capability of CXDI to image 10–20 µm specimens with even greater detail and in a shorter time. Such improvements result in a better tool for visualizing cells, bone fragments, exoskeletons of marine algae and micrometre-sized nanostructures, as well as enhanced capabilities for time-resolved studies of the formation of crystalline microparticles, biominerals *etc.*

To benefit fully from these new opportunities, further development of instrumentation and detectors is necessary, along with numerical algorithms for phase retrieval, noise reduction and ultimate resolution.

Acknowledgements

The authors are grateful to Benoit Maillot and Petra Pernot for preparation and help with the imaging of *Deinococus radiodurans*, to Giuseppe Faraci for provision of the porous silicon sample, to the ID10 beamline staff, and to the ESRF for beamtime.

Funding information

The Research Council of Norway is acknowledged for the financial funding of the ICONIC project (FRINATEK project No. 303252).

References

- Aidukas, T., Phillips, N. W., Diaz, A., Poghosyan, E., Müller, E., Levi, A. F. J., Aeppli, G., Guizar-Sicairos, M. & Holler, M. (2024). *Nature*, **632**, 81–88.
- Ayyer, K., Lan, T.-Y., Elser, V. & Loh, N. D. (2016). J. Appl. Cryst. 49, 1320–1335.
- Ayyer, K., Xavier, P. L., Bielecki, J., Shen, Z., Daurer, B. J., Samanta,
 A. K., Awel, S., Bean, R., Barty, A., Bergemann, M., Ekeberg, T.,
 Estillore, A. D., Fangohr, H., Giewekemeyer, K., Hunter, M. S.,
 Karnevskiy, M., Kirian, R. A., Kirkwood, H., Kim, Y., Koliyadu, J.,
 Lange, H., Letrun, R., Lübke, J., Michelat, T., Morgan, A. J., Roth,
 N., Sato, T., Sikorski, M., Schulz, F., Spence, J. C. H., Vagovic, P.,
 Wollweber, T., Worbs, L., Yefanov, O., Zhuang, Y., Maia, F. R. N. C.,
 Horke, D. A., Küpper, J., Loh, N. D., Mancuso, A. P. & Chapman,
 H. N. (2021). Optica, 8, 15–23.
- Bajt, S., Prasciolu, M., Fleckenstein, H., Domaracký, M., Chapman, H. N., Morgan, A. J., Yefanov, O., Messerschmidt, M., Du, Y.,

- Murray, K. T., Mariani, V., Kuhn, M., Aplin, S., Pande, K., Villanueva-Perez, P., Stachnik, K., Chen, J. P., Andrejczuk, A., Meents, A., Burkhardt, A., Pennicard, D., Huang, X., Yan, H., Nazaretski, E., Chu, Y. S. & Hamm, C. E. (2018). *Light Sci. Appl.* **7**, 17162– 17162.
- Bartels, M., Priebe, M., Wilke, R. N., Krüger, S. P., Giewekemeyer, K., Kalbfleisch, S., Olendrowitz, C., Sprung, M. & Salditt, T. (2012). *Opt. Nanoscopy*, **1**, 10.
- Baruchel, J., Bleuet, P., Bravin, A., Coan, P., Lima, E., Madsen, A., Ludwig, W., Pernot, P. & Susini, J. (2008). C. R. Phys. 9, 624–641.
- Bellisario, A., Maia, F. R. N. C. & Ekeberg, T. (2022). J. Appl. Cryst. 55, 122–132.
- Bérujon, S., Ziegler, E., Cerbino, R. & Peverini, L. (2012). Phys. Rev. Lett. 108, 158102.
- Beuvier, T., Chushkin, Y., Zontone, F., Gibaud, A., Cherkas, O., Da Silva, J. & Snigireva, I. (2022). *IUCrJ*, **9**, 580–593.
- Beuvier, T., Probert, I., Beaufort, L., Suchéras-Marx, B., Chushkin, Y., Zontone, F. & Gibaud, A. (2019). *Nat. Commun.* **10**, 751.
- Biasci, J. C., Bouteille, J. F., Carmignani, N., Chavanne, J., Coulon, D., Dabin, Y., Ewald, F., Farvacque, L., Goirand, L., Hahn, M., Jacob, J., LeBec, G., Liuzzo, S., Nash, B., Pedroso-Marques, H., Perron, T., Plouviez, E., Raimondi, P., Revol, J. L., Scheidt, K. & Serrière, V. (2014). Synchrotron Radiation News, 27(6), 8–12.
- Björling, A., Marçal, L. A. B., Solla-Gullón, J., Wallentin, J., Carbone, D. & Maia, F. R. N. C. (2020). *Phys. Rev. Lett.* **125**, 246101.
- Carnis, J., Gao, L., Labat, S., Kim, Y. Y., Hofmann, J. P., Leake, S. J., Schülli, T. U., Hensen, E. J. M., Thomas, O. & Richard, M.-I. (2019). *Sci. Rep.* **9**, 17357.
- Chang, H., Lou, Y., Ng, M. K. & Zeng, T. (2016). SIAM J. Sci. Comput. 38, A3672–A3695.
- Chao, W., Harteneck, B. D., Liddle, J. A., Anderson, E. H. & Attwood, D. T. (2005). *Nature*, **435**, 1210–1213.
- Chapman, H. N., Barty, A., Marchesini, S., Noy, A., Hau-Riege, S. P., Cui, C., Howells, M. R., Rosen, R., He, H., Spence, J. C. H., Weierstall, U., Beetz, T., Jacobsen, C. & Shapiro, D. (2006). *J. Opt. Soc. Am. A*, 23, 1179–1200.
- Chapman, H. N. & Nugent, K. A. (2010). Nat. Photon. 4, 833-839.
- Chattopadhyay, B., Madathiparambil, A. S., Mürer, F. K., Cerasi, P., Chushkin, Y., Zontone, F., Gibaud, A. & Breiby, D. W. (2020). *J. Appl. Cryst.* **53**, 1562–1569.
- Chen, C.-C., Miao, J., Wang, C. W. & Lee, T. K. (2007). *Phys. Rev. B*, **76**, 064113.
- Cherkas, O., Beuvier, T., Breiby, D. W., Chushkin, Y., Zontone, F. & Gibaud, A. (2017). Cryst. Growth Des. 17, 4183–4188.
- Cherkas, O., Beuvier, T., Zontone, F., Chushkin, Y., Demoulin, L., Rousseau, A. & Gibaud, A. (2018). *Adv. Powder Technol.* **29**, 2872–2880.
- Chushkin, Y., Zontone, F., Cherkas, O. & Gibaud, A. (2019). J. Appl. Cryst. 52, 571–578.
- Chushkin, Y., Zontone, F., Lima, E., De Caro, L., Guardia, P., Manna, L. & Giannini, C. (2014). J. Synchrotron Rad. 21, 594–599.
- Cloetens, P., Ludwig, W., Baruchel, J., Van Dyck, D., Van Landuyt, J., Guigay, J. P. & Schlenker, M. (1999). *Appl. Phys. Lett.* **75**, 2912– 2914.
- De Andrade, V., Nikitin, V., Wojcik, M., Deriy, A., Bean, S., Shu, D., Mooney, T., Peterson, K., Kc, P., Li, K., Ali, S., Fezzaa, K., Gürsoy, D., Arico, C., Ouendi, S., Troadec, D., Simon, P., De Carlo, F. & Lethien, C. (2021). Adv. Mater. 33, 2008653.
- Dierolf, M., Menzel, A., Thibault, P., Schneider, P., Kewish, C. M., Wepf, R., Bunk, O. & Pfeiffer, F. (2010). *Nature*, 467, 436–439.
- Donnelly, C., Guizar-Sicairos, M., Scagnoli, V., Gliga, S., Holler, M., Raabe, J. & Heyderman, L. J. (2017). *Nature*, 547, 328–331.
- Donnelly, C. & Scagnoli, V. (2020). J. Phys. Condens. Matter, 32, 213001.
- Donnelly, C., Scagnoli, V., Guizar-Sicairos, M., Holler, M., Wilhelm, F., Guillou, F., Rogalev, A., Detlefs, C., Menzel, A., Raabe, J. & Heyderman, L. J. (2016). *Phys. Rev. B*, **94**, 064421.

- Einfeld, D., Schaper, J. & Plesko, M. (1995). Proceedings Particle Accelerator Conference, Dallas, TX, USA, Vol. 1, pp. 177–179.
- Eisebitt, S., Lüning, J., Schlotter, W. F., Lörgen, M., Hellwig, O., Eberhardt, W. & Stöhr, J. (2004). *Nature*, **432**, 885–888.
- ESRF (2014). ESRF Upgrade Programme Phase II (2015–2022), Technical Design Study. http://www.esrf.fr/Apache_files/Upgrade/ ESRF-orange-book.pdf.
- Favre-Nicolin, V., Girard, G., Leake, S., Carnis, J., Chushkin, Y., Kieffer, J., Paleo, P. & Richard, M.-I. (2020a). J. Appl. Cryst. 53, 1404–1413.
- Favre-Nicolin, V., Leake, S. & Chushkin, Y. (2020b). Sci. Rep. 10, 2664.
- Fienup, J. R. (1987). J. Opt. Soc. Am. A, 4, 118–123.
- Flewett, S., Schaffert, S., Mohanty, J., Guehrs, E., Geilhufe, J., Günther, C. M., Pfau, B. & Eisebitt, S. (2012). *Phys. Rev. Lett.* 108, 223902.
- Frank, V., Chushkin, Y., Fröhlich, B., Abuillan, W., Rieger, H., Becker, A. S., Yamamoto, A., Rossetti, F. F., Kaufmann, S., Lanzer, M., Zontone, F. & Tanaka, M. (2017). *Sci. Rep.* 7, 14081.
- Gao, Y., Harder, R., Southworth, S. H., Guest, J. R., Huang, X., Yan,
 Z., Ocola, L. E., Yifat, Y., Sule, N., Ho, P. J., Pelton, M., Scherer,
 N. F. & Young, L. (2019). *Proc. Natl Acad. Sci. USA*, **116**, 4018–4024.
- Gerchberg, R. W. & Saxton, W. O. (1972). Optik (Stuttgart), 35, 237–246.
- Giewekemeyer, K., Aquila, A., Loh, N.-T. D., Chushkin, Y., Shanks, K. S., Weiss, J. T., Tate, M. W., Philipp, H. T., Stern, S., Vagovic, P., Mehrjoo, M., Teo, C., Barthelmess, M., Zontone, F., Chang, C., Tiberio, R. C., Sakdinawat, A., Williams, G. J., Gruner, S. M. & Mancuso, A. P. (2019). *IUCrJ*, 6, 357–365.
- Grimes, M., Pauwels, K., Schülli, T. U., Martin, T., Fajardo, P., Douissard, P.-A., Kocsis, M., Nishino, H., Ozaki, K., Honjo, Y., Nishiyama Hiraki, T., Joti, Y., Hatsui, T., Levi, M., Rabkin, E., Leake, S. J. & Richard, M.-I. (2023). J. Appl. Cryst. 56, 1032–1037.
- Grote, L., Seyrich, M., Döhrmann, R., Harouna-Mayer, S. Y., Mancini, F., Kaziukenas, E., Fernandez-Cuesta, I. A., Zito, C., Vasylieva, O., Wittwer, F., Odstrčzil, M., Mogos, N., Landmann, M., Schroer, C. G. & Koziej, D. (2022). *Nat. Commun.* **13**, 4971.
- Harder, R. (2021). IUCrJ, 8, 1-3.
- Hinsley, G. N., Westermeier, F., Wang, B., Ngoi, K. H., Singh, S., Rysov, R., Sprung, M., Kewish, C. M., van Riessen, G. A. & Vartanyants, I. A. (2024). *Nano Lett.* 24, 13702–13707.
- Holler, M., Guizar-Sicairos, M., Tsai, E. H. R., Dinapoli, R., Müller, E., Bunk, O., Raabe, J. & Aeppli, G. (2017). *Nature*, 543, 402–406.
- Howells, M., Beetz, T., Chapman, H., Cui, C., Holton, J., Jacobsen, C., Kirz, J., Lima, E., Marchesini, S., Miao, H., Sayre, D., Shapiro, D., Spence, J. & Starodub, D. (2009). J. Electron Spectrosc. Relat. Phenom. 170, 4–12.
- Jacobsen, C. (2019). X-ray Microscopy. Cambridge University Press.
- Jiang, H., Ramunno-Johnson, D., Song, C., Amirbekian, B., Kohmura, Y., Nishino, Y., Takahashi, Y., Ishikawa, T. & Miao, J. (2008). *Phys. Rev. Lett.* **100**, 038103.
- Jiang, H., Song, C., Chen, C.-C., Xu, R., Raines, K. S., Fahimian, B. P., Lu, C.-H., Lee, T.-K., Nakashima, A., Urano, J., Ishikawa, T., Tamanoi, F. & Miao, J. (2010). *Proc. Natl Acad. Sci. USA*, **107**, 11234–11239.
- Johansson, U., Carbone, D., Kalbfleisch, S., Björling, A., Kahnt, M., Sala, S., Stankevic, T., Liebi, M., Rodriguez Fernandez, A., Bring, B., Paterson, D., Thånell, K., Bell, P., Erb, D., Weninger, C., Matej, Z., Roslund, L., Åhnberg, K., Norsk Jensen, B., Tarawneh, H., Mikkelsen, A. & Vogt, U. (2021). J. Synchrotron Rad. 28, 1935– 1947.
- Jong, E. M. L. D. de, Mannino, G., Alberti, A., Ruggeri, R., Italia, M., Zontone, F., Chushkin, Y., Pennisi, A. R., Gregorkiewicz, T. & Faraci, G. (2016). Sci. Rep. 6, 25664.
- Kalbfleisch, S., Zhang, Y., Kahnt, M., Buakor, K., Langer, M., Dreier, T., Dierks, H., Stjärneblad, P., Larsson, E., Gordeyeva, K.,

Chayanun, L., Söderberg, D., Wallentin, J., Bech, M. & Villanueva-Perez, P. (2022). J. Synchrotron Rad. 29, 224–229.

- Kolb, T., Albert, S., Haug, M. & Whyte, G. (2015). J. Biophotonics, 8, 239–246.
- Latychevskaia, T., Chushkin, Y., Zontone, F. & Fink, H.-W. (2015). *Appl. Phys. Lett.* **107**, 183102.
- Lima, E., Wiegart, L., Pernot, P., Howells, M., Timmins, J., Zontone, F. & Madsen, A. (2009). *Phys. Rev. Lett.* **103**, 198102.
- Lo, Y. H., Zhao, L., Gallagher-Jones, M., Rana, A. J., Lodico, J., Xiao, W., Regan, B. C. & Miao, J. (2018). *Nat. Commun.* 9, 1826.
- Loh, N. D. & Elser, V. (2009). Phys. Rev. E, 80, 026705.
- Luke, D. R. (2005). Inverse Probl. 21, 37-50.
- Maiden, A. M., Humphry, M. J. & Rodenburg, J. M. (2012). J. Opt. Soc. Am. A, 29, 1606–1614.
- Malm, E. (2021). PhD thesis, Lund University, Sweden.
- Malm, E. & Chushkin, Y. (2025). J. Synchrotron Rad. 32, 210-216.
- Marchesini, S., He, H., Chapman, H. N., Hau-Riege, S. P., Noy, A., Howells, M. R., Weierstall, U. & Spence, J. C. H. (2003). *Phys. Rev. B*, 68, 140101.
- Masto, M., Favre-Nicolin, V., Leake, S., Schülli, T., Richard, M.-I. & Bellec, E. (2024). J. Appl. Cryst. 57, 966–974.
- McIntosh, J. R. (2007). *Cellular Electron Microscopy*, edited by J. R. McIntosh, pp. xxi-xxvii. Academic Press.
- McNulty, I., Kirz, J., Jacobsen, C., Anderson, E. H., Howells, M. R. & Kern, D. P. (1992). *Science*, **256**, 1009–1012.
- Miao, J., Charalambous, P., Kirz, J. & Sayre, D. (1999). Nature, 400, 342–344.
- Miao, J., Ishikawa, T., Anderson, E. H. & Hodgson, K. O. (2003). *Phys. Rev. B*, **67**, 174104.
- Miao, J., Nishino, Y., Kohmura, Y., Johnson, B., Song, C., Risbud, S. H. & Ishikawa, T. (2005). *Phys. Rev. Lett.* **95**, 085503.
- Miao, J., Sayre, D. & Chapman, H. N. (1998). J. Opt. Soc. Am. A, 15, 1662–1669.
- Mokso, R., Cloetens, P., Maire, E., Ludwig, W. & Buffière, J.-Y. (2007). *Appl. Phys. Lett.* **90**, 144104.
- Müller, P., Schürmann, M., Chan, C. J. & Guck, J. (2015). *Proc. SPIE*, **9548**, 95480U.
- Nakano, M., Miyashita, O., Joti, Y., Suzuki, A., Mitomo, H., Niida, Y., Yang, Y., Yumoto, H., Koyama, T., Tono, K., Ohashi, H., Yabashi, M., Ishikawa, T., Bessho, Y., Ijiro, K., Nishino, Y. & Tama, F. (2022). *Optica*, **9**, 776–784.
- Neutze, R., Wouts, R., van der Spoel, D., Weckert, E. & Hajdu, J. (2000). *Nature*, **406**, 752–757.
- Nishino, Y., Eltsov, M., Joti, Y., Ito, K., Takata, H., Takahashi, Y., Hihara, S., Frangakis, A. S., Imamoto, N., Ishikawa, T. & Maeshima, K. (2012). *EMBO J.* **31**, 1644–1653.
- Pfeifer, M. A., Williams, G. J., Vartanyants, I. A., Harder, R. & Robinson, I. K. (2006). *Nature*, 442, 63–66.
- Pfeiffer, F. (2018). Nat. Photon. 12, 9–17.
- Pham, M., Yin, P., Rana, A., Osher, S. & Miao, J. (2019). *Opt. Express*, **27**, 2792–2808.
- Raimondi, P., Benabderrahmane, C., Berkvens, P., Biasci, J. C., Borowiec, P., Bouteille, J.-F., Brochard, T., Brookes, N. B., Carmignani, N., Carver, L. R., Chaize, J.-M., Chavanne, J., Checchia, S., Chushkin, Y., Cianciosi, F., Di Michiel, M., Dimper, R., D'Elia, A., Einfeld, D., Ewald, F., Farvacque, L., Goirand, L., Hardy, L., Jacob, J., Jolly, L., Krisch, M., Le Bec, G., Leconte, I., Liuzzo, S. M., Maccarrone, C., Marchial, T., Martin, D., Mezouar, M., Nevo, C., Perron, T., Plouviez, E., Reichert, H., Renaud, P., Revol, J.-L., Roche, B., Scheidt, K.-B., Serriere, V., Sette, F., Susini, J., Torino, L., Versteegen, R., White, S. & Zontone, F. (2023). *Commun. Phys.* 6, 82.
- Robinson, I. & Harder, R. (2009). Nat. Mater. 8, 291-298.
- Robinson, I. K., Vartanyants, I. A., Williams, G. J., Pfeifer, M. A. & Pitney, J. A. (2001). *Phys. Rev. Lett.* **87**, 195505.

- Rodenburg, J. M., Hurst, A. C., Cullis, A. G., Dobson, B. R., Pfeiffer, F., Bunk, O., David, C., Jefimovs, K. & Johnson, I. (2007). *Phys. Rev. Lett.* 98, 034801.
- Rodriguez, J. A., Xu, R., Chen, C.-C., Huang, Z., Jiang, H., Chen, A. L., Raines, K. S., Pryor, A. Jr, Nam, D., Wiegart, L., Song, C., Madsen, A., Chushkin, Y., Zontone, F., Bradley, P. J. & Miao, J. (2015). *IUCrJ*, 2, 575–583.
- Rose, M., Bobkov, S., Ayyer, K., Kurta, R. P., Dzhigaev, D., Kim, Y. Y., Morgan, A. J., Yoon, C. H., Westphal, D., Bielecki, J., Sellberg, J. A., Williams, G., Maia, F. R. N. C., Yefanov, O. M., Ilyin, V., Mancuso, A. P., Chapman, H. N., Hogue, B. G., Aquila, A., Barty, A. & Vartanyants, I. A. (2018). *IUCrJ*, 5, 727–736.
- Sanzaro, S., Zontone, F., Grosso, D., Bottein, T., Neri, F., Smecca, E., Mannino, G., Bongiorno, C., Spinella, C., La Magna, A. & Alberti, A. (2019). *Nanomaterials*, 9, 1300.
- Sayre, D. (1952). Acta Cryst. 5, 843-843.
- Schropp, A., Hoppe, R., Patommel, J., Samberg, D., Seiboth, F., Stephan, S., Wellenreuther, G., Falkenberg, G. & Schroer, C. G. (2012). Appl. Phys. Lett. 100, 253112.
- Shang, M., Liao, M., Li, Y., Lu, D., Deng, D., Zhang, C., Chen, H. & Lu, H. (2025). Opt. Laser Technol. 181, 111726.
- Skjønsfjell, E. T. B., Chushkin, Y., Zontone, F. & Breiby, D. W. (2018a). J. Synchrotron Rad. 25, 1162–1171.
- Skjønsfjell, E. T. B., Kleiven, D., Patil, N., Chushkin, Y., Zontone, F., Gibaud, A. & Breiby, D. W. (2018b). J. Opt. Soc. Am. A, 35, A7– A17.
- Soltau, J., Vassholz, M., Osterhoff, M. & Salditt, T. (2021). *Optica*, **8**, 818–823.
- Song, C., Ramunno-Johnson, D., Nishino, Y., Kohmura, Y., Ishikawa, T., Chen, C.-C., Lee, T.-K. & Miao, J. (2007). *Phys. Rev. B*, **75**, 012102.
- Stockmar, M., Cloetens, P., Zanette, I., Enders, B., Dierolf, M., Pfeiffer, F. & Thibault, P. (2013). *Sci. Rep.* **3**, 1927.
- Stroke, G. & Falconer, D. (1964). Phys. Lett. 13, 306–309.
- Sun, Y. & Singer, A. (2024). Chem. Phys. Rev. 5, 031310.
- Takahashi, Y., Abe, M., Uematsu, H., Takazawa, S., Sasaki, Y., Ishiguro, N., Ozaki, K., Honjo, Y., Nishino, H., Kobayashi, K., Hiraki, T. N., Joti, Y. & Hatsui, T. (2023). J. Synchrotron Rad. 30, 989–994.
- Takahashi, Y., Nishino, Y., Tsutsumi, R., Zettsu, N., Matsubara, E., Yamauchi, K. & Ishikawa, T. (2010). *Phys. Rev. B*, 82, 214102.
- Takazawa, S., Dao, D.-A., Abe, M., Uematsu, H., Ishiguro, N., Hoshino, T., Dam, H. C. & Takahashi, Y. (2023). *Phys. Rev. Res.* 5, L042019.
- Thibault, P., Dierolf, M., Menzel, A., Bunk, O., David, C. & Pfeiffer, F. (2008). *Science*, **321**, 379–382.
- Thibault, P., Elser, V., Jacobsen, C., Shapiro, D. & Sayre, D. (2006). *Acta Cryst.* A**62**, 248–261.
- Tsai, E. H. R., Usov, I., Diaz, A., Menzel, A. & Guizar-Sicairos, M. (2016). *Opt. Express*, **24**, 29089–29108.
- Ukleev, V., Yamasaki, Y., Morikawa, D., Kanazawa, N., Okamura, Y., Nakao, H., Tokura, Y. & Arima, T. (2018). *QuBS*, **2**, 3.
- Verezhak, M. (2016). PhD thesis, Université Grenoble Alpes, France.
- Vila-Comamala, J., Diaz, A., Guizar-Sicairos, M., Mantion, A., Kewish, C. M., Menzel, A., Bunk, O. & David, C. (2011). *Opt. Express*, **19**, 21333–21344.
- Wilke, R. N., Vassholz, M. & Salditt, T. (2013). Acta Cryst. A69, 490– 497.
- Wu, L., Juhas, P., Yoo, S. & Robinson, I. (2021). IUCrJ, 8, 12-21.
- Yokoyama, Y., Yamasaki, Y., Okada, M. & Mizumaki, M. (2022). J. Phys. Soc. Jpn, 91, 034701.
- Yoreo, J. J. De, Gilbert, P. U. P. A., Sommerdijk, N. A. J. M., Penn, R. L., Whitelam, S., Joester, D., Zhang, H., Rimer, J. D., Navrotsky, A., Banfield, J. F., Wallace, A. F., Michel, F. M., Meldrum, F. C., Cölfen, H. & Dove, P. M. (2015). *Science*, **349**, aaa6760.