

DISCO synchrotron-radiation circular-dichroism endstation at SOLEIL

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The new synchrotron-radiation circular-dichroism (SRCD) endstation on the UV-visible synchrotron beamline DISCO has been commissioned at the SOLEIL synchrotron. The design has been focused on preservation of a high degree of linear polarization at high flux and moderate resolving power covering the vacuum ultraviolet to visible spectral range (125–600 nm). The beam dimensions have been set to 4 mm × 4 mm at 1 nm bandwidth for lower sample degradation. The nitrogen-purged sample chamber fits three types of sample holders accommodating conventional round cell mounting, automated rotation of the samples, as well as a microfluidic set-up. Automated temperature-controlled data collection on microvolumes is now available to the biology and chemistry communities. Macromolecules including membrane proteins, soluble proteins, bio-nanotubes, sugars, DNA and RNAs are now routinely investigated.

Keywords: spectroscopy; SRCD; far-UV; linear dichroism; circular dichroism; high throughput.

1. Introduction

In the past ten years synchrotron-radiation circular dichroism (SRCD), a sensitive spectroscopic probe of biological and chemical macromolecules, has opened up new fields in structural biology (Matsuo & Gekko, 2004; Hoffmann *et al.*, 2007; Bürck *et al.*, 2008; Cowieson *et al.*, 2008; Meinert *et al.*, 2012). The chiral properties of biomolecules such as sugars, proteins and nucleotides are now commonly investigated on several international synchrotron light sources by comparing differences in the absorption of right and left circularly polarized light (Liu *et al.*, 2010; Tao *et al.*, 2007; Jávorfi *et al.*, 2010; Matsuo *et al.*, 2005; Clarke & Jones, 2004). The structural information content of circular dichroism (CD) spectra is significantly enhanced by high-intensity sources and by an extended spectral range which spans from the visible to the vacuum ultraviolet. Together, these advantages access more electronic transitions and reduce sample volumes of precious biological macromolecules within absorbing buffers (Bulheller *et al.*, 2008; Wallace, 2010).

The wide spectral range (600 nm to 125 nm) now accessible on the SRCD beamline at DISCO is not yet available in its integrity with conventional light sources. SRCD not only allows rapid screening of protein folding behaviour in various buffer environments including salts, lipids and detergents, but also provides access to spectral information of poorly folded proteins with low CD amplitudes such as observed for amyloid and fibril forming proteins (Bousset *et al.*, 2010; Gohon *et al.*, 2011). This is important for structural biologists who have limited access to the folding properties of aggregating

proteins with high-resolution techniques such as NMR, X-ray crystallography or electron microscopy (Miron *et al.*, 2005). Indeed, SRCD, like small-angle X-ray scattering and Fourier transform infrared spectroscopy, is becoming a promising contributor in integrated biology (Stanley *et al.*, 2004).

Designs and optical properties of the SRCD experiments vary from synchrotron to synchrotron, therefore users are provided with a diversity of experimental set-ups. It is in the interest of the growing SRCD community that CD spectra are viable and reproducible at the various stations as well as comparable with conventional CD spectra (Wallace *et al.*, 2010). A previous international study concerning conventional CD spectrophotometers has highlighted the 'improved comparability and data quality' obtained in different laboratories (Ravi *et al.*, 2010).

2. Beamline overview

An outline of the beamline, including the light extraction at the front-end with its 1.7 T dipole magnet (bending magnet), the optical design and the concept of the three experimental stations, has been described previously (Giuliani *et al.*, 2009).

Here we focus on the SRCD endstation, with a horizontal and vertical acceptance of 50 mrad × 10 mrad. The collimated synchrotron radiation beam is focused and propagated (Fig. 1) through four grazing incident mirrors (silicon) into the monochromator (Cinel Scientific Instruments, Italy) and refocused onto the sample with

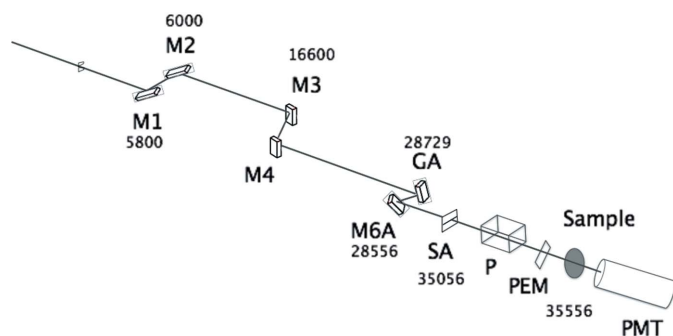


Figure 1
Schematic representation of the part of the beamline of interest for the SRCD endstation. The beam is focused by M2 just before passing through the radioprotection wall. A plane variable-line-spacing grating (GA) monochromatizes the beam before refocusing by a toroidal mirror (M6A). To ensure pure polarization, it passes through a Rochon-type polarizer (P) before being changed to circular light *via* a photoelastic modulator (PEM), and the detection is provided by locking the high tension (HT) of a photomultiplier tube (PMT). Numbers refer to the distance to the source point in millimetres.

mirror M6A. The beam divergence after the monochromator is 10 mrad (horizontally) and 9 mrad (vertically), reducing the original divergence.

Preservation of the inherent synchrotron beam polarization is an important constraint in the beamline design. The reflectivity of the surfaces as well as the number of reflective planes and their angles have been taken into account, balancing the reflectivities of the s-polarized and the p-polarized radiation through the six optical elements, which include the focusing mirrors and the grating (Table 1).

The ultra-high-vacuum monochromator design includes a normal-incident variable-line-spaced grating ($400 \text{ lines mm}^{-1}$) (HORIBA, Longjumeau, France) of moderate resolving power and a toroidal normal-incident mirror focusing vertically on the exit slits defining the bandwidth and horizontally onto the sample behind the exit window (CaF_2).

In order to maximize the circular polarized light intensities, the monochromatic beam is linearly polarized using a MgF_2 Rochon-type polarizer (Bernhard Halle Nachfolger GmbH, Berlin, Germany) and modulated with a PEM90 photoelastic modulator (HINDS, USA). A diaphragm shields the sample from the scattered radiation minimizing contaminating light of opposite polarization.

Compared with reported monochromator designs (Haass *et al.*, 1994), this design is distinct in two ways: (i) the entrance slit is defined by a diaphragm close to the bending-magnet exit crotch; (ii) the focusing mirror M6A is set right after the grating focusing vertically on the sample. The wavelength range covered by the monochromator extends from 600 nm down to 125 nm and is selected through rotation of the incident angle. Possible interference, such as stray light or zero- and higher-orders reflections along the tubing, have been eliminated by the shape of the monochromator housing that includes a light baffle. In the UV range, using inorganic KCl filters, stray light was found to be less than 2.5%. In addition, in the full accessible range (visible to UV), the recorded vitamin B spectra do not show any interference.

Within the monochromator the variable-line-spaced (VLS) grating and the focusing mirror have been set at normal-incident angles and have been coated with ruthenium. Oxidation of the Al surface would degrade the s *versus* p polarization, while ruthenium does preserve both polarizations. This choice was calculated to optimize for a maximum degree of linear polarization of the synchrotron radiation and to preserve flux ($2 \times 10^{10} \text{ photons s}^{-1}$) at a moderate resolving

Table 1
Beamline details.

Beamline name	DISCO SRCD endstation
Source type	Bending magnet, 50 (H) mrad \times 10 (V) mrad
Mirrors	Silicon, covered with ruthenium
Monochromator	Normal-incidence VLS grating monochromator (Cinel)
Wavelength range	122–600 nm
Beam size on sample	4 mm \times 4 mm
Flux at 400 mA	$2 \times 10^{10} \text{ photons s}^{-1}$ at 180 nm
Sample mounting	Three possibilities (see text)
Detector type	Photomultiplier tube
Detector models	Perkin-Elmer C921 (\varnothing 10.5 mm), C1321 (\varnothing 15.5 mm) or Electron Tube 9125B (\varnothing 29 mm)
Standard acquisition parameters	Scan speed: 35 nm min^{-1} ; 300 ms time constant/1.2 s integration

power ($R = \lambda/\Delta\lambda \simeq 300\text{--}120$). The slits opening may be adapted automatically or kept fixed for bandwidth choice.

The focusing mirror (M6A) focuses on the sample 0.5 m distant from the monochromator slits to a final beam size of 4 mm \times 4 mm (Fig. 1). The beam size was measured using fluorescence from graph paper at 270 nm.

Care has been taken in the design of the exit window. A high-purity CaF_2 window (Hellma GmbH, Müllheim, Germany) was thermal-bonded into an ultra-high-vacuum flange minimizing distortion due to pressure differences between the vacuum and the nitrogen-purged atmospheric pressurized sample chamber. The cut-off of 122 nm for CaF_2 at 10% transmission at 300 K (Hunter & Malo, 1969) corresponds to the measured nitrogen absorption spectra obtained with an empty chamber (see Fig. 2a); nitrogen peaks are used for absolute calibration of the monochromator (Okabe, 1978); however, for everyday calibration of the spectrometer, a transfer standard, namely the peak positions and ratio of D-10-camphorsulfonic acid (CSA), is used (Fig. 2b). This allows the complete ratio of polarization to be determined (Kelly *et al.*, 2005). Ultra-high vacuum is required in all optical chambers in order to preserve the surfaces from carbon contamination. The vacuum in the front-end and optical chambers is kept in the low 10^{-9} mbar range under beam exposition (Giuliani *et al.*, 2009).

2.1. Different drawers for different applications

For faster sample exchange, the sample chamber is only under strong nitrogen purge, nitrogen being sufficiently transparent for VUV spectroscopy down to 120 nm. The sample chamber volume of 2.5 L has been minimized to reduce nitrogen consumption, which runs normally at 10 L min^{-1} . The chamber holder was designed for fast sample changing and high versatility. The door is constructed over sliders with magnetic sealing for safe and quick purging after sample changing. Three sample holders were designed for specific applications (Fig. 3b):

(i) The classical sample holder [Fig. 3(b), far right] includes many ports for a range of applications, including laser activation. Using this holder, samples are deposited between two CaF_2 windows (Wien *et al.*, 2005). With this design it is possible to vary the optical pathway between 6 and 100 μm .

(ii) The rotating port [Fig. 3(b), far left] allows precise rotation of the sample between each spectrum, with an angular resolution of 0.1° . This holder is dedicated to linear dichroism measurement of films or oriented samples. It is possible to launch a sequence of measurements with a spectrum recorded at each selected angle and to measure anisotropy spectra (Meinert *et al.*, 2012).

(iii) The microfluidic holder [Fig. 3(b), center] includes a microchip in quartz designed to load the sample into position. Moving of the

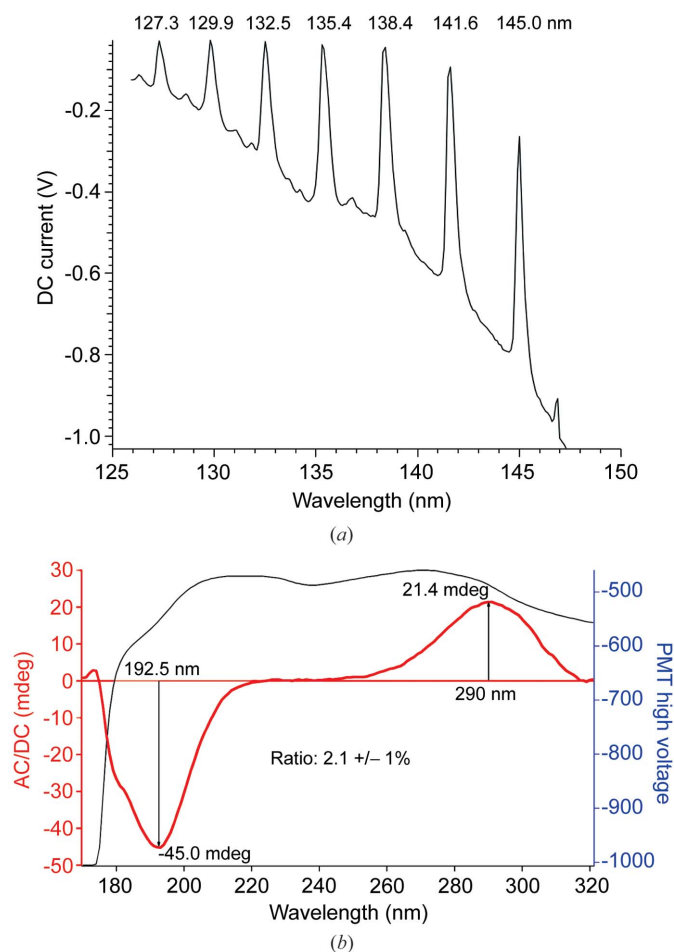


Figure 2 Calibration of the SRCD spectrometer. (a) Absolute calibration of the monochromator using N_2 peaks in the VUV. The chamber was purged with nitrogen, spectral acquisition settings were 0.1 nm steps, 300 ms time constant, HT current constant. Peak positions measured in volts of DC current correspond to the literature-referenced absorption peaks. For routine optimization of the optical alignment and electronic recording, peak positions and amplitudes ratio are verified, with a standard CSA solution 6.3 g L^{-1} in $100 \mu\text{m}$ pathlength at 298 K (b). The thick line shows the CD signal measured in mdeg; the thin line corresponds to the recorded HT measured in volts.

fluids is performed by liquid chromatography micro-pumps. It allows high-throughput experiments with high repeatability, minimizing experimental errors and a high degree of automation for screening; the necessary sample loading volume is around $10 \mu\text{L}$ for a $3 \mu\text{L}$ cell volume capacity. Pathlengths vary from $15 \mu\text{m}$ to $100 \mu\text{m}$ and the flow rate is adjustable between $10 \mu\text{L min}^{-1}$ and $50 \mu\text{L min}^{-1}$. Once the sample is in the observation chamber, the valve of the liquid chromatography pump will be automatically switched to pass-by position during a pre-defined time which allows the necessary numbers of data acquisitions. Thereafter the microfluidic cell is flushed out by a cleaning solvent such as water or Hellmanex as required for 5 min at $25 \mu\text{L min}^{-1}$. Such a microfluidic device with multiple inputs and a mixing path makes it possible to conduct automatic titration and interaction studies of proteins. A home-made program is developed in parallel in order to automate data processing.

3. Ancillary facilities overview

Directly on the beamline, users have access to biochemistry workbenches with a UV spectrometer, dryers, spins and everyday

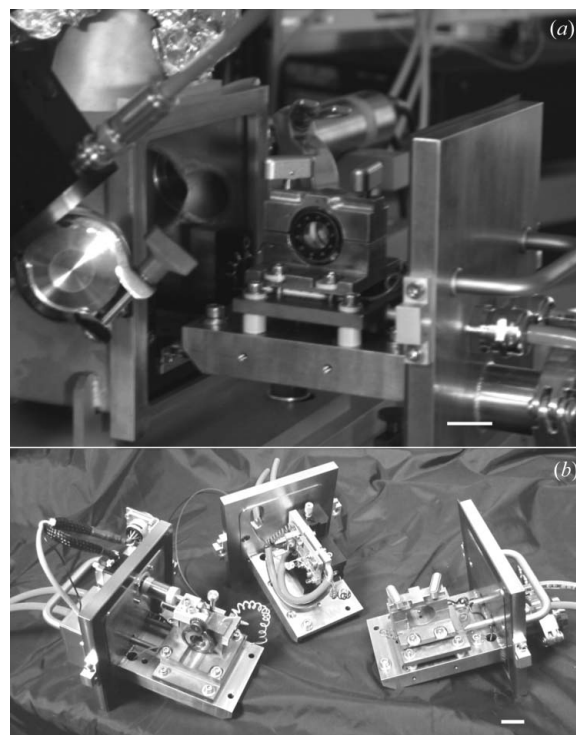


Figure 3 (a) Photograph showing the endstation with the sample chamber and an open drawer. (b) The three thermostated sample-holder drawers can easily be exchanged and repositioned; from left to right, rotating chamber, microfluidic chamber and classic chamber. Scale bars are 22 mm.

consumables. They may also obtain beam time on the two other branches for complementary mass spectrometry (Giuliani *et al.*, 2012) or deep ultraviolet (DUV) fluorescence (Jamme *et al.*, 2010). For deeper biological characterizations, a structural biology laboratory open for users 24/7 is situated only 100 m away from the endstation; it is fully equipped for protein purification and characterization.

4. Facility access

SOLEIL beamlines are under a general user program, accessible through SUNset (<http://sunset.synchrotron-soleil.fr/sun>). Access to SOLEIL beamlines is open to scientist from around the world. There are two calls for proposals per year with 15 February and 15 September deadlines for submissions.

5. Highlight

5.1. SRCD classics versus high throughput

An automatic sample handling set-up allowing high-throughput experiments with high reliability and reproducibility is in place. This technique exploits a fully automated synchrotron-radiation circular-dichroism spectrometer connected to a microfluidic device and suited to high-throughput screening of different protein conditions. In order to reduce the eventual radiation damage of the samples, the beam is kept broad ($4 \text{ mm} \times 4 \text{ mm}$) at the sample position. No spectral decrease owing to radiation damage could be recorded after 50 consecutive scans. Therefore, care has to be taken for driving the sample to its recording position. Between two measurements, the system is cleaned. Peltier temperature-controlled data acquisition permits automatic denaturation studies between 273 and 383 K for

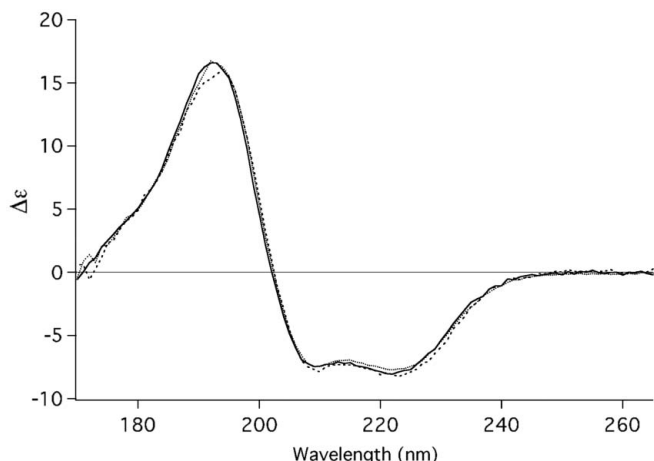


Figure 4
Comparison of myoglobin spectra: charged manually (dotted) in the round cell holders using a 10 μm pathlength at $\pm 7 \text{ mg ml}^{-1}$ in CaF_2 and in a microfluidic cell at 15 μm pathlength (dashes). The presented spectra were averaged over three raw spectra and their respective baselines subtracted, smoothed with step size of 5 (Savitzky Golay) followed by calibration to corresponding concentrations pathlengths and standardized to $\Delta\epsilon$ units using the mean residue weight of myoglobin (111.5 Da). For comparison a myoglobin spectrum from the reference data set SP175 (continuous line) is shown.

the classical and rotating sample holders in steps of 0.1 K. The range of applications includes denaturation and folding analysis, screening for better crystallization parameters and optimization, as well as ligand binding assessments. The recorded spectra are perfectly matched to ‘classical’ SRCD spectra (Fig. 4) with no loss of information.

5.2. 3 min per spectrum at very high signal to noise

The influence of the time constant (locking amplifier) and the resulting integration time (four times the time constant) have been assessed, minimizing the noise level (Fig. 5).

6. Discussion and conclusions

The SOLEIL synchrotron is a 2.75 GeV third-generation photon source. The flux is well matched to absorption biospectroscopies, therefore the main design efforts were focused on speed of acquisition and conservation of the polarization.

Temperature-controlled circular and linear dichroism as well as absorption spectroscopy in combination with automated sample rotation and continuous microfluidic cell set-ups are now commissioned and routinely used on the DISCO SRCD beamline. The performance of the beamline and especially the high throughput of samples have been the focus of the chamber design along with user-friendly software. Data acquisition, handling and data analysis are at the heart of the software design and adapted to the needs of the users. The whole set-up is controlled through *Igor* (Wavemetrics) for better data handling and analysis afterwards.

The capabilities of the DUV SRCD endstation of DISCO are presented, opening new ways to analyse biomolecules in solution. The next step will be to increase again the speed of acquisition in order to be able to follow fast kinetics in real time.

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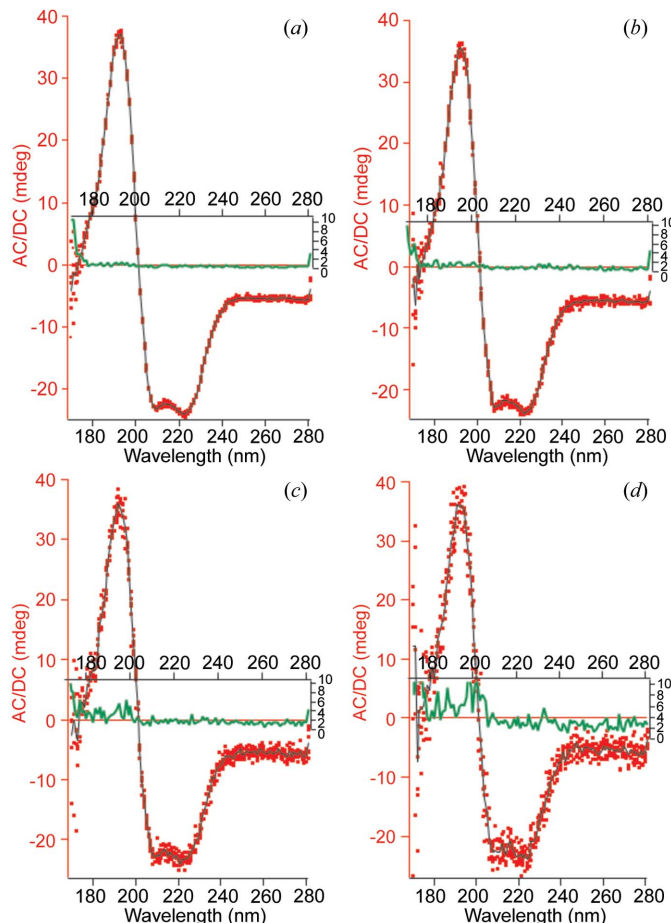


Figure 5
Influence of the speed of acquisition on the signal-to-noise ratio. Four different acquisition time constants (TC set in the lock-in amplifier) resulting in different acquisition speeds demonstrate the signal-to-noise behaviour. (a) 300 ms; (b) 100 ms; (c) 30 ms; (d) 10 ms. The acquisition time per data point corresponds to four times the time constant, also referred to as the integration time. Residuals are overlaid. Three full consecutive scans of myoglobin, 10 mg ml^{-1} in a 15 μm pathlength [raw data, black squares (red online)], have been measured for each dataset, including the return of the monochromator, and the stepping mode of the monochromator of theoretical speed 50 nm min^{-1} (corresponding to 1.2 s integration time) is reduced to a scan speed of 35 nm min^{-1} . Therefore three consecutively acquired spectra with good signal-to-noise ratio are currently produced within 9 min. The residual in percentiles [grey line (green online)] is superimposed on the zero line.

Jean-Claude Maurizot, the chaperon of the DISCO SRCD experiment.

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